

Antimicrobial Susceptibility Pattern of *Escherichia coli* Isolated from Urinary Tract Infected Patients Attending Bir Hospital

Amit Raj Sharma¹, Dwij Raj Bhatta¹, Jyotsna Shrestha² and Megha Raj Banjara¹

¹Central Department of Microbiology, Tribhuvan University, Kathmandu

²Pathology Department, Bir Hospital, Kathmandu

e-mail: amritrsharma@gmail.com

Abstract

Antibiotic resistance among uropathogens is emerging public health problem. This study was done for assessing antibiotic and multidrug resistance (MDR) patterns of *Escherichia coli* at Bir Hospital, Kathmandu, among suspected urinary tract infection (UTI) patients from January to March, 2011. Altogether, 739 urine samples were analysed by semi-quantitative culture method and uropathogens were identified by conventional methods. *E. coli* was tested (109 samples) for antimicrobial susceptibility by Kirby Bauer disc diffusion method as per Clinical and Laboratory Standard Institute (CLSI) guidelines. Out of 739 samples, 27.3% gave significant growth while 3.1% and 29.2% samples gave mixed and non-significant growth respectively. *E. coli* was found to be most predominant isolate (54.0%) followed by coagulase negative *Staphylococci* (CoNS) (21.3%) and *Enterococcus* spp. (7.3%). Nitrofurantoin was found to be the most effective antibiotic followed by ciprofloxacin and ofloxacin while cephalexin was least effective. Out of 109 *E. coli* isolates, 90.8% were MDR strains and most of the isolates had a very high multiple antibiotic resistance (MAR) index, suggesting the origin of the isolates to be of high antibiotic usage. *E. coli* showed higher rate of resistance towards commonly used oral antibiotics. However, nitrofurantoin is still active against organisms. Thus, nitrofurantoin could be the choice for empirical therapy of UTI.

Key words: *Escherichia coli*, multidrug resistance, multiple antibiotic resistance index, urinary tract infection

Introduction

Urinary tract infections (UTIs) are one of the most common infectious diseases ranking next to upper respiratory tract infection. UTIs are often associated with significant morbidity and mortality (Ramesh *et al.* 2008). Worldwide, about 150 million people are diagnosed with urinary tract infection each year, costing the global economy in excess of 6 billion dollars (Gonzalez & Schaeffer 1999). *E. coli*, the most common member of the family Enterobacteriaceae, accounts for 75.0-90.0% of all UTIs in both inpatients and outpatients (Dromigny *et al.* 2005). *E. coli* present in the gastrointestinal tract as a commensal provide the pool for initiation of UTI and certain serotypes of *E. coli* responsible for uropathogenicity were traditionally designated as uropathogenic *E. coli* (UPEC) (Raksha *et al.* 2003). UTI is a common disease ailment among Nepalese population as well as one of the commonest nosocomial infection (Kattel *et al.* 2008). According to the annual report of fiscal year (2010/2011) published by the Department of Health Services,

morbidity of UTI among outpatients were 265,143. The present study was conducted to determine the prevalence and antibiotic susceptibility pattern of *E. coli* associated urinary tract infection among inpatients and outpatients.

Methodology

In a descriptive cross sectional study conducted from January to March 2011, a total of 739 urine samples from suspected UTI patients visiting Bir Hospital, Kathmandu were processed in the microbiology laboratory of this hospital for the isolation of *E. coli*. Each sample was mixed well and aseptically inoculated with 4 mm diameter nichrome wire loop on blood agar and MacConkey agar plates and incubated at 37°C for 24 hours aerobically. Significant UTI was defined as the presence of >10⁵ colony forming unit (CFU)/ml in the culture. All positive cultures were further identified by their cultural characteristics, Gram stain and battery of biochemical reaction. *E. coli* was identified on the

basis of triple sugar iron agar (TSI), sulphide indole motility (SIM) test medium, citrate utilization and urease production. The antimicrobial susceptibility testing (AST) of *E. coli* isolates was done by Kirby Bauer Disc Diffusion Method as per CLSI guideline (CLSI 2011). The antibiotic discs of HiMedia (India) used were ampicillin (10 µg), cephalexin (30 µg), nalidixic acid (30 µg), ciprofloxacin (5 µg), ofloxacin (5 µg), norfloxacin (10 µg), nitrofurantoin (300 µg) and co-trimoxazole (1.25/23.75 µg). A reference strain, *E. coli* ATCC 25922 was used as control. Multiple antibiotic resistance (MAR) index was determined using the formula $MAR = x/y$, where x was number of antibiotics to which test isolate displayed resistance and y is the total number of antibiotics to which the test organism has been evaluated for sensitivity (Akinjogunla & Enabulele 2010). Data collected were analysed by using PASW (Predictive Analytical Software), version 18.0, the premier vendor for (Statistical Package for the Social Sciences) program. A p-value of less than or equal to 0.05 was considered to be statistically significant ($p \leq 0.05$).

Results and Discussion

A total of 739 urine samples, 202 (27.3%) samples showed significant growth, whereas majority of the

samples i.e. 298 (40.3%) showed no growth, 216 showed non-significant growth, and out of the total only 23 samples showed mixed growths. *E. coli* (54.0%) was the most predominant uropathogen followed by CoNS (21.3%), *Enterococcus* spp. (7.3%), *Pseudomonas aeruginosa* (4.0%) and *Klebsiella* spp. (2.7%). *Acinetobacter* spp. and *Candida* spp. were 2.2% each. Other bacterial isolates were 1.0% or less than 1.0%. The distribution of *E. coli* were found to be the most frequent in age group 16-49 years in both sexes i.e. 13 and 60 in male and female respectively. In age group 1-15 years only two *E. coli* were isolated from females. The sexwise distribution showed that females (65.3%) were more susceptible to UTI than males (34.7%).

E. coli, the most common uropathogen isolated more commonly from the female patients comparatively to the male patients and isolation of *E. coli* among female patients is statistically significant ($p < 0.05$) whereas, isolate of CoNS was more common among male patient; however, isolation of CoNS more among male patients is statistically insignificant ($p > 0.05$) (Table 1).

Table 1. Distribution of uropathogens among males and females

Uropathogens	Male	Female	p- value
<i>Escherichia coli</i>	24(34.3)	85(64.4)	0.0001
Coagulase negative Staphylococcus (CoNS)	22(31.4)	21(15.9)	0.084
<i>Enterococcus</i> spp.	9(12.9)	7(5.3)	0.145
<i>Pseudomonas aeruginosa</i>	4(5.7)	6(4.5)	1.0
<i>Acinetobacter</i> spp.	1(1.4)	4(3.0)	0.654
<i>Candida</i> spp.	2(2.9)	3(2.3)	1.0
<i>Klebsiella</i> spp.	3(4.3)	3(2.3)	0.682
<i>Staphylococcus aureus</i>	2(2.9)	2(1.5)	0.643
<i>Citrobacter</i> spp.	2(2.9)	0(0.0)	0.149
<i>Enterobacter</i> spp.	0(0.0)	1(0.8)	1.0
<i>Salmonella</i> Typhi	1(1.4)	0(0.0)	0.387
Total	70(100.0)	132(100.0)	

Majority of *E. coli* showed susceptibility towards nitrofurantoin (94.5%) followed by ciprofloxacin and ofloxacin with the susceptibility of 50.5% for each drug. Cephalexin (7.3%) was found least effective drug followed by ampicillin (18.3%). Norfloxacin, nalidixic acid and co-trimoxazole were found effective only for less than half of the isolates of *E. coli* (Table 2).

Table 2. Antibiotic susceptibility pattern of *E. coli*

Antibiotics	Sensitive	Resistant
Ampicillin	20(18.3)	89(81.7)
Cephalexin	8(7.3)	101(92.7)
Nalidixic acid	23(21.1)	86(78.9)
Ciprofloxacin	55(50.5)	54(49.5)
Ofloxacin	55(50.5)	54(49.5)
Norfloxacin	51(46.8)	58(53.2)
Co-trimoxazole	50(45.9)	59(54.1)
Nitrofurantoin	103(94.5)	6(5.5)

Taking resistant to two or more classes of antibiotics as MDR, it was detected in 90.8% isolates. Among the

99 (90.8%) MDR strains, 21(19.3%) were resistant to two antibiotics and 78 (71.6%) were resistant to three or more antibiotics (Table 3).

Table 3. Multidrug resistance (MDR) pattern of *E. coli*

Bacterial isolate	No. of isolates	Resistance to antibiotic			Total	Percent
		0 Drug	1 Drug	2 Drug		
<i>Escherichia coli</i>	109	0	10	21	99	90.8

Sixteen multidrug resistance patterns were observed in *E. coli* for the eight antimicrobial agents tested. Resistance to Amp-Na-Cp-Cip-Of-Nx-Nit was the most frequent pattern observed in 30.3% of *E. coli* isolates, whereas Na-Cp-Cot, Amp-Na-Cp-Nx, Amp-Na-Cp-Cip-Nx, Na-Cip-Of-Nx-Cot and Amp-Na-Cp-Cip-Of-Nx-Nit were the least frequent pattern observed in 1.0% of *E. coli* for each (Table 4).

Table 4. Antibiotic resistance pattern of MDR *E. coli* strains

Antibiotic resistant pattern	Number (%)
Amp-Cp	13(13.1)
Na-Cp	4(4.0)
Na-Cot	4(4.0)
Amp-Cp-Cot	2(2.0)
Amp-Na-Cp	5(5.1)
Na-Cp-Cot	1(1.0)
Amp-Na-Cp-Nx	1(1.0)
Amp-Na-Cp-Cot	12(12.1)
Amp-Na-Cp-Nx-Cot	2(2.0)
Amp-Na-Cp-Cip-Nx	1(1.0)
Na-Cip-Of-Nx-Cot	1(1.0)
Amp-Na-Cp-Cip-Of-Nx	15(15.2)
Amp-Na-Cp-Of-Nx-Cot	2(2.0)
Amp-Na-Cp-Cip-Of-Nx-Cot	30(30.3)
Amp-Na-Cp-Cip-Of-Nx-Nit	1(1.0)
Amp-Na-Cp-Cip-Of-Nx-Nit-Cot	5(5.1)
Total MDR isolates	99(100.0)

Note: Amp-Ampicillin, Cp-Cephalexin, Na-Nalidixic acid, Cot-Co-trimoxazole, Nx-Norfloxacin, Cip-Ciprofloxacin, Of-Ofloxacin, Nit-Nitrofurantoin

Out of 109 *E. coli* isolates, only 10 showed MAR index of 0.1 i.e. these isolates were only resistant to one antibiotic. However, five isolates showed MAR index of 1 i.e. these isolates were resistant to all the antibiotics used in antibiotic susceptibility testing (Table 5).

Table 5. Multiple antibiotic resistance (MAR) indices of *E. coli*

MAR index	Frequency of MAR index <i>E. coli</i> (n=109)
0	0(0.0)
0.1	10(9.2)
0.2	21(19.3)
0.3	8(7.3)
0.4	0(0.0)
0.5	13(11.9)
0.6	4(3.7)
0.7	17(15.6)
0.8	31(28.4)
0.9	0(0.0)
1.0	5(4.6)

Our study was conducted among outpatients and inpatients suspected of urinary tract infection, attending Bir Hospital, Kathmandu. In this study, 27.3% urine specimens from suspected UTI patients gave significant growth. The results are in agreement with other investigators from Nepal (Chhetri *et al.* 2001, Karki *et al.* 2004, Kumari *et al.* 2005, Rai *et al.* 2008, Acharya *et al.* 2011) and rest of the world (Levitt 1993, Bashar *et al.* 2009). The majority of urine specimens showed no growth (40.3%). This may be due to patients undergoing antibiotics therapy which has inhibited or destroyed the bacterial growth (Okonofua & Okonofu 1989), or slow growing organisms, and which were not able to grow on the routine culture media used (Kattel *et al.* 2008). This study showed that the commonest isolates were *E. coli* (54%), CoNS (21.3%), *Enterococcus* spp. (7.3%), *Pseudomonas aeruginosa* (4.0%) and *Klebsiella* spp. (2.7%). In a study from Kathmandu, in combined group, outpatients as well as inpatients, *E. coli* (59.6%) was the commonest one followed by *Staphylococcus aureus* (12.5%), *Klebsiella* spp. (10.8), *Enterococcus* spp. (7.9%) and *Pseudomonas aeruginosa* (5.0%) (Kattel *et al.* 2008). *E. coli* was isolated as the most predominant isolate and accounted for 54.0% of the total uropathogens

and agreed with other studies (Chhetri *et al.* 2001, Dhakal *et al.* 2002, Jha & Bapat 2005, Das *et al.* 2006, Kattel *et al.* 2008) in Nepal. The results are also agreed with the study done at international context, which indicated that the Gram negative bacteria mostly *E. coli* was the commonest one isolated in patients with UTI (Tambekar *et al.* 2006, Ahmed & Imran 2008, Biadlegne & Abera 2009, Omigie *et al.* 2009, Manjunath *et al.* 2011, Alzohairy & Khadri 2011).

E. coli, the most common uropathogen isolated more commonly from female patients comparatively to the male patients and isolation of *E. coli* more among female patients is statistically significant ($p < 0.05$) whereas, isolate of CoNS was more common among male patient; however, isolation of CoNS more among male patients is statistically insignificant ($p > 0.05$). The reason for the higher prevalence of CoNS in males is not clear, though lack of circumcision, receptive anal intercourse and HIV infection recognised as risk factor for males (Orrett & Davis 2006). The study revealed that females (65.3%) were more susceptible to UTI than males (34.7%), which is also similar to other studies (Dimitrov *et al.* 2003, Akram *et al.* 2007, Arjunan *et al.* 2010, Manjunath *et al.* 2011, Alzohairy & Khadri 2011, Acharya *et al.* 2011). The increased incidence of the urinary tract infection in women is conditioned by favouring anatomic factors, by hormonal changes and by the urodynamic disturbance occurring with age (Bobos *et al.* 2010).

Most *E. coli* isolates in our study were resistant to ampicillin (81.7%) which resembles other studies (Rashedmarandi *et al.* 2008, Behroozi *et al.* 2010, Farshad *et al.* 2010a, Farshad *et al.* 2011). Overall quinolone resistance of *E. coli* were 57.7%. Increased resistance in quinolones against *E. coli* may reflect the overuse of these drugs for the treatment of UTI (Saleh *et al.* 2009). Another factor could be the generalized use of fluoroquinolones in animals feed (especially in poultry), and the subsequent transmission of resistant to strains from animals to humans (Miller & Tang 2004). In our study, only 5.5% isolates were resistant to nitrofurantoin. These findings are in agreement in with other workers (Moniri *et al.* 2003, Bean *et al.* 2008, Biadlegne & Abera 2009, Raza *et al.* 2011, Bahadin *et al.* 2011, Baral *et al.* 2012). Nitrofurantoin was also found as the most effective antimicrobial in UTI caused by *E. coli* from studies in Nepal (Karki *et al.* 2004, Shrestha *et al.* 2007, Sharma

et al. 2011, Rijal *et al.* 2012) and rest of the world (Sahm *et al.* 2001, Jamie *et al.* 2002, Alos *et al.* 2004, Okonko *et al.* 2009, Behroozi *et al.* 2010, Eryilmaz *et al.* 2010). However, Akram *et al.* (2007), Kausar *et al.* (2009), and Arjunan *et al.* (2010) were reported higher rate of *E. coli* resistance to nitrofurantoin. In case of the present study, nitrofurantoin was found to be the most effective antimicrobial. Resistance to nitrofurantoin among *E. coli* isolates from UTIs remained low despite of more than 50 year's widespread use of the drug (Kahlmeter 2000, Mazzulli *et al.* 2001). Reason for the lack of emerging resistance are not fully understood, but likely include restricting use to indication for urinary infection, limited systemic absorption, and the need for multiple genetic mutations for the bacteria to develop resistance (Nicolle *et al.* 2006). Most of the *E. coli* isolates showed the multidrug resistant (90.8%) in agreement with other studies that found multidrug resistant *E. coli* ranging from 67.0 to 100.0% (Bashar *et al.* 2009, Moyo *et al.* 2010, Farshad *et al.* 2010a, Hassan *et al.* 2011). The higher MDR may be due to large portion of *E. coli* isolates being previously exposed to several antibiotics. In this study the antimicrobial agents showed that *E. coli* were highly resistant to commonly used antibiotics i.e. cephalexin, ampicillin, nalidixic acid, ciprofloxacin, ofloxacin, norfloxacin, and co-trimoxazole. The resistance rate of *E. coli* to commonly used antibiotics was: ampicillin (81.7%), cephalexin (92.7%), nalidixic acid (78.9%), ciprofloxacin (49.5%), ofloxacin (49.5%), norfloxacin (53.2%), co-trimoxazole (54.1%) and nitrofurantoin (5.5%). The increasing resistance of co-trimoxazole to *E. coli* has been reported in other studies from Nepal and other countries (Kattel *et al.* 2008, Rai *et al.* 2008, Bashar *et al.* 2009, Biadlegne & Abera 2009, Abou-Dobara *et al.* 2010, Jadhav *et al.* 2011). They were higher than the rate reported in our study. Ampicillin resistance among *E. coli* was 81.7%, which is comparable with other studies (Rafique *et al.* 2002, Biadlegne & Abera 2009, Behroozi *et al.* 2011, Alzohairy & Khadri 2011). On the basis of our finding, antimicrobials such as ampicillin and co-trimoxazole should no longer be recommended for initial empirical therapies for UTIs.

Multiple antibiotic resistance (MAR) index is a tool that reveals the spread of bacterial resistance in a given population (Krumpermann 1983). An MAR index greater than 0.2 implies that the strains of such bacteria originate from an environment where several

antibiotics are used (Ehinmidu 2003). The MAR indices of *E. coli* obtained in this study is a possible indication that a very large proportion of the bacterial isolates have been exposed to several antibiotics.

Higher resistance rate to all antibiotics used in this study except nitrofurantoin may be explained by high and uncontrolled use of these antibiotics in our institutions.

Most *E. coli* isolates are highly resistant to commonly prescribed antibiotics (ampicillin, cephalexin, quinolones and co-trimoxazole), but are still susceptible to nitrofurantoin which should be considered as preferred therapeutic agent once the organism is identified.

Acknowledgements

We greatly acknowledge the support provided by Bir Hospital and the Central Department of Microbiology, Tribhuvan University for conducting this research.

References

- Abou-Dobara, M.I., M.A. Deyab, E.M. Elsayy and H.H. Mohamed. 2010. Antibiotic susceptibility and genotype patterns of *Escherichia coli*, *Klebsiella pneumonia* and *Pseudomonas aeruginosa* isolated from urinary tract infected patients. *Pol. J. Microbiol.* **59** (3): 207-212.
- Acharya, A., R. Gautam and L. Subedee. 2011. Uropathogens and their antimicrobial susceptibility pattern in Bharatpur. *Nepal Med. Coll. J.* **13**(1): 30-33.
- Ahmed, K. and Imran. 2008. Prevalence and antibiogram of uncomplicated lower urinary tract infections in human population of Gilgit, Northern areas of Pakistan. *Pakistan J. Zool.* **40**(4):295-301.
- Akinjogunla, O.J. and I.O. Enabulele. 2010. Virulence factors, plasmid profiling and curing analysis of multidrug resistant *Staphylococcus aureus* and coagulase negative *Staphylococcus* spp. isolated from patients with acute otitis media. *Journal of American science* **6**(11): 1022-1033.
- Akram, M., M. Shahid and A.U. Khan. 2007. Etiology and antibiotic resistance patterns of community-acquired urinary tract infections in JNMC hospital Aligarh, India. *Ann. Clin. Microbiol. Antimicrob.* **6**: 4.
- Alos, J.I., M.G. Serrano and J.L. Gomez-Garces. 2004. Antibiotic resistance of *Escherichia coli* from community-acquired urinary tract infections in relation to demographic and clinical data. *Clin. Microbiol. Infect.* **11**: 199-203
- Alzohairy, M. and H. Khadri. 2011. Frequency and antibiotic susceptibility pattern of uro-pathogens isolated from community and hospital –acquired infections in Saudi Arabia- A prospective case study. *Br. J. Med. Res.* **1**(2): 45-56.
- Arjunan, M., A.A. Al-Salamah and M. Amuthan. 2010. Prevalence and antibiotic susceptibility of uropathogens in patients from a rural environment, Tamilnadu. *American Journal of Infectious Disease* **6**(2): 29-33.
- Bahadin, J., S.S.H.Teo and S. Methew. 2011. Aetiology of community-acquired urinary tract infection and antimicrobial susceptibility patterns of uropathogens isolated. *Singapore Med. J.* **52**(6): 415-420.
- Baral, P., S. Neupane, B.P. Marasini, K.R. Ghimire, B. Lekhak and B. Shrestha. 2012. High prevalence of multidrug resistance in bacterial uropathogens from Kathmandu, Nepal. *BMC Res. Notes.* **5**:38.
- Bashar, M.A., M.F. Ahmed, S.R. Rahman and D.J. Gomes. 2009. Distribution and resistance trends of *Escherichia coli* from urinary tract infection isolated in Dhaka city. *Bangladesh J. Med. Sci.* **15**(2): 93-98.
- Bean, D.C., D. Krahe and D.W. Wareham. 2008. Antimicrobial resistance in community and nosocomial *Escherichia coli* urinary tract isolates, London 2005-2006. *Ann. Clin. Microbiol. Antimicrob.* **7**: 13.
- Behroozi, A., M. Rahbar and J.A. Yousefi. 2010. A survey on epidemiology of urinary tract infections and resistance pattern of uropathogens in an Iranian 1000-bed tertiary care hospital. *Afr. J. Microbiol. Res.* **4**(9): 753-756.
- Biadlegne, F. and B. Abera. 2009. Antimicrobial resistance of bacterial isolates from urinary tract infections at Felge Hiwot Referral Hospital, Ethiopia. *Ethiop. J. Health Dev.* **23**(3): 236-238.
- Bobos, C., C. Hodarnau, D. Terec, L. Feticu, F. Iencica and H. Alina. 2010. Prevalence and susceptibility to chemotherapeutic agents of bacterial species isolated from urinary tract infections. *Clujul Medical.* **83**(1): 69-75.
- Chhetri, P.K., S.K. Rai and U.N. Pathak *et al.* 2001. Retrospective study on urinary tract infection at Nepal Medical College Teaching Hospital, Kathmandu. *Nepal Med. Coll. J* **3**: 83-85.
- CLSI. 2011. Performance standards for antimicrobial susceptibility testing; Twenty-first Informational Supplement. CLSI M100-S21:**31**(1).Clinical and Laboratory Standards Institute, Wayne, PA.
- Das, R.N., T.S. Chandrashekhar, H.S. Joshi, M. Gurung, N. Shrestha and P.G. Shivananda. 2006. Frequency and susceptibility profile of pathogens causing urinary tract infections at a tertiary care hospital in Western Nepal. *Singapore Med. J.* **47**(4): 281-285.
- DoHS. 2012. Annual report 2010/2011. Department of Health Services (DoHS), Ministry of Health,

- Government of Nepal, Teku, Kathmandu, Nepal, 301 pp.
- Dhakal B.K., B.M. Pokhrel and J. Ahnn. 2002. Microscopic detection of urinary tract infection in Nepalese patients. *J. Microbiol.* **40**(4): 267-273.
- Dimitrov, T.S., E.E. Udo, M. Emara, F. Awni and R. Passadilla. 2003. Etiology and antibiotic susceptibility patterns of community-acquired urinary tract infections in a Kuwait hospital. *Med Princ Pract.* **13**(6): 334-339.
- Dromigny, J.A., P. Nabeth, A. Juergens-Behr and J.D. Perrier-Gros-Claude. 2005. Risk factors for antibiotic-resistant *Escherichia coli* isolated from community-acquired urinary tract infections in Dakar, Senegal. *J. Antimicrob. Chemother.* **56**(1): 236-239.
- Ehinmidu J.O. 2003. Antibiotic susceptibility patterns of urine bacterial isolates in Zaria, Nigeria. *Trop. J. Pharm. Res.* **2**(2): 223-228.
- Eryilmaz M., M.E. Bozkurt, M. Murat, M.M. Yildiz and A. Akin. 2010. Antimicrobial resistance of urinary *Escherichia coli* isolates. *Trop. J. Pharm. Res.* **9**(2): 205-209.
- Farshad, S., F. Emamghoraishi and A. Japoni. 2010a. Association of virulent genes hly, sfa, cnf-1 and pap with antibiotic sensitivity in *Escherichia coli* strains isolated from children with community-acquired UTI. *Iran Red Crescent Med. J.* **12**(1): 33-37.
- Farshad, S., M. Anvarinejad and A.M. Tavana. 2011. Molecular epidemiology of *Escherichia coli* strains isolated from children with community acquired urinary tract infection. *African Journal of Medical Research* **5**(26): 4476-4483.
- Farshad, S., R. Ranjbar, M. Anvarinejad, M.A. Shahidi and M. Hosseini. 2010b. Emergence of Multi Drug Resistant Strains of *Escherichia coli* isolated from Urinary Tract Infection. *Open Conf. Proc. J.* **1**: 192-196.
- Gonzalez, C.M. and A.J. Schaeffer. 1999. Treatment of urinary tract infection: what's old, what's new, and what works. *World J. Urol.* **17**: 372-382.
- Hassan, S.A., S.A. Jamal and M. Kamal. 2011. Occurrence of multidrug resistant and ESBL producing *E. coli* causing urinary tract infections. *J. Basic Appl. Sci.* **7**(1): 39-43.
- Jadhav, S., A. Hussain, S. Devi, A. Kumar, S. Parveen, N. Gandham, L.H. Wieler, C. Ewers and N. Ahmed. 2011. Virulence characteristics and genetic affinities of multiple resistant uropathogenic *Escherichia coli* from a semi-urban locality in India. *PLoS One* **6**(3): e18063.
- Jamie, W.E., R.K. Edwards and P. Duff. 2002. Antimicrobial susceptibility of gram-negative uropathogens. *Infect. Dis. Obstet. Gynecol.* **10**: 123-126.
- Jha, N. and S.K. Bapat. 2005. A study of sensitivity and resistance of pathogenic microorganisms causing UTI in Kathmandu valley. *J Med.* **3**: 123-129.
- Kahlmeter, G. 2000. The ECO*SENS Project: a prospective, multinational, multicentre epi-demiological survey of the prevalence and antimicrobial susceptibility of urinary tract pathogens-interim report. *J. Antimicrob. Chemother.* **46**(Suppl A): 15-22.
- Karki, A., B.R. Tiwari, and S.B. Pradhan. 2004. Study of bacteria isolated from urinary tract infection and their sensitivity pattern. *J. Nepal. Med. Assoc.* **43**: 200-204
- Kattel, H.P., J. Acharya, S.K. Mishra, B.P. Rijal and B.M. Pokhrel. 2008. Bacteriology of urinary tract infection among patients attending Tribhuvan University Teaching Hospital, Kathmandu, Nepal. *JNAMLS* **9**(1): 25-29.
- Kausar, Y., S.K. Chunchanur, S.D. Nadagir, L.H. Halesh and M.R. Chandrasekhar. 2009. Virulence factors, serotypes and antimicrobial susceptibility pattern of *Escherichia coli* in urinary tract infections. *Al. Ameen. J. Med. Sci.* **2**(1): 47-51.
- Krumpermann, P.H. 1983. Multiple antibiotics resistance indexing of *E. coli* to identify high risks sources of fecal contamination of foods. *App. Environ. Microbiol.* **46**: 165-170.
- Kumari, N., G. Ghimire, J.K. Magar, T.M. Mohapatra, and A. Rai. 2005. Antibiogram pattern of isolates from UTI cases in Eastern part of Nepal. *Nepal Med. Coll. J.* **7**: 116-118.
- Levitt, P.N. 1993. Analysis of pathogens isolated from urinary tract infection in Barbados. *West Indi. Med. J.* **42**: 72-76.
- Manjunath, G.N., R. Prakash, A. Vamseedhar and K. Shetty. 2011. Changing trends in the spectrum of antimicrobial drug resistance pattern of uropathogens isolated from hospital and community patients with urinary tract infections in Tumkur and Bangalore. *Int. J. Biol. Med. Res.* **2**(2): 504-507.
- Mazzulli ,T., M. Skulnick, G. Small, W. Marshall, D.J. Hoban, G.G. Zhanel, S. Finn and D.E. Low 2001. Susceptibility of community Gram-negative urinary tract isolates to mecillinam and other oral agents. *Can. J. Infect. Dis.* **12**(5): 289-292.
- Miller, L.G. and A.W. Tang. 2004. Treatment of uncomplicated urinary tract infections in an era of increasing antimicrobial resistance. *Mayo Clin. Proc.* **79**: 1048-1054.
- Moniri, R., A. Khorshidi, and H. Akbari. 2003. Emergence of multidrug resistant strains of *Escherichia coli* isolated from urinary tract infection. *Iranian J. Publ. Health* **32**(4): 42-46.
- Moyo, S.J., S. Aboud, Kasubi M, Lyamuya E.F. and S.Y. Maselle. 2010. Antimicrobial resistance among procedures and non-producers of extended spectrum beta-lactamase in urinary isolates at a tertiary hospital in Tanzania. *BMC Research Notes* **3**: 348.
- Nicolle, L., P.A.M. Anderson, J. Conly, T.C. Mainprize, J. Meuser, J.C. Nickel, V.M. Senikas and G.G. Zhanel.

2006. Uncomplicated urinary tract infection in women. *Can. Fam. Physician.* **52**(5): 612-618.
- Okonko, I.O., O.B. Donbraye and L.A. Ijandipe. 2009. Antibiotic sensitivity and resistance patterns of uropathogens to nitrofurantoin and nalidixic acid in pregnant women with urinary tract infections in Inbadan, Nigeria. *Middle East Journal of Scientific Research* **4**(2):105-109.
- Okonofua, E.E. and B.N. Okonofu. 1989. Incidence and pattern of asymptomatic bacteriuria of pregnancies in Nigerian women. *Nig. Med. Pract.* **17**: 354-358.
- Omigie, O., L. Okoror, P. Umolu, and I.G. Kuuh. 2009. Increasing resistance to quinolones: A four-year prospective study of urinary tract infection pathogen. *Int. J. Gen. Med.* **2**:171-175.
- Orrett, F.A., and G.K. Davis. 2006. A comparison of the antimicrobial susceptibility profile of urinary pathogens for the years 1999 and 2003. *West Indian Med. J.* **55**: 95-99.
- Rafique, S., A. Mehmood, M. Qayyum and A.B. Qazilbash. 2002. Prevalence patterns of community-based and nosocomial urinary tract infection caused by *Escherichia coli*. *Pakistan Journal of Biological Sciences* **5**(4): 494-496.
- Rai, G.K., H.C. Upreti, S.K. Rai, K.P. Shah and R.M. Shrestha. 2008. Causative agents of urinary tract infections in children and their antibiotic sensitivity pattern: a hospital based study. *Nepal Med. Coll. J.* **10**(2): 86-90.
- Raksha, R., H. Srinivasa, and R.S. Macaden. 2003. Occurrence and characterization of uropathogenic *Escherichia coli* in urinary tract infections. *Indian J. Med. Microbiol.* **21**(2): 102-107.
- Ramesh, N., C.S. Sumathi, V. Balasubramanian, K.R. Palaniappan and V.R. Kannan. 2008. Urinary tract infection and antimicrobial susceptibility pattern of extended spectrum of beta lactamase producing clinical isolates. *Advan Biol Res.* **2**(5-6): 78-82.
- Rashedmarandi, F., M. Rahnamayefarzami, M. Saremi and R. Sabouri 2008. A survey on urinary pathogens and their antimicrobial susceptibility among patients with significant bacteriuria. *Iran J Pathol.* **14**: 191-196.
- Raza, S., S. Pandey and C.P. Bhatt. 2011. Microbiological analysis of the urine isolates in Kathmandu Medical College Teaching Hospital, Kathmandu, Nepal. 2011. *Kathmandu Univ. Med. J.* **36**(4): 295-297.
- Rijal, A., G. Ghimire, K. Gautam and A. Barakoti. 2012. Antibiotic susceptibility of organism causing urinary tract infection in patients presenting to a teaching hospital. *J. Nepal Health Res. Counc.* **10**(20): 24-27.
- Sahm, D.F., C. Thornsberry, D.C. Mayfield, M.E. Jones and J.A. Karlowsky. 2001. Multidrug resistant urinary tract isolates of *Escherichia coli*: Prevalence and patients demographics in the United States in 2000. *Antimicrob. Agents Chemother.* **45**(5): 1402-1406.
- Saleh, A.A., S.S. Ahmed, M. Ahmed, A.N.I. Sattar and Md.R.A. Miah 2009. Changing trends in uropathogens and their antimicrobial sensitivity pattern. *Bangladesh J Med Microbiol.* **3**(1): 9-12.
- Sharma, A., S. Shrestha, S. Upadhyay and P. Rijal 2011. Clinical and bacteriological profile of urinary tract infection in children at Nepal Medical College Teaching Hospital. *Nepal Med. Coll. J.* **13**(1): 24-26.
- Shrestha P., S. Malla, S. Basnvat, S. Dumre, B. Upadhaya and S. Lamichhane. 2007. Antimicrobial resistance pattern of bacterial isolates causing urinary tract infection. *J. Nepal Health Res. Counc.* **5**(2): 49-54.
- Tambekar, D.H., D.V. Dhanorkar, S.R. Gulhane, V.K. Khandelwal and M.N. Dudhane. 2006. Antibacterial susceptibility of some urinary tract pathogens to commonly used antibiotic. *Afr. J. Biotechnol.* **5**(17): 1562-1565.

