# Green Synthesis of Silver Nanoparticles from *Ocimum sanctum* Linn. and Study of Their Antioxidant Activity

#### Ishwor Pathak<sup>1</sup>\*, Muna Niraula<sup>2</sup>, Khaga Raj Sharma<sup>3</sup>, Prakash Thapa<sup>4</sup>, Hari Bhakta Oli<sup>1</sup>, Surya Kant Kalauni<sup>3</sup>\*

<sup>1</sup> Department of Chemistry, Amrit Campus, Tribhuvan University, Thamel, Kathmandu, Nepal

<sup>2</sup> Department of Forests and Soil Conservation, Babarmahal, Kathmandu, Nepal

<sup>3</sup> Central Department of Chemistry, Tribhuvan University, Kirtipur, Kathmandu, Nepal

<sup>4</sup> University of Toledo, Department of Chemistry and Biochemistry, Ohio, USA

\*E-mail: pathakishwor14@gmail.com, skkalauni@gmail.com

(Received: August 21, 2024; Received in Revised Form: November 9, 2024; Accepted: November 12, 2024; Available online)

DOI: https://doi.org/10.3126/arj.v5i1.73553

# Highlights

- Methanol extract of Ocimum sanctum Linn prepared and phytochemically tested.
- Silver nanoparticles were green synthesized using methanol extract.
- FESEM, EDS, and XRD analysis confirm successful synthesis of OCE-AgNPs.
- OCE-AgNPs exhibit remarkable antioxidant activity.

# Abstract

Green synthesis of silver nanoparticles (AgNPs) has gained considerable attention due to their unique properties and wide-ranging applications. This study focuses on the synthesis of AgNPs utilizing the methanol extract of aerial parts of *Ocimum sanctum* Linn., commonly known as holy basil or Tulsi. The synthesis process is environmentally friendly and offers a sustainable alternative to conventional methods. Characterization of the synthesized AgNPs is performed using various techniques, including field emission scanning electron microscopy (FESEM), energy-dispersive X-ray spectroscopy (EDX), and X-ray diffraction (XRD). The results demonstrate the successful synthesis of AgNPs with distinct morphologies and crystalline structures. Additionally, the antioxidant activity of the AgNPs is evaluated using the 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay, with ascorbic acid as a reference standard. The synthesized OCE-AgNPs exhibit an IC50 value of 49.71  $\mu$ g/mL, which is close to that of the standard ascorbic acid (41.34  $\mu$ g/mL). The results of this study highlight the potential applications of OCE-AgNPs in biomedical and pharmaceutical fields.

Keywords: Silver nanoparticles, Ocimum sanctum, green synthesis, antioxidant activity, DPPH assay

# Introduction

Silver nanoparticles (AgNPs) have emerged as promising materials in various fields, including medicine, electronics, and catalysis, owing to their unique physical, chemical, and biological properties. [1], [2], [3] These nanoparticles are characterized by their high surface area-to-volume ratio, exceptional electrical and thermal conductivity, high antioxidant activity, and potent antimicrobial properties, making them highly valuable for a myriad of applications. [4], [5] In particular, AgNPs are being explored for their potential in drug delivery systems, diagnostic tools, and as antioxidant and antimicrobial agents in medical

<sup>\*</sup>Corresponding author

#### I. Pathak et al., 2024

devices. [6], [7] The increasing demand for multifunctional nanomaterials has driven extensive research into optimizing their synthesis and functionalization.

*Ocimum sanctum*, commonly known as holy basil or tulsi, is a medicinal plant revered in traditional medicine, particularly in Ayurveda, for its extensive health benefits. [8], [9] This plant is rich in various phytochemicals, including flavonoids, alkaloids, phenolic acids, and essential oils, which confer diverse biological activities such as antioxidant, anti-inflammatory, antimicrobial, and anticancer properties. [10], [11] The high antioxidant value is particularly noteworthy, as it is crucial in neutralizing free radicals. [12], [13], [14], [15] Free radicals are unstable molecules that can cause oxidative stress, leading to cellular damage and contributing to the development of chronic diseases such as cancer, cardiovascular diseases, and neurodegenerative disorders. [16], [17] By scavenging these free radicals, the antioxidants in *O. sanctum* help in preventing oxidative stress and its associated health issues.[18] Moreover, the plant's anti-inflammatory properties aid in reducing inflammation, while its antimicrobial properties help in combating various pathogens, making it a versatile and highly valued plant in traditional and modern medicine. [19], [20]

In recent years, there has been growing interest in the green synthesis of AgNPs using plant extracts. This method offers several advantages over conventional chemical synthesis, which often involves toxic solvents, hazardous reducing agents, and complex procedures. [21], [22] Green synthesis is not only cost-effective and eco-friendly but also scalable, making it suitable for large-scale production. Plant extracts serve as natural reducing and stabilizing agents, simplifying the process and eliminating the need for harmful chemicals. [23] This approach leverages the inherent biological activity of the plant extracts, which can enhance the functional properties of the synthesized nanoparticles. The use of plant-based synthesis aligns with the principles of sustainable development, reducing the environmental footprint of nanomaterial production. Additionally, green synthesis methods contribute to the circular economy by utilizing renewable resources and generating less hazardous waste. This environmentally benign approach also opens the possibility of integrating traditional medicinal knowledge with modern nanotechnology, potentially leading to the discovery of novel therapeutic agents and treatments. The integration of green chemistry principles in nanoparticle synthesis is a significant step forward in achieving sustainable and responsible nanotechnology development. All in all, the utilization of *O. sanctum* for the green synthesis of AgNPs not only takes advantage of the plant's rich phytochemical profile and diverse biological activities but also aligns with the global shift towards more sustainable and eco-friendly synthetic processes. This holistic approach not only enhances the therapeutic potential of AgNPs but also underscores the importance of integrating traditional medicinal medicinal knowledge with cutting-edge nanotechnology.

Taking into consideration of the above-discussed advantages, we utilized methanol extract of aerial parts of *O. sanctum* for the green synthesis of silver nanoparticles (AgNPs). The synthesized nanoparticles were extensively characterized and evaluated for their antioxidant activity. The extract served as a natural reducing and stabilizing agent in the synthesis process. The IC50 value of OCE-AgNPs is comparable to that of L-ascorbic acid, demonstrating its potent antioxidant efficacy. This study confirms the successful formation of AgNPs with significant antioxidant properties, highlighting the potential of *O. sanctum* in eco-friendly nanoparticle synthesis and suggesting promising applications in biomedical research and healthcare

# **Experimental Section**

#### Chemicals and plant materials

Silver nitrate (AgNO<sub>3</sub>), L-Ascorbic acid, 1,1-Diphenyl-2-picrylhydrazyl (DPPH), and methanol were purchased from Sigma– Aldrich Co., St. Louis, MO, USA. Deionized water was utilized in the present analysis. Fresh and healthy aerial parts of the experimental plant, *Ocimum sanctum L*. were collected from Dhading, Nepal. The plant parts were thoroughly rinsed with tap water, followed by distilled water, to remove dust particles, pathogens, and adhered matter. After that, it was shade-dried for 4 days and ground to a fine powder.

#### Preparation of the O. sanctum extract

The grounded powder was subjected to a cold percolation technique using methanol as the solvent (500 mL  $\times$  24 hours  $\times$  5 cycles). The methanol extract was then concentrated using a rotary evaporator. The concentrated extract was dried and stored in a sealed glass vial in a refrigerator until further experimentation.

#### Green synthesis of silver nano particles

Silver nanoparticles were synthesized using a slightly modified version of the previously reported methods. [24], [25], [26] Initially, the biological reduction of AgNO<sub>3</sub> was performed as follows: 3 mL of plant extract was added to 2 mL of 0.02 M AgNO<sub>3</sub> solution, and the volume was adjusted to 20 mL with deionized water in a 50 mL volumetric flask. This mixture was kept at ambient temperature ( $25\pm0.5$  °C) for 24 hours. The mixture's color transitioned from light to yellowish, then to reddish, and finally to colloidal. This color change was monitored periodically. After the synthesis and completion of the reaction, the solution was centrifuged at 10,000 rpm for 20 minutes and the silver nanoparticles were collected. The as-synthesized *O. sanctum* extract-derived silver nanoparticles were abbreviated as OCE-AgNPs.

#### **Phytochemical screening**

The methanol extract (1 g) was completely dissolved in 100 mL of methanol to prepare a stock solution. This stock solution was then used for phytochemical screening, following established protocols with slight modifications. [13], [27], [28]

#### **Physical characterization**

The morphology and the constituent elements of the synthesized OCE-AgNPS were studied using a Field Emission Scanning Electron Microscopy (FESEM, Hitachi, Japan) equipped with an energy dispersive spectrometer (EDX). The crystallinity of the nanoparticles was studied with an X-ray diffractometer (Rigaku Corporation, Japan).

#### Antioxidant activity test (DPPH radical scavenging activity)

1,1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging activity of the methanol extract and OCE-AgNPs was assessed following our previous study with slight modifications.[29], [30], [31], [32] DPPH (7.886 g) was dissolved in 100 mL methanol to prepare a 0.2 mM solution. Various concentrations (10, 30, 50, 70, 90, and 100  $\mu$ g/mL) of methanol extract, OCE-AgNPs, and ascorbic acid (positive control) were tested. Each concentration (2 mL) was mixed with 2 mL of 0.2 mM DPPH solution, and absorbance was measured at 517 nm using a UV-Visible spectrophotometer (Shimadzu UV professional double beam) after 30 minutes. The experiment was performed in triplicate, and radical scavenging activity was calculated using the equation 1:

% radical scavenging activity =  $[(Abs._{control} - Abs._{sample})/Abs._{control})] \times 100\%$  (1)

where, Abs.<sub>control</sub> is the absorbance of the control (1 mL MeOH + 0.5 mL DPPH) and Abs.<sub>sample</sub> is the absorbance of the sample. IC50 values, representing the concentration needed to scavenge 50% of free radicals, were determined from concentration versus percentage scavenging activity curves for each sample.

## **Results and Discussion**



Fig 1. Digital photographs of the synthetic steps involved in the preparation of OCE-AgNPs.

**Figure 1** depicts each step involved in the synthesis of the OCE-AgNPs. As illustrated in the figure, cleaned and dried aerial parts of the *O. sanctum* were shed dried, and ground into powder. The methanol extract was extracted from the dried powder and finally, Ag NPs were synthesized using the as-prepared extract. Phytochemical screening of methanol extract of *O. sanctum Linn*. was carried out according to the standard procedure. The results indicated that the extract is rich in flavonoids and polyphenols, along with the presence of alkaloids, terpenoids, tannins, and steroids. The high content of flavonoids and polyphenols suggests that the plant has significant antioxidant potential. However, the extract lacks glycosides, quinones, proteins, and saponins.

S.N.	Class of compounds	Methanol extract
1	Alkaloids	+
2	Flavonoids	++
4	Terpenoids	+
5	Glycosides	-
6	Quinones	-
7	Polyphenols	++
8	Tannins	+
9	Steroids	+
10	Proteins	-
11	Saponins	-

Table 1.	Phytochemical	screening	of the	methanol	extract
----------	---------------	-----------	--------	----------	---------

(+) indicates present and (-) indicates absent.

FESEM analysis (**Figure 2a-d**) revealed the formation of almost spherical AgNPs with a uniform size distribution. The EDX color mapping images and EDX spectrum (**Figure 3a-k**) confirmed the homogeneous distribution of elemental silver, along with other elements such as C, N, O, Mg, Cl, and K, which are originated from the plant extract. The XRD analysis (**Figure 4**) identified the crystalline nature of the synthesized AgNPs, which perfectly matched the JCPDS card of Silver (PDF#04-0783). The characteristic peaks at 38.12 °, 44.27 °, 64.43 °, 77.47 °, and 81.54 ° corresponding to the (111), (200), (220), (311), and (222) planes of face-centered cubic (fcc) silver nanoparticles. Furthermore, the broad peak at around 25 ° is due to the (002) peak of carbon which arises from the secondary metabolites present in the plant. Furthermore, the carbon content as seen in the XRD peaks is supported by the intense carbon peak and elemental color mapping in the EDX results.



Fig 2. (a-d) OCE-Ag NPs at increasing magnifications.



Fig 3. EDX analysis of OCE-Ag NPs (a) EDX color mapping area, (b) superimposition of all elements. EDX color mapping image for (c) C, (d) N, (e) O, (f) Mg, (g) Cl, (h) Ag, (i) K, and (j) Ca. (i) EDX spectrum (inset: element percentage).

The DPPH assay demonstrated that the synthesized OCE-AgNPs exhibited significant antioxidant activity, as evidenced by their ability to scavenge DPPH radicals in a concentration-dependent manner. The antioxidant activity of OCE-AgNPs was found to be comparable to that of ascorbic acid, indicating their potential as natural antioxidants.



Fig 4. XRD pattern of OCE-Ag NPs.

A DPPH assay was conducted to analyze the antioxidant activity of methanol extract and OCE-AgNPs, using ascorbic acid as a standard reference. Different concentrations of methanol extract, OCE-AgNPs, and ascorbic acid were incubated at room temperature, and their absorbance was measured with a spectrophotometer. The percentage of free radical scavenging and the IC50 values for each samples were calculated and are presented in **Table 2** and **Figure 5**.

Concentration (µg/mL)	% free radical scavenging activity				
	Methanol extract	OCE-Ag NPs	Ascorbic acid		
0	0	0	0		
10	27.36	42.69	48.57		
30	40.2	53.72	57.14		
50	48.61	58.62	62.85		
70	53.61	63.52	67.14		
90	56.11	67.3	74.28		
110	57.75	69.89	77.14		
IC <sub>50</sub> value(µg/mL)	72.78	49.71	41.34		

**Table 2:** DPPH free radical scavenging activity and  $IC_{50}$  values of different samples.

values are expressed as mean  $\pm$  SD (n=3)



Fig 5. % radical scavenging activity of methanol extract, OCE-AgNPs and ascorbic acid at different concentrations.

From the calculations, the inhibitory concentration for 50% inhibition (IC50) value of OCE-AgNPs is 49.71 µg/mL, which is close to that of standard ascorbic acid (41.34 µg/mL). The IC50 value of methanol extract is 72.78 µg/mL, substantially lower than that of both OCE-AgNPs and ascorbic acid. The closer the IC50 value of a sample is to that of ascorbic acid, the higher its antioxidant activity. The antioxidant activity of OCE-AgNPs is highly effective, being comparable to or even surpassing the results reported in earlier studies on *Ocimum sanctum* extracts and green-synthesized silver nanoparticles. This highlights the enhanced potential of OCE-AgNPs, which combine the bioactive properties of the plant extract with the unique capabilities of silver nanoparticles for superior free radical scavenging.[8], [33], [34], [35], [36] The presence of secondary metabolites such as flavonoids and polyphenols, which have the ability to scavenge free radicals, might explain the antioxidant activity exhibited by the OCE-AgNPs. [5], [37] Furthermore, the close IC50 value of OCE-AgNPs to that of ascorbic acid can be attributed to several factors. First, silver nanoparticles themselves can exhibit antioxidant properties. Second, when synthesized with plant extracts, the nanoparticles are often capped and stabilized by phytochemicals from the plant, potentially enhancing their antioxidant capacity. Third, the combination of silver nanoparticles with flavonoids, polyphenols, and other bioactive compounds from the extract can lead to higher antioxidant activity compared to the extract alone.[38]

### Conclusions

In conclusion, this study successfully synthesized AgNPs using the leaf extract of *Ocimum sanctum* via a green and sustainable approach. The synthesized OCE-AgNPs exhibited distinct morphologies, crystalline structures, and notable antioxidant activity,

#### Amrit Research Journal, Dec 2024, Vol. 5

highlighting their potential for applications in various fields, including the biomedical and pharmaceutical field. Further research is warranted to explore the therapeutic efficacy, toxicity, and stability of OCE-AgNPs for practical applications.

# **Acknowledgments:**

The authors acknowledge the Department of Forests and Soil Conservation, Babarmahal, Kathmandu, for providing the laboratory facilities to conduct this research.

# **References:**

- K. Baruah, M. Haque, L. Langbang, S. Das, K. Aguan, A. Singha Roy, *Ocimum sanctum* mediated green synthesis of silver nanoparticles: A biophysical study towards lysozyme binding and anti-bacterial activity, Journal of Molecular Liquids, 2021, 337, 116422. (DOI: https://doi.org/10.1016/j.molliq.2021.116422).
- M.M. U Din, A. Batool, R.S. Ashraf, A. Yaqub, A. Rashid, N.M. U Din, Green Synthesis and Characterization of Biologically Synthesized and Antibiotic-Conjugated Silver Nanoparticles followed by Post-Synthesis Assessment for Antibacterial and Antioxidant Applications, ACS Omega, 2024, 9(17). (DOI: 18909-18921. https://doi.org/10.1021/acsomega.3c08927.
- D.K. Shrestha, D.R. Jaishi, I. Ojha, D.R. Ojha, I. Pathak, A.B. Magar, N. Parajuli, K.R. Sharma, Plant assisted synthesis of silver nanoparticles using *Persicaria perfoliata* (L.) for antioxidant, antibacterial, and anticancer properties, Heliyon, 2024, 10(23). (DOI: https://doi.org/10.1016/j.heliyon.2024.e40543).
- W.M. Alamier, M. D Y Oteef, A.M. Bakry, N. Hasan, K.S. Ismail, F.S. Awad, Green Synthesis of Silver Nanoparticles Using *Acacia ehrenbergiana* Plant Cortex Extract for Efficient Removal of Rhodamine B Cationic Dye from Wastewater and the Evaluation of Antimicrobial Activity, ACS Omega, 2023, 8(21). (DOI: 18901-18914. https://doi.org/10.1021/ acsomega.3c01292.
- L.N. Khanal, K.R. Sharma, H. Paudyal, K. Parajuli, B. Dahal, G.C. Ganga, Y.R. Pokharel, S.K. Kalauni, Green Synthesis of Silver Nanoparticles from Root Extracts of *Rubus ellipticus* Sm. and Comparison of Antioxidant and Antibacterial Activity, Journal of Nanomaterials, 2022, (1), 1832587. (DOI: https://doi.org/https://doi.org/10.1155/2022/1832587).
- O.A. Adeleye, O.K. Aremu, H. Iqbal, M.O. Adedokun, O.A. Bamiro, O.L. Okunye, M.N. Femi-Oyewo, K.O. Sodeinde, Z.S. Yahaya, A.O. Awolesi, Green Synthesis of Silver Nanoparticles Using Extracts of *Ehretia cymosa* and Evaluation of Its Antibacterial Activity in Cream and Ointment Drug Delivery Systems, Journal of Nanotechnology, 2023, (1), 2808015. (DOI: https://doi.org/https://doi.org/10.1155/2023/2808015).
- M. Kandiah, K.N. Chandrasekaran, Green Synthesis of Silver Nanoparticles Using *Catharanthus roseus* Flower Extracts and the Determination of Their Antioxidant, Antimicrobial, and Photocatalytic Activity, Journal of Nanotechnology, 2021, (1), 5512786. (DOI: https://doi.org/https://doi.org/10.1155/2021/551278).
- S. Mishra, S. Sundaram, S. Srivastava, R. Dhar, Phytosynthesis of Silver Nanoparticles Using *Ocimum sanctum* Leaf Extract and Studies on Its Antidiabetic, Antioxidant, and Antibacterial Properties, ACS Applied Bio Materials, 2023, 6(10), 4127-4137. (DOI: https://doi.org/10.1021/acsabm.3c00234).
- 9. I. Pathak, M. Niraula, Assessment of total phenolic, flavonoid content and antioxidant activity of *Ocimum sanctum* Linn, Journal of Nepal Chemical Society, 2019, 40, 30-35. (DOI: https://doi.org/10.3126/jncs.v40i0.27275)
- D. Singh, P.K. Chaudhuri, A review on phytochemical and pharmacological properties of Holy basil (*Ocimum sanctum* L.), Industrial Crops and Products, 2018, 118, 367-382. (DOI: https://doi.org/10.1016/j.indcrop.2018.03.048).
- 11. Y. Sharma, M. Bharadwaj, N. Srivastava, A. Kaur, M. Kumar, M. Agarwal, Y. Bahl, K. Bala, In vitro antioxidant activity of defatted seed extracts of *Ocimum sanctum* on rat PC-12 cells and its inhibitory efficacy with receptors of oral squamous cell carcinoma, Industrial Crops and Products, 2020, 154, 112668. (DOI: https://doi.org/10.1016/j.indcrop.2020.112668).

- 12. I. Pathak, R. Budhathoki, N. Yadav, M. Niraula, S.K. Kalauni, Phytochemical screening, cytotoxic and antioxidant activity of *Alternathera sessilis* and *Moringa oleifera*, Amrit Research Journal, 2020, 1(1), 65-71.
- 13. I. Pathak, S. Rokaha, K.B. Bajracharya, Phytoconstituents and biological activities of *Zanthoxylum armatum* fruit extract, Journal of Nepal Chemical Society, 2021, 42(1), 125-131.
- 14. R. Timilsina, P. Tandukar, I. Pathak, Biological and chemical studies of essential oil and extracts of rhizome of *Acorus calamus* Linn, Journal of Nepal Chemical Society, 2022, 43(1), 35-42.
- 15. I. Pathak, M. Niraula, P. Thapa, Biological and chemical studies of essential oil from *Vitex negundo* of Nepalese origin, Journal of Nepal Chemical Society, 2018, 39, 18-24.
- 16. K. Jomova, R. Raptova, S.Y. Alomar, S.H. Alwasel, E. Nepovimova, K. Kuca, M. Valko, Reactive oxygen species, toxicity, oxidative stress, and antioxidants: Chronic diseases and aging, Archives of toxicology, 2023, 97(10), 2499-2574.
- R. Marahatha, K. Gyawali, K. Sharma, N. Gyawali, P. Tandan, A. Adhikari, G. Timilsina, S. Bhattarai, G. Lamichhane, A. Acharya, I. Pathak, H.P. Devkota, N. Parajuli, Pharmacologic activities of phytosteroids in inflammatory diseases: Mechanism of action and therapeutic potentials, Phytotherapy Research, 2021, 35(9), 5103-5124. (DOI: https://doi.org/ https://doi.org/10.1002/ptr.7138).
- D.K. Shrestha, A.B. Magar, M. Bhusal, R. Baraili, I. Pathak, P.R. Joshi, N. Parajuli, K.R. Sharma, Synthesis of Silver and Zinc Oxide Nanoparticles Using *Polystichum lentum* Extract for the Potential Antibacterial, Antioxidant, and Anticancer Activities, Journal of Chemistry, 2024, (1), 1876560. (DOI: https://doi.org/https://doi.org/10.1155/2024/1876560).
- 19. M.R. Hasan, B.S. Alotaibi, Z.M. Althafar, A.H. Mujamammi, J. Jameela, An Update on the Therapeutic Anticancer Potential of *Ocimum sanctum* L.: "Elixir of Life", Molecules, 2023, 28(3), 1193.
- 20. A.K. Singh, *Ocimum sanctum* mediated phytosynthesis of metallic nanoparticles: A review, Bioresource Technology Reports, 2022, 19, 101118. (DOI: https://doi.org/https://doi.org/10.1016/j.biteb.2022.101118).
- H.B. Habeeb Rahuman, R. Dhandapani, S. Narayanan, V. Palanivel, R. Paramasivam, R. Subbarayalu, S. Thangavelu, S. Muthupandian, Medicinal plants mediated the green synthesis of silver nanoparticles and their biomedical applications, IET Nanobiotechnology, 2022, 16(4), 115-144. (DOI: https://doi.org/https://doi.org/10.1049/nbt2.12078).
- N.S. Alharbi, N.S. Alsubhi, A.I. Felimban, Green synthesis of silver nanoparticles using medicinal plants: Characterization and application, Journal of Radiation Research and Applied Sciences, 2022, 15(3), 109-124. (DOI: https://doi.org/10.1016/j. jrras.2022.06.012).
- N.K. Sharma, J. Vishwakarma, S. Rai, T.S. Alomar, N. AlMasoud, A. Bhattarai, Green Route Synthesis and Characterization Techniques of Silver Nanoparticles and Their Biological Adeptness, ACS Omega, 2022, 7(31), 27004-27020. (DOI: https:// doi.org/10.1021/acsomega.2c01400).
- V. Ravichandran, S. Vasanthi, S. Shalini, S.A. Ali Shah, R. Harish, Green synthesis of silver nanoparticles using *Atrocarpus altilis* leaf extract and the study of their antimicrobial and antioxidant activity, Materials Letters, 2016, 180, 264-267. (DOI: https://doi.org/10.1016/j.matlet.2016.05.172).
- 25. M. Bhusal, I. Pathak, A. Bhadel, D.K. Shrestha, K.R. Sharma, Synthesis of silver nanoparticles assisted by aqueous root and leaf extracts of *Rhus chinensis* Mill and its antibacterial activity, Heliyon, 2024, 10, e33603.
- A. Rana, A.K. Chaudhary, S. Saini, R. Srivastava, M. Kumar, S.N. Sharma, Ultrafast transient absorption spectroscopic (UFTAS) and antibacterial efficacy studies of phytofabricated silver nanoparticles using *Ocimum Sanctum* leaf extract, Inorganic Chemistry Communications 2023, 147, 110233. (DOI: https://doi.org/10.1016/j.inoche.2022.110233).
- B. Subba, A. Sharma, A. Budhathoki, Assessment of phytochemical content, antioxidant and antibacterial activities of three medicinal plants of Nepal, Journal of Medicinal Plants Research, 2016, 10(45), 829-837.
- A. Hassan, Z. Akmal, N. Khan, The Phytochemical Screening and Antioxidants Potential of *Schoenoplectus triqueter* L. Palla, Journal of Chemistry, 2020, (1), 3865139. (DOI: https://doi.org/10.1155/2020/3865139).

#### Amrit Research Journal, Dec 2024, Vol. 5

- 29. S.K. Kalauni, M. Niraula, P. Thapa, I. Pathak, Comparative Studies on Antioxidant Activity of Ten Medicinal Plants Collected From the Ilam District of Nepal, Nepal Journal of Science and Technology, 2021, 20(1), 136-145.
- 30. S.K. Kalauni, R. Maharjan, I. Pathak, K. Khadayat, M. Niraula, P. Thapa, Different Crude Extracts of *Cinnamomum tamala* with Antioxidant and Antibacterial Capabilities, Amrit Research Journal, 2021, 2(01), 68-74.
- 31. P. Tandukar, N. Das, I. Pathak, D.R. Gautam, GC-MS Profiling and Bioactivities of Essential Oil and Extracts of *Cinnamomum tamala* (Buch.-Ham.) Nees & Eberm. Leaves from Kathmandu Valley, Nepal, Amrit Research Journal, 2022, 3(01), 56-66.
- N. Das, P. Tandukar, M. Niraula, D.R. Gautam, I. Pathak, Phytochemical Analysis and Biological Activities of Different Extracts of Walnut (*Juglans regia* Linn.) Kernels, Journal of Nepal Chemical Society, 2024, 44(2), 78-89. (DOI: https://doi. org/10.3126/jncs.v44i2.68319).
- 33. D.B. Manikandan, A. Sridhar, R. Krishnasamy Sekar, B. Perumalsamy, S. Veeran, M. Arumugam, P. Karuppaiah, T. Ramasamy, Green fabrication, characterization of silver nanoparticles using aqueous leaf extract of *Ocimum americanum* (Hoary Basil) and investigation of its in vitro antibacterial, antioxidant, anticancer and photocatalytic reduction, Journal of Environmental Chemical Engineering, 2021, 9(1), 104845. (DOI: https://doi.org/10.1016/j.jece.2020.104845).
- H. Bagur, C.C. Poojari, G. Melappa, R. Rangappa, N. Chandrasekhar, P. Somu, Biogenically Synthesized Silver Nanoparticles Using Endophyte Fungal Extract of *Ocimum tenuiflorum* and Evaluation of Biomedical Properties, Journal of Cluster Science, 2020, 31(6), 1241-1255. (DOI: https://doi.org/10.1007/s10876-019-01731-4).
- 35. K. Kavitha, S. Ponne, In Vitro Antioxidant and Free Radical Scavenging Activities of Methanolic Extract of *Ocimum* sanctum Linn. Seed, 2015, 2(1).
- 36. J. Samson, R. Sheeladevi, R. Ravindran, Oxidative stress in brain and antioxidant activity of *Ocimum sanctum* in noise exposure, NeuroToxicology, 2007, 28(3), 679-685. (https://doi.org/10.1016/j.neuro.2007.02.011).
- M.S. Swallah, H. Sun, R. Affoh, H. Fu, H. Yu, Antioxidant Potential Overviews of Secondary Metabolites (Polyphenols) in Fruits, International Journal of Food Science, 2020, (1), 9081686. (https://doi.org/10.1155/2020/9081686).
- D.R. Jaishi, I. Ojha, G. Bhattarai, R. Baraili, I. Pathak, D.R. Ojha, D.K. Shrestha, K.R. Sharma, Plant-mediated synthesis of zinc oxide (ZnO) nanoparticles using *Alnus nepalensis* D. Don for biological applications, Heliyon, 2024, 10(20). (DOI: https://doi.org/10.1016/j.heliyon.2024.e39255).