DOI: https://doi.org/10.3126/asta.v1i1.30270



Research Article

Diversity and prevalence of gut parasites in urban macaques

Bikram Sapkota^{1,2}, Roshan Babu Adhikari^{3,4,6}, Ganga Ram Regmi^{4,5,7}, Bishnu Prasad Bhattarai^{1,8}, and Tirth Raj Ghimire^{3,9,*}

Received: May 20, 2020; Accepted: June 14, 2020; Published: June 25, 2020

Abstract: Rhesus macaques (*Macaca mulatta*) are commonly found to inhabit various religious sites and cities in Nepal. Similar to other nonhuman primates, they are also the natural or reservoir host of several gut parasites. However, the status of gut parasitism, particularly in the urban dweller macaques, remains largely unexplored in the country. This study aimed to assess the prevalence and diversity of gut parasites in the monkeys inhabiting Bajrabarahee, an urban temple area in Lalitpur District, Nepal. A total of 42 fresh fecal samples of macaques belonging to five different troops, were collected and preserved in 2.5% (w/v) potassium dichromate solution. The fecal samples were processed by direct wet mount, concentration, and acid-fast techniques and examined under an optical microscope. All the fecal samples were positive with gut parasites. The parasites detected were Ascarid spp., *Balantidium coli, Cryptosporidium* sp., *Eimeria* sp., *Entamoeba coli, Entamoeba* spp., *Giardia* sp., hookworm, *Strongyloides* sp., Strongylid spp., *Trichomonas* sp., and *Trichuris* sp. Cent percent prevalence rate and high species richness with 12 parasites may indicate that they have impact on the gut health of these monkeys. This suggests the need of deworming the macaque population and enhancing public awareness for pro-active control of parasitic infection as well as of adopting the preventive measures to lessen the zoonotic transmission of the pathogenic parasites.

Keywords: Cryptosporidium, Entamoeba, gastro-intestinal (GI), Macaca mulatta, zoonosis.

सारांश: रातो बाँदरहरू प्रायः नेपालका विभिन्न धार्मिकस्थल र शहरहरूमा पाइन्छन् । अन्य गैर-मानव प्राइमेटहरू जस्तै तिनीहरू पिन पेटको धेरै परजीवीहरूको प्राकृतिक आश्रयस्थलहरु हुन्, यद्यपि, खासगरी नेपालको सहरवासी रातो बाँदरको पेटमा परजीवीको स्थिति भने अभै अन्वेषण गर्नुपर्ने रहेको छ । यसैले नेपालको लिलतपुरस्थित शहरीक्षेत्रको मन्दिर बज्रबाराहीमा रहेको बाँदरको आन्द्रामा पाइने परजीवीहरूको व्यापकता र विविधताको आँकलन गर्नु यस अध्ययनको लक्ष्य हो । फरक-फरक पाँच प्रकारका समूहमा रहेका बाँदरहरुबाट कुल ४२ वटा ताजा दिसाका नमूनाहरू जम्मा गरि २.५% पोटाशियम डाइकोमेटमा संकलन गरी संरक्षित गरिएको थियो । उक्त नमूनाहरु डाइरेक्ट, कन्सनट्रेसन, र एसीड-फास्ट तरिकाहरुबाट एउटा अप्टीकल माइकोस्कोपद्वारा विश्लेषण गरिएको थियो, जसले आन्द्राहरुमा पाइने १२ वटा परजीवीहरु देखायो, जस्तै एस्कारिड, बालाण्टिडयम, किप्टोस्पोरिडियम, आइमेरिया, एन्टअमिबा कोली, एन्टअमिबा इस्पेसिज, जिआर्डिया, अंकुशे जूका, स्टूङ्गगाइलिड, स्टूङ्गगाइलोइडिस, ट्राइकोमोनस, र ट्राइचुरिस थिए । धेरै परजीवीहरुको उच्च दर र व्यप्तताले गर्दा यी बाँदरहरुको आन्द्राको स्वास्थ्यमा नकरात्मक असर पर्न सक्छ र मानवमा परजीवीहरु प्रसारणको जोखिममा उल्लेखनीय वृद्धि गर्दछ । यसले परजीवीहरुको जनोटिक प्रसारण कम गर्नका लागि रोकथामको उपायहरू अनकलन गर्न सफाव दिन्छ ।

¹Central Department of Zoology, Tribhuvan University, Kathmandu/NP.

³Animal Research Laboratory, Faculty of Science, Nepal Academy of Science and Technology (NAST), Lalitpur, Nepal.

⁴Third Pole Conservancy, Bhaktapur-10, Nepal.

⁵Institute of Forestry, Tribhuvan University, Nepal.

²E-mail: bik.saapkotaa@gmail.com.

⁶E-mail: srkroshanbabu@gmail.com, ORCID ID: 0000-0002-5876-667X

⁷E-mail: gangarregmi@gmail.com, ORCID ID: 0000-0002-8392-5863

⁸E-mail: bpbhattarai@cdztu.edu.np, ORCID ID: 0000-0001-5741-6179

⁹E-mail: tirth.ghimire@nast.gov.np, ORCID ID: 0000-0001-9952-1786

^{*} Corresponding author, E-mail: tirth.ghimire@nast.gov.np; Tel.: +977-1-5547715

[©] RECAST/TU

1. Introduction

Rhesus Macaca mulatta macaques, (Zimmermann, 1780) (Rato bander in Nepali) are well-known Old-World nonhuman primates. They are physiologically and genetically similar to humans as both are believed to share a common ancestor that diverged about 25 million years ago and developed independently (Kumar and Hedges 1998; Gibbs et al., 2007). They are listed as 'Least Concern' **IUCN** by Red List (https://www.iucnredlist.org/species/12554/3356486, accessed on May 22, 2020). Compared with other macaques M. mulatta have a high level of adaptation and are well-adapted to co-exist with the human in urban settlements (Rathoure 2014; Kumar et al., 2013).

Monkeys have an important status in mythology and religion particularly, in Hinduism and Buddhism (Jokinen 2014; Fuentes 2017; Ale et al., 2020) and are commonly found in religious sites like temples, monasteries, and many urban areas (Ale et al., 2020). There is an extensive, unregulated, and close contact of macaques with humans because the religious sites are always full of local people, worshippers, and visitors (Fuentes 2005). Their feeding ecology and habitat are more or less similar to that of humans; thus, in the nearby areas, they often invade the houses, gardens, or agricultural fields for sharing the niches. They are also known to share disease-causing pathogens like the gut or gastrointestinal (GI) parasites, for example, from many years, these mammals have been linked for the outbreaks of emerging parasitic diseases in humans (Chapman et al., 2005; Jones-Engel et al., 2006; Ghimire et al., 2020). Gut parasitism has been evidenced to result in the high morbidity and mortality in nonhuman primates including many types of macaques (Fremming et al., 1955; Remfry 1978; Toft 1986; Chapman et al., 2005) as well as in human (Stauffer and Ravdin 2003; Haque 2007; WHO 2020) around the world. Thus, it is crucial to know the status of gut parasitism in the monkeys, especially in the anthropogenic ones, to reduce the possible health consequences in macaques as well as humans. The current study was conducted to assess the prevalence and diversity of gut parasites in the monkeys inhabiting Bajrabarahee, an urban temple area with increasing human-macaque interactions in the Lalitpur district, Nepal.

2. Materials and Methods

2.1. Study area

The study had been conducted in Bajrabarahee, a religious Hindu temple area located in Godawari Municipality, Lalitpur, Nepal. The area (27.60610 N and 85.32930 E) is covered by a sacred forest and is a typical habitat of several species of birds, reptiles,

and mammals, including the *M. mulatta*. By direct counting methods, we assessed 50 monkeys in the forest. The area is surrounded by agricultural land to the south and east, and human settlement to the north and west. Besides the religious importance, it is also a bird-watching and recreational spot, and thus many religious people, tourists, and local people usually visit the site. It has also been developed as a picnic spot, so macaque-human interaction is typical in the study area.





Figure 1. *M. mulatta* in the study area. (a) Closecontact scenario. (b) Feeding on garbage.

2.2. Sample collection, preservation, and transportation

From June to August 2019, a total of 42 fresh fecal samples of *M. mulatta* belonging to five different troops were collected non-invasively from various sites in the study area (Figure 1). The fecal samples were immediately preserved in 2.5% (w/v) potassium dichromate solution in 20 mL sterile vials. Then, the samples were transported to the Animal Research Laboratory (ARL) of the Nepal Academy of Science and Technology (NAST) and stored in the refrigerator (4° Celsius) for further analysis.

2.3. Laboratory processing and examination

The fecal samples stored in 2.5% (w/v) potassium dichromate were microscopically examined by four different techniques - direct wet mount, sedimentation, saturated salt (45% w/v NaCl) flotation, and modified acid-fast techniques as previously described (Ghimire and Bhattarai 2019).

2.4. Parasite identification

All the fecal samples were observed under an optical microscope (Optika Microscopes Italy, B-383PLi) at X100, X400, and X1000 total magnifications. Parasitic images were taken by the camera (SXView 2.2.0.172 Beta (Nov 6, 2014) Copyright (C) 2013-2014). The micrometry of parasitic bodies was assessed using ImageJ 1.51k (National Institute of Health, USA) and identification was carried out as previously described (Petrášová et al., 2010; Soulsby 2012; Li et al., 2017)

2. 5. Data analysis

Data were expressed as numbers of positive samples as well as prevalence rates in the table using Microsoft Word 2007. Prevalence rates were calculated by dividing the number of parasite positive samples (total or particular species) by the total number of samples observed (Ghimire and Bhattarai 2019).

3. Results

In the current study, we reported a 100% prevalence rate of gut parasites. The prevalence of protozoa was higher (90.5%) compared to that of the helminths (47.6%). Furthermore, a total of 12 gut parasitic species were detected. They were protozoa - *Entamoeba* spp. (66.7%), *Balantidium coli* (59.5%), *Entamoeba coli* (57.1%), *Cryptosporidium* sp. (11.9%), *Eimeria* sp. (7.1%), *Giardia* sp. (4.8%), and *Trichomonas* sp. (2.4%) and helminths such as Ascarid spp. (21.4%), *Strongyloides* sp. (21.4%), hookworm (19%), *Trichuris* sp. (14.3%), and Strongylid spp. (9.5%) (**Figure 2**) (**Table 1**).

All samples were found to be mixed infections with two or more gut parasitic species. Triplet infection was the highest (57.1%) followed by the duplet (26.2%), and pentuplet (4.8%) infections were the least (**Table 1**). Further, two morphotypes of Ascarid eggs were detected. Some of these eggs were similar to human *Ascaris* (size range: 54–58x40–44μm), and others were similar to animal *Toxocara* (size range: 72–84x61–68μm). Similarly, based on the morphology and micrometry, three morphotypes of Strongylid eggs were detected (size range: 72–111x38–65μm) in the current study (**Figure 2**).

4. Discussion

The current study explores the status and diversity of the gut parasites in monkeys inhabiting an urban temple area situated in between human settlements in Nepal. In this study, the overall prevalence of the gut parasites was 100% which was similar to the result from Bangladesh (100%) (Tabasshum et al., 2018), higher than the findings

from Nepal (61.9% – 86%) (Jha et al., 2011; Adhikari and Dhakal 2018; Bhattarai et al., 2019), and India (40%–66.5%) (Parmar et al., 2012; Jaiswal et al., 2014; Kumar et al., 2018). Besides, cent percent concomitant infections with maximum triplet co-infection rate suggested the parasitic richness in the gut of the macaques.

Table 1. Gut parasitic species, their concurrency, and prevalence in *M. mulatta. N represents total samples collected and n represents number of positive sample/s.*

| Infecting Parasitic species | Overall Prevalence (nX100/N) (N=42) |
|-----------------------------|--|
| Protozoa | |
| Entamoeba spp. | 28 (66.7%) |
| Entamoeba coli | 24 (57.1%) |
| Eimeria sp. | 3 (7.1%) |
| Cryptosporidium sp. | 5 (11.9%) |
| Giardia sp. | 2 (4.8%) |
| Balantidium coli | 25 (59.5%) |
| Trichomonas sp. | 1 (2.4%) |
| Helminths | |
| Ascarid spp. | 24 (21.4%) |
| Strongylid spp. | 4 (9.5%) |
| Hookworm | 8 (19%) |
| Strongyloides sp. | 9 (21.4%) |
| Trichuris sp. | 6 (14.3%) |
| Total Protozoan infection | 38 (90.5%) |
| Total Helminth infection | 20 (47.6%) |
| Total Mixed infection | 42 (100%) |
| Duplet Infection | 11 (26.2 %) |
| Triplet Infection | 24 (57.1 %) |
| Quadruplet Infection | 5 (11.9%) |
| Pentuplet Infection | 2 (4.8%) |

The higher parasitic prevalence might be related to the applied study design, sampling techniques, laboratory-based techniques, sampling geography, including the climatic conditions and the lifestyle pattern of the existing macaque population. In the current study, we used a purposive sampling technique and collected the fresh fecal samples 2016). Also, laboratory techniques like direct wet and trophozoites of gut parasites (Zvinorova et al., fecal sample.

immediately after defecation by the macaques. The mount, concentrations (sedimentation and flotation), sampling period was warm and wet, the period and acid-fast staining techniques were used to favorable for the survival of larva, cysts, oocysts, enhance the detection of the parasites from each

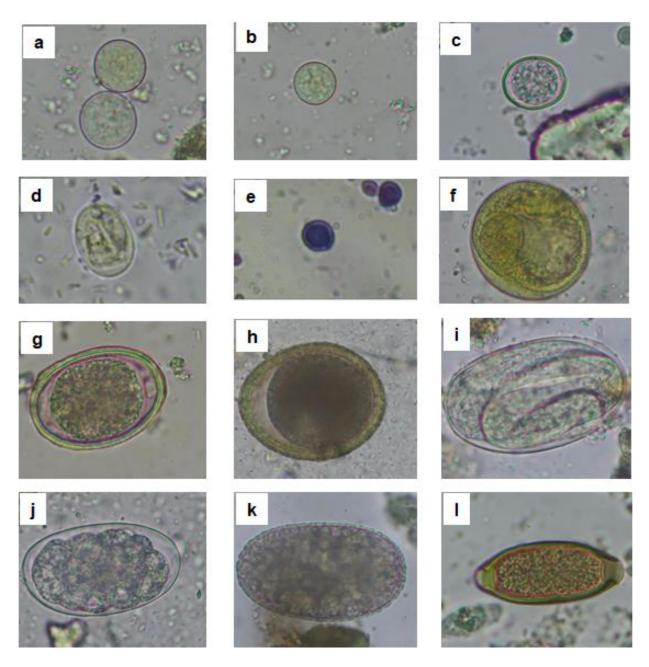


Figure. 2. Photomicrographs of various gut parasitic species. (a) Cysts of Entamoeba coli, 19µmx19µm, X400. (b) Cyst of Entamoeba sp., 13μmx13μm, X400. (c) Oocyst of Eimeria sp., 20μmx14μm, X400. (d) Cyst of Giardia sp., 14μmx9μm, X1000. (e) Oocyst of Cryptosporidium sp., 6µmx6µm, X1000. (f) Cyst of Balantidium coli, 54µmx51µm, X400. (g) Egg of Ascarid sp. 56µmx41µm, X400. (h) Egg of Ascarid (Toxocara sp.), 82µmx67µm, X400. (i) Egg of Strongyloides sp., 69μmx43μm, X400. (j) Egg of hookworm, 66μmx39μm, X400. (k) Egg of Strongylid sp., 111μmx65μm, X400. (l) Egg of Trichuris sp., 55µmx24µm, X400.

Macaques are the natural and reservoir hosts of by visitors/outsiders in the forest areas and nearby semi-nomadic life that might massively increase the parasite dispersal (Macpherson 1994; Swedell 2012). The increasing soil and water pollution by waste food and garbage, especially during the festive and

many gut parasites. They live in a group and spend a water sources, are the risk factors of parasite transmission. In this scenario, macaques are usually in contact with contaminated soil and water, and consumption of garbage foods may lead to the acquisition and transmission success of the gut picnic programs, and the occasional open defecation parasites in them, which explains their species

richness.

The current prevalence of gut protozoa was higher than that of the helminthes; however, this result contrasts with the previous findings (Adhikari and Dhakal 2018; Bhattarai et al., 2019) that recorded higher prevalence rates in helminths. Regarding protozoa, the prevalence of Entamoeba spp. in the current study was 66.7% which was lower than the findings from China (89.96%) (Zhang et al., 2019) and higher than those reported from Nepal (13.97% - 32%) (Jha et al., 2011; Pokhrel and Maharjan 2014; Adhikari and Dhakal 2018; Bhattarai et al., 2019) and from India (10% 23.07%) (Parmar et al., 2012; Jaiswal et al., 2014). Several species of these pseudopodial amoebas like Entamoeba histolytica, E nuttalli, E. dispar, E. moshkovskii, E. hartmanni, E. chattoni, and E. polecki (Tachibana et al., 2007; Jiang et al., 2008; Tachibana et al., 2013; Guan et al., 2016; Zhang et al., 2019) have already been reported from macaques all over the world; however, majority of them are considered harmless and do not exhibit pathologic illness in the macaques. Pathologically, E. histolytica and E. nuttali are critical because they induce fatal intestinal and extraintestinal amebiasis (Fremming et al. 1955; Loomis et al., 1983; Haq et al., 1985; Beaver et al., 1988; Pang et al., 1993; Verweij et al., 2003; Tachibana et al., 2007; Jiang et al., 2008; Levecke et al., 2010). In our study, Entamoeba coli showed 57.1% prevalence rate which was higher than the previous findings from Nepal (9.52% – 24.44%) (Jha et al., 2011; Bhattarai et al., 2019), India (10% - 26.92%) (Parmar et al., 2012; Jaiswal et al., 2014), and China (42%) (Guan et al., 2016). Although this species is typically asymptomatic in primates (Chapman et al., 2005), its presence should be taken as the indication of other pathogens inside the gut (Ghimire 2014).

It was notable that, the rate of prevalence of Cryptosporidium sp. was 11.9%, which was lower than the findings from Nepal (41.1%) (Bhattarai et al., 2019) and India (26.92%) (Jaiswal et al., 2014), and higher than the findings from China (10.94%) (Ye et al., 2012) and Thailand (1%) (Sricharern et al., 2016). This coccidian parasite causes highly fatal types of intestinal and extraintestinal pathologies (Kuhn et al., 1997; Kaup et al., 1998) and might be among transmitted humans and primates zoonotically (Ye et al., 2012; Zhao et al., 2019). Another coccidian parasite Eimeria sp. was reported in 7.1% of the fecal samples. This rate was lower than the findings from Nepal (16.12%) (Adhikari and Dhakal 2018) and higher than the results from India (3%) (Arunachalam et al., 2015). Eimeria sp. severe pathologic consequences, especially in the young monkeys compared to the old ones (Burrows 1972).

Giardia and Trichomonas are the two flagellated parasites reported in the current study. The prevalence rate of Giardia sp. was 4.8% which was lower than the findings from India (31%) (Debenham et al., 2017), China (8.51%) (Ye et al., 2012), Nepal (6.67%) (Bhattarai et al., 2019), Thailand (7%) (Sricharern et al., 2016), and higher than the reports from India (1.2%) (Kumar et al., 2018). Giardia causes enteritis in macaques (Toft 1986; Chapman et al., 2005) and is a zoonotically critical parasite for the public and veterinary health (Ye et al., 2012, 2014). The prevalence rate of Trichomonas sp. was 2.2% when examined in the fecal samples. Reports of this species are found in M. mulatta following histopathologic studies in USA (Blanchard and Baskin 1988) and Germany (Kuhn et al., 1997; Kondova et al., 2005). This parasite is associated with mild to moderate gastritis (Blanchard and Baskin 1988; Blanchard 1993; Kaup et al., 1998), including many other severe GI pathologic consequences in immunocompromised macaques (Kondova et al., 2005).

In the same way, *Balantidium coli*, a ciliate protozoan had a prevalence rate of 59.5% which was higher than the findings from Nepal (27.95% – 36%) (Jha et al., 2011; Pokhrel and Maharjan 2014; Adhikari and Dhakal 2018; Bhattarai et al., 2019) and India (8.7% – 19%) (Knezevich 1999; Kumar et al., 2018). This zoonotic parasite can also cause severe pathology in the intestinal tract of macaques, including diarrhea and rectal prolapse (Burrows 1972; Toft 1986; Kuhn et al., 1997).

Regarding helminths, the prevalence rate of Ascarid spp. was 21.4% which was lower than the findings from Bangladesh (90.90%) (Tabasshum et al., 2018), India (25.5% – 26.66%) (Parmar et al., 2012; Kumar et al., 2018), and Nepal (22.22%) (Bhattarai et al., 2019) and higher than the findings from Nepal (10.58% - 11.82%) (Pokhrel and Maharjan 2014; Adhikari and Dhakal 2018), India (5%) (Arunachalam et al., 2015), and Thailand (1%) (Schurer et al., 2019). The presence of *Toxocara* sp. in macaques, was also explained in various research findings from Nepal (Jha et al., 2011; Bhattarai et al., 2019) and Bangladesh (Tabasshum et al., 2018). Ascarid infection is associated with zoonosis (https://www.cdc.gov/parasites/ascariasis/index.html , Accessed on: June 2, 2020). It is one of the leading causes of high morbidity and mortality in macaques (Weiszer et al., 1968; Richards et al., 1983; Toft 1986; Chapman et al., 2005). Another nematode species Strongyloides was detected with the prevalence rate of 21.4% that was lower than the findings from England (55.6%) (Remfry 1978), Nepal (30% – 34%) (Jha et al., 2011), India (26.66%) - 57%) (Knezevich 1999; Parmar et al., 2012), and higher than the findings from Nepal (10.75%)

(Adhikari and Dhakal 2018), India (13%) (Kumar et al., 2018), and Thailand (2%) (Schurer et al., 2019). Mild to severe strongyloidiosis caused by migratory larva as well as the adult of this zoonotically significant parasite may lead to deaths, especially in young macaques (Remfry 1978; Toft 1986; Toft and Eberhard 1998; Chapman et al., 2005).

Strongylid nematode is one of the critical nematode groups representing different species. The species cannot be easily distinguished by the morphology of the eggs or without performing larval cultures and development (Ghimire and Bhattarai 2019). Its current prevalence rate 9.5% was lower than the findings from India (33%) (Arunachalam et al., 2015). Strongylids are zoonotic and can cause serious illnesses resulting in the deaths of macaques (Remfry 1978; Toft 1986; Chapman et al., 2005).

Trichuris sp. was reported with a prevalence rate of 14.3% that was in accordance with the findings from Nepal (14.05% – 14.44%) (Jha et al., 2011; Bhattarai et al., 2019) and lower than the reports from Bangladesh (50%) (Tabasshum et al., 2018), Thailand (19.6%) (Schurer et al., 2019), Nepal (23.65%) (Adhikari and Dhakal 2018), and higher than the findings from India (3.7% – 12%) (Knezevich 1999; Kumar et al., 2018) and England (11.2%) (Remfry 1978). Trichuriasis in macaques leads to intestinal disorder accompanied by rectal prolapse and may even induce death (Thienpont et al., 1962; Remfry 1978; Toft and Eberhard 1998)

The current prevalence 19% of hookworm was lower than the findings from Thailand (23%) (Schurer et al., 2019), Bangladesh (22.72%) (Tabasshum et al., 2018) and was higher than the findings from India (15.38%) (Jaiswal et al., 2014) and Nepal (6.67%) (Bhattarai et al., 2019). This zoonotic nematode may cause mild to severe clinical pathologies in the macaques, including anemia and diarrhea (Toft 1986; Toft and Eberhard 1998).

5. Conclusions

In conclusions, the Rhesus macaques are heavily infected with ameboid, flagellate, ciliate, coccidian, and nematode parasites. Besides, concomitant infections up to five different species of parasites in the fecal samples may indicate that these pathogens may pose negative impacts on the gut of macaques. Most of the parasitic species detected in this study are zoonotically significant, and thus, the macaques may enhance the risk of transmitting the parasites. Thus, practical control actions like deworming of the macaques should be conducted by the concerned authorities. There is a need of enhanced awareness among local people and visitors towards adopting hygienic precautions and decreasing the humanmacaque interaction in order to control the zoonotic transmission of the pathogens.

Acknowledgments: The authors would like to acknowledge Prof. Dr. Tej Bahadur Thapa, Head of Department, Central Department of Zoology, Tribhuvan University, Kathmandu, for permitting Bikram Sapkota for the M.Sc. Dissertation. We are grateful to Jaishree Sijapati, Chief, Faculty of Science, Nepal Academy of Science and Technology (NAST) for allowing us working in the Animal Research Laboratory. We thank the authorities of the Government of Nepal, Ministry of Forests and Environment for granting the permission of the works.

Author Contributions: BS: Conceptualization, Methodology, and Investigation. **RBA**: Investigation and original draft preparation. **GRR**: Writing-Reviewing. **BPB**: Supervision, and **TRG**: Writing-Reviewing, Resources, and Supervision.

Conflicts of Interest: The authors declare no conflict of interest.

Ethical approval

The authors declare that the study was conducted on naturally infected Rhesus monkeys. No experimental infection was established during this research work. The required permission for the collection of the fecal samples was issued by Government of Nepal, Ministry of Forests and Environment, Department of Forestry (Permission number: 202/2018).

Funder information

No specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Competing interests

None declared.

Availability of data and materials

All data generated or analyzed during the research work are included in this article

References

Adhikari PP, Dhakal P. Prevalence of gastro-intestinal parasites of Rhesus Macaque (*Macaca mulatta* Zimmermann, 1780) and Hanuman Langur (*Semnopithecus entellus* Dufresne, 1797) in Devghat, Chitwan, Nepal. Journal of Institute of Science and Technology. 2018 Apr 9;22(2):12-18. DOI: 10.3126/jist.v22i2.19590

Ale PB, Kandel K, Ghimire TR, Huettmann F, Regmi GR. Persistent Evidence for a Dramatic Decline in Langurs (*Semnopithecus* spp.) in Nepal and Elsewhere: Science Data and Personal Experiences Converge on a Landscape-Scale. In G. Regmi and F. Huettmann (Eds.) Hindu Kush-Himalaya Watersheds Downhill: Landscape Ecology and Conservation Perspectives 2020 (pp. 663-676). Springer, Cham. DOI: 10.1007/978-3-030-36275-1_33

Arunachalam K, Senthilvel K, Anbarasi P. Endo parasitic infections in free living Rhesus Macaque: (*Macaca mulatta*)

- of Namakkal, Tamil Nadu, India. Zoo's Print. 2015 Jun 21;30(6):20-1
- Beaver PC, Blanchard JL, Seibold HR. Invasive amebiasis in naturally infected New World and Old-World monkeys with and without clinical disease. The American Journal of Tropical Medicine and Hygiene. 1988 Oct 1;39(4): 343-52. DOI:10.4269/ajtmh.1988.39.343
- Bhattarai BP, Adhikari JN, Dhakal DN. Impact of prevalence of gastrointestinal parasites in rhesus macaque (*Macaca mulatta* Zimmermann, 1780) in Chitwan-Annapurna landscape, Nepal. International Journal of Zoology Studies. 2019 March; 4(2): 34-42
- Blanchard JL, Baskin GB. Trichomonas gastritis in rhesus monkeys infected with the simian immunodeficiency virus. Journal of Infectious Diseases. 1988 May 1;157(5):1092-3. DOI: 10.1093/infdis/157.5.1092
- Blanchard JL. Trichomonas gastritis. In Nonhuman Primates 1993 (pp. 38-40). Springer, Berlin, Heidelberg. DOI: 10.1007/978-3-642-84924-4_9
- Burrows RB. Protozoa of the intestinal tract. In Pathology of Simian Primates 1972 (pp. 2-28). Karger Publishers
- Chapman CA, Gillespie TR, Goldberg TL. Primates and the ecology of their infectious diseases: how will anthropogenic change affect host-parasite interactions? Evolutionary Anthropology: Issues, News, and Reviews: Issues, News, and Reviews. 2005 Jul;14(4):134-44. DOI: 10.1002/evan.20068
- Debenham JJ, Tysnes K, Khunger S, Robertson LJ. Occurrence of *Giardia, Cryptosporidium*, and *Entamoeba* in wild rhesus macaques (*Macaca mulatta*) living in urban and semi-rural North-West India. International Journal for Parasitology: Parasites and Wildlife. 2017 Apr 1;6(1):29-34. DOI: 10.1016/j.ijppaw.2016.12.002
- Fremming BD, Vogel FS, Benson RE, Young RJ. A fatal case of amebiasis with liver abscesses and ulcerative colitis in a chimpanzee. Journal of the American Veterinary Medical Association. 1955 May;126(938):406
- Fuentes A. Monkey forests and human landscapes: is extensive sympatry sustainable for *Homo sapiens* and *Macaca fascicularis* in Bali?. Commensalism and conflict: The primate-human interface. American Society of Primatology Publications; 2005. pp. 168–195
- Fuentes A. The International Encyclopedia of Primatology, 3 Volume Set. John Wiley & Sons; 2017 Apr 24
- Ghimire TR. Parasites and parasitic diseases in Nepal: Principle of Parasitology. LAP Lambert Academic Publishing. 2014. ISBN 978-3-659-53650-2
- Ghimire TR, Bhattarai N. A survey of gastrointestinal parasites of goats in a goat market in Kathmandu, Nepal. Journal of Parasitic Diseases. 2019 Dec 1; 43(4): 686-95. DOI: 10.1007/s12639-019-01148-w
- Ghimire TR, Regmi GR, Huettmann F. When Micro Drives the Macro: A Fresh Look at Disease and its Massive Contributions in the Hindu Kush-Himalaya. In G. Regmi and F. Huettmann (Eds.) Hindu Kush-Himalaya Watersheds Downhill: Landscape Ecology and Conservation Perspectives 2020 (pp.771-811). Springer, Cham. DOI: 10.1007/978-3-030-36275-1_40
- Gibbs RA, Rogers J, Katze MG, Bumgarner R, Weinstock GM, Mardis ER, Remington KA, Strausberg RL, Venter JC, Wilson RK, Batzer MA. Evolutionary and biomedical insights from the rhesus macaque genome. Science. 2007 Apr 13;316(5822): 222-34. DOI: 10.1126/science.1139247
- Guan Y, Feng M, Cai J, Min X, Zhou X, Xu Q, Tan N, Cheng X, Tachibana H. Comparative analysis of genotypic diversity in Entamoeba nuttalli isolates from Tibetan macaques and rhesus macaques in China. Infection, Genetics and Evolution. 2016 Mar 1; 38: 126-31. DOI: 10.1016/j.meegid.2015.12.014
- Haq A, Sharma A, Ahmad S, Khan HM, Khan N. Experimental

- infection of rhesus monkeys with *Entamoeba histolytica* mimics human infection. Laboratory Animal Science. 1985 Oct;35(5):481-4
- Haque R. Human intestinal parasites. Journal of Health, Population, and Nutrition. 2007 Dec;25(4):387
- Jaiswal AK, Sudan V, Kanojiya D, Sachan A, Shanker D. A pilot study on gastrointestinal parasites of monkeys (*Macaca mulatta*) of Mathura-Vrindavan areas, India. Journal of Veterinary Parasitology. 2014;28(1): 66-8
- Jha A, Chalise MK, Shrestha RM, Karki K. Intestinal parasitic investigation in temple Rhesus Monkeys of Kathmandu. The Initiation. 2011; 4: 1-7. DOI: 10.3126/init.v4i0.5530
- Jiang B, Yang GY, Yu XM, Cheng AC, Bi FJ. A case report: *Entamoeba histolytica* infections in the rhesus macaque, China. Parasitology Research. 2008 Sep 1;103(4): 915-7. DOI: 10.1007/s00436-008-1076-3
- Jokinen A. Monkeys and monkey gods in mythology, folklore, and religion. 2007. Accessed from: http://www.luminarium.org/mythology/monkeygods.htm, Retrieved: May 22, 2020
- Jones-Engel L, Engel GA, Heidrich J, Chalise M, Poudel N, Viscidi R, Barry PA, Allan JS, Grant R, Kyes R. Temple monkeys and health implications of commensalism, Kathmandu, Nepal. Emerging Infectious Diseases. 2006 Jun;12(6): 900. DOI: 10.3201%2Feid1206.060030
- Kaup FJ, Mätz-Rensing K, Kuhn EM, Hünerbein P, Stahl-Hennig C, Hunsmann G. Gastrointestinal pathology in rhesus monkeys with experimental SIV infection. Pathobiology. 1998;66(3-4): 159-64. DOI: 10.1159/000028015
- Knezevich M. Geophagy as a therapeutic mediator of endoparasitism in a free-ranging group of rhesus macaques (*Macaca mulatta*). American Journal of Primatology. 1998;44(1): 71-82. DOI: 10.1002/(sici)1098-2345(1998)44:1%3C71::aid-ajp6%3E3.0.co;2-u
- Kondova I, Simon MA, Klumpp SA, MacKey J, Widmer G, Domingues HG, Persengiev SP, O'Neil SP. Trichomonad gastritis in rhesus macaques (*Macaca mulatta*) infected with simian immunodeficiency virus. Veterinary Pathology. 2005 Jan;42(1): 19-29. DOI: 10.1354%2Fvp.42-1-19
- Kuhn EM, Mätz Rensing K, Stahl Hennig C, Makoschey B, Hunsmann G, Kaup FJ. Intestinal manifestations of experimental SIV infection in Rhesus Monkeys (*Macaca mulatta*): A histological and ultrastructural study. Journal of Veterinary Medicine, Series B. 1997 Jan 12;44(1-10): 501-12. DOI: 10.1111/j.1439-0450.1997.tb01001.x
- Kumar R, Sinha A, Radhakrishna S. Comparative demography of two commensal macaques in India: implications for population status and conservation. Folia Primatologica. 2013;84(6): 384-93. DOI: 10.1159/000351935
- Kumar S, Hedges SB. A molecular timescale for vertebrate evolution. Nature. 1998 Apr;392(6679): 917-20. DOI: 10.1038/31927
- Kumar S, Sundararaj P, Kumara HN, Pal A, Santhosh K, Vinoth S. Prevalence of gastrointestinal parasites in bonnet macaque and possible consequences of their unmanaged relocations. PloS one. 2018;13(11). DOI: 10.1371%2Fjournal.pone.0207495
- Levecke B, Dreesen L, Dorny P, Verweij JJ, Vercammen F, Casaert S, Vercruysse J, Geldhof P. Molecular identification of *Entamoeba* spp. in captive nonhuman primates. Journal of Clinical Microbiology. 2010 Aug 1;48(8): 2988-90. DOI: 10.1128/JCM.00013-10
- Li J, Dong H, Wang R, Yu F, Wu Y, Chang Y, Wang C, Qi M, Zhang L. An investigation of parasitic infections and review of molecular characterization of the intestinal protozoa in nonhuman primates in China from 2009 to 2015. International Journal for Parasitology: Parasites and Wildlife. 2017 Apr 1;6(1): 8-15. DOI: 10.1016/j.ijppaw.2016.12.003

- Loomis MR, Britt Jr JO, Gendron AP, Holshuh HJ, Howard EB. Hepatic and gastric amebiasis in black and white colobus monkeys. Journal of the American Veterinary Medical Association. 1983 Dec 1;183(11): 1188-91
- Macpherson CN. Epidemiology and control of parasites in nomadic situations. Veterinary Parasitology. 1994 Aug 1;54(1-3): 87-102. DOI: 10.1016/0304-4017(94)90085-X
- Pang VF, Chang CC, Chang WF. Concurrent gastric and hepatic amebiasis in a dusky leaf monkey (Presbytis obscurus). Journal of Zoo and Wildlife Medicine. 1993 Jun 1:204-7. https://www.jstor.org/stable/20095265
- Parmar SM, Jani RG, Mathakiya RA. Study of parasitic infections in non-human primates of Gujarat state, India. Veterinary World. 2012 Jun 1;5(6): 362. DOI: 10.5455/vetworld.2012.362-364
- Petrášová J, Modrý D, Huffman MA, Mapua MI, Bobáková L, Mazoch V, Singh J, Kaur T, Petrželková KJ. Gastrointestinal parasites of indigenous and introduced primate species of Rubondo Island National Park, Tanzania. International Journal of Primatology. 2010 Oct 1;31(5): 920-36. DOI: 10.1007/s10764-010-9439-x
- Pokhrel G, Maharjan M. Gastro-intestinal Parasites of Assamese Macaque (*Macaca assamensis* Hodgson, 1840) in Shivapuri Nagarjun National Park, Kathmandu, Nepal. Journal of Institute of Science and Technology. 2014;19(2): 53-7. DOI:10.3126/jist.v19i2.13852
- Rathoure AK. Conservation measures for monkey (*Macaca mulatta*) and Languor (*Presbytis entellus*). Octa Journal of Environmental Research. 2014 Dec 1; 3(4)
- Remfry J. The incidence, pathogenesis and treatment of helminth infections in rhesus monkeys (*Macaca mulatta*). Laboratory Animals. 1978 Oct 1;12(4): 213-8. DOI: 10.1258%2F002367778781088440
- Richards IM, Eady RP, Jackson DM, Orr TS, Pritchard DI, Vendy K, Wells E. Ascaris-induced bronchoconstriction in primates experimentally infected with *Ascaris suum* ova. Clinical and Experimental Immunology. 1983 Nov;54(2): 461, PMCID: PMC1535898, PMID: 6652969
- Schurer JM, Ramirez V, Kyes P, Tanee T, Patarapadungkit N, Thamsenanupap P, Trufan S, Grant ET, Garland-Lewis G, Kelley S, Nueaitong H. Long-Tailed Macaques (*Macaca fascicularis*) in Urban Landscapes: Gastrointestinal Parasitism and Barriers for Healthy Coexistence in Northeast Thailand. The American Journal of Tropical Medicine and Hygiene. 2019 Feb 6;100(2): 357-64. DOI: 10.4269/ajtmh.18-0241
- Soulsby EJ. Helminths, arthropods and protozoa of domesticated animals. Helminths, arthropods and protozoa of domesticated animals. 2012. Seventh Edition. Affiliated East-West Press Private Limited, New Delhi
- Sricharern W, Inpankaew T, Keawmongkol S, Supanam J, Stich RW, Jittapalapong S. Molecular detection and prevalence of *Giardia duodenalis* and *Cryptosporidium* spp. among longtailed macaques (*Macaca fascicularis*) in Thailand. Infection, Genetics and Evolution. 2016 Jun 1; 40: 310-4. DOI: 10.1016/j.meegid.2016.02.004
- Stauffer W, Ravdin JI. *Entamoeba histolytica*: an update. Current opinion in infectious diseases. 2003 Oct 1;16(5):479-85
- Swedell L. Primate sociality and social systems. Nature Education Knowledge. 2012;3(10):84
- Tabasshum T, Mukutmoni M, Begum A. Occurrence of gastrointestinal helminths in captive rhesus macaques (*Macaca mulatta*). Bangladesh Journal of Zoology. 2018 Dec 2;46(2):231-7. DOI: 10.3329/bjz.v46i2.39065

- Tachibana H, Yanagi T, Akatsuka A, Kobayashi S, Kanbara H, Tsutsumi V. Isolation and characterization of a potentially virulent species *Entamoeba nuttalli* from captive Japanese macaques. Parasitology. 2009 Sep;136(10):1169-77. DOI: 10.1017/S0031182009990576
- Tachibana H, Yanagi T, Lama C, Pandey K, Feng M, Kobayashi S, Sherchand JB. Prevalence of *Entamoeba nuttalli* infection in wild rhesus macaques in Nepal and characterization of the parasite isolates. Parasitology International. 2013 Apr 1;62(2):230-5. DOI: 10.1016/j.parint.2013.01.004
- Tachibana H, Yanagi T, Pandey K, Cheng XJ, Kobayashi S, Sherchand JB, Kanbara H. An *Entamoeba* sp. strain isolated from rhesus monkey is virulent but genetically different from *Entamoeba histolytica*. Molecular and Biochemical Parasitology. 2007 Jun 1;153(2):107-14. DOI: 10.1016/j.molbiopara.2007.02.006
- Thienpont D, Mortelmans J, Vercruysse J. Contribution to the study of trichuriasis in the chimpanzee and of its treatment with methyridine. Annales de la Societe belge de Medecine Tropicale (1920). 1962 Apr 30; 42:211
- Toft II JD, Eberhard ML. Parasitic diseases. In Nonhuman primates in biomedical research 1998 Jan 1 (pp. 111-205). Academic Press. DOI: 10.1016/B978-012088665-4/50005-1
- Toft JD. The pathoparasitology of nonhuman primates: a review. In Primates 1986 (pp. 571-679). Springer, New York, NY. DOI: 10.1007/978-1-4612-4918-4_45
- Verweij JJ, Vermeer J, Brienen EA, Blotkamp C, Laeijendecker D, van Lieshout L, Polderman AM. *Entamoeba histolytica* infections in captive primates. Parasitology Research. 2003 Jun 1;90(2): 100-3. DOI: 10.1007/s00436-002-0808-z
- Weiszer I, Patterson R, Pruzansky JJ. *Ascaris* hypersensitivity in the rhesus monkey: I. A model for the study of immediate type hypersensitivity in the primate. Journal of Allergy and Clinical Immunology. 1968 Jan 1;41(1):14-22. DOI: 10.1016/0021-8707(68)90004-X
- Ye J, Xiao L, Li J, Huang W, Amer SE, Guo Y, Roellig D, Feng Y. Occurrence of human-pathogenic *Enterocytozoon bieneusi*, *Giardia duodenalis* and *Cryptosporidium* genotypes in laboratory macaques in Guangxi, China. Parasitology International. 2014 Feb 1;63(1):132-7. DOI: 10.1016/j.parint.2013.10.007
- Ye J, Xiao L, Ma J, Guo M, Liu L, Feng Y. Anthroponotic enteric parasites in monkeys in public park, China. Emerging Infectious Diseases. 2012 Oct;18(10):1640. DOI: 10.3201%2Feid1810.120653
- Zhang Q, Liu K, Wang C, Luo J, Lu J, He H. Molecular characterization of *Entamoeba* spp. in wild Taihangshan macaques (*Macaca mulatta tcheliensis*) in China. Acta Parasitologica. 2019 Jun 1;64(2): 228-31. DOI: 10.2478/s11686-019-00026-y
- Zhao W, Zhou H, Jin H, Liu M, Qiu M, Li L, Yin F, Chan JF, Lu G. Molecular prevalence and subtyping of *Cryptosporidium hominis* among captive long-tailed macaques (*Macaca fascicularis*) and rhesus macaques (*Macaca mulatta*) from Hainan Island, southern China. Parasites & Vectors. 2019 Dec;12(1): 192. DOI: 10.1186/s13071-019-3449-0
- Zvinorova PI, Halimani TE, Muchadeyi FC, Matika O, Riggio V, Dzama K. Prevalence and risk factors of gastrointestinal parasitic infections in goats in low-input low-output farming systems in Zimbabwe. Small Ruminant Research. 2016 Oct 1; 143:75-83. DOI: 10.1016/j.smallrumres.2016.09.005