

Phytochemical Screening, Antioxidant and Antidiabetic Activities of Extracts of Leaves and Seeds of *Carica papaya*

Shanta Pokhrel^{1*} and Puja Karki¹

¹Department of Chemistry, Tri-Chandra Multiple Campus, Tribhuvan University, Nepal

*CORRESPONDING AUTHOR:

Shanta Pokhrel

Email: shantabhattacharai2014@gmail.com

ISSN : 2382-5359(Online),
1994-1412(Print)

DOI:

<https://doi.org/10.3126/njst.v20i1.43362>

ACCESS THE ARTICLE ONLINE



CONFLICT OF INTEREST: None

Copyright: The Author(s) 2021. This is an open access article under the [CC BY](https://creativecommons.org/licenses/by-nc/4.0/) license.



ABSTRACT

The phytochemicals, phenolic, flavonoid contents, and antibacterial, antioxidant and antidiabetic activities of leaves and seeds extract of *Carica papaya* via Soxhlet extraction were investigated. The phytochemical screening indicated different bioactive compounds such as saponins, alkaloids, glycosides, tannins, polyphenols, steroids, and flavonoids in different extracts. The total phenolic content (TPC) was determined spectrophotometrically by Folin–Ciocalteu reagent method and aluminium chloride reagent to measure total flavonoid content (TFC). TPC was found more in leaf methanol extract (LME) (93.18 ± 0.40 mg GAE/g) than that in seed methanol extract (SME) (89.14 ± 0.45 mg GAE/g). Comparatively, the flavonoid content was found more in LME (18.85 ± 0.061 mgQE/g) than in SME (16.64 ± 0.065 mgQE/g). The antioxidant activity was assessed for LME and SME by DPPH radical scavenging activity and IC_{50} values. The IC_{50} values of LME and SME were 43.54 ± 0.007 and 15.48 ± 0.13 μ g/mL, respectively. Thus, seeds methanol extract showed better antioxidant activity than leaves methanol extract. The antidiabetic assay was performed via α -amylase inhibition method. Among the selected parts (seeds and leaves), methanol seed extract exhibited high α -amylase inhibition with an IC_{50} value of 46.99 ± 0.018 μ g/mL. The study explored the potential value of *Carica papaya* in medicinal applications.

Keywords: α -Amylase inhibition, IC_{50} , Total flavonoid content, Total phenolic content

1. INTRODUCTION

Carica papaya, locally known as Pawpaw belongs to Caricaceae family, is an evergreen shrub. It is a large tree with a solo stem growing in height upto 10 m tall, with spirally arranged leaves confined to the trunk's top. The lower trunk contains leaves and fruits. The leaves are large, 50-70

cm in diameter (Aravind *et al.* 2013). *Carica papaya* is a nutraceutical plant having a wide range of pharmacological activities (Peter *et al.* 2014). Its fruit is a rich source of nutrients but papaya leaves and seeds also contain various phytochemicals, including phenols. The edible black seeds of the papaya with sharp and spicy tests are ground and used as a substitute for black pepper (Nna *et al.* 2019). However, the seeds seem to have more potent medicinal values than the flesh (Peter *et al.* 2014; Nna *et al.* 2019). *Carica papaya* seeds possess various pharmacological activities such as antifertility, contraceptive, anthelmintic, anti-inflammatory, analgesic and antimicrobial property (Agarwal *et al.* 2016). Other pharmacological uses of papaya seeds include carminative, emmenagogue, abortifacient, counterirritant, paste in psoriasis, ringworm disease, and an antifertility agent in males (Peter *et al.* 2014). Water extract of Papaya seeds protects fibroblasts from H_2O_2 , induced stress due to the antioxidant activity (Panzarini *et al.* 2014). *Carica papaya* seeds are also used in the treatment of hypertension, diabetes mellitus and hypercholesterolemia. The vegetative parts of the papaya plant have enormous medicinal values (Asghar *et al.* 2016). Papaya leaves are made into tea as a treatment for malaria. The antimalarial and antiplasmodial activity has been noted due to the presence of Karpain compound, which kills microorganisms that often interfere with the digestive function (Udoh *et al.* 2005, Kousika & Priyadharshini 2020). The leaves of *Carica papaya* are also used traditionally to cure various ailments like malaria, dengue, jaundice, immunomodulatory, and antiviral activity. Other diseases that can be controlled by *Carica papaya* include; abdominal discomfort, pain, diabetes, obesity, infection, and oral drug poisonings. It also possesses therapeutic, anti-inflammatory, and sugar reducing tendency (Ogundele *et al.* 2017). Therefore, the present study was carried out for the comparative study extracts of seeds and leaf's phytoconstituents (quantitative & qualitative analysis) and their biological activity, i.e. antibacterial, antioxidant and antidiabetic.

2. METHODOLOGY

2.1 Collection of Plant Materials and Preparation of Seed and Leaves Extracts

The leaves and fruits of *Carica papaya* were collected from Suklagandaki-3, Tanahun district Gandaki zone of Nepal at the beginning of April 2019. The dried and powdered form (50 g) of leaves and seeds were extracted successively with hexane (350 mL), chloroform (300 mL), ethylacetate (250 mL), methanol (200 mL) by Soxhlet method. The remaining residue after extraction with methanol was refluxed with water (500 mL) for 3h. The extracts were filtered and concentrated using Rota vapour to get solid or semi-solid mass and kept in the freezer for further analysis.

2.2 Phytochemical Screening

Preliminary phytochemical screening of different extracts of leaves and seeds were done using the standard procedure put forward by Ciulei I (Vongsak *et al.* 2013).

2.3 Quantitative Analysis of Extracts of Leaves and Seeds of *Carica papaya*

2.3.1 Total Phenolic Content (TPC)

The total phenolic content in plant extracts was determined using the Folin-Ciocalteu colourimetric method (Waterhouse 2002). Various gallic acid solutions in methanol (10, 25, 50, and 100 $\mu\text{g/mL}$) were prepared. In a 20 mL test tube, 1 mL of gallic acid of each concentration was added to that 5 mL of 10% FCR (1:10 v/v), and 4 mL of 7% Na_2CO_3 were added to get a total volume of 10 mL. The blue coloured mixture was shaken well and incubated for 30 minutes at 40 °C in a water bath. Then, the absorbance was measured at 760 nm. Similarly, the absorbance was measured against a blank containing all reagents except gallic acid. All experiments were carried out in triplicate. The average absorbance values obtained at different concentration was used to plot the calibration curve.

Absorbance for various extracts (10, 25, 50, 80 and 100 $\mu\text{g/mL}$) was recorded by following the standard procedure. Total phenolic content in the extracts of leaves and seeds was calculated as mg gallic acid equivalents (GAE) per gram dry extract (mg/g).

2.3.2 Total Flavonoid Content (TFC)

The total flavonoid present in the different plants' methanolic extract was estimated by the aluminium chloride colourimetric assay involving quercetin as standard (Pallab *et al.* 2011; Pandey & Rajbhandari 2014). The absorbance values for different concentration of each extract were recorded at 510 nm. The total flavonoid content in different extract was calculated from the calibration curve using regression equation $y = 0.0113x - 0.0837$, $R^2 = 0.988$ followed by the formula $C = cV/m$ expressed as mg quercetin equivalents per gram of extract in dry weight (mg QE/g) (Ariffin *et al.* 2021). The various standard quercetin concentrations (10, 25, 50, 100 and 125 $\mu\text{g/mL}$) were prepared. 1.0 mL of quercetin was taken from each concentration, and 2.0 mL of methanol and 4.0 mL of distilled water were added. At the zero time, 0.3 mL of 5 % sodium nitrite solution was added, and 0.3 mL of 10% AlCl_3 was added after 5 min, and 2.0 mL of 1M NaOH solution was added after 6 min to the mixture. Immediately, the total volume of the mixture was made 10.0 mL by adding the distilled water. Finally, pink colour mixture was recorded at 510 nm against a blank containing all reagents except quercetin. The average absorbance values obtained at different concentrations of quercetin were used to plot the calibration curve. Various concentration of seeds and leaves extract (10, 25, 50, 100 and 125 $\mu\text{g/mL}$) were prepared. Following the procedure described for standard quercetin, absorbance for each concentration of the extract was measured. Total Flavonoid content in extract was expressed as mg quercetin equivalent (QE) per gram of dry extract (mg/g).

Total phenolic and flavonoid content were calculated by using the formula given by Zuhair *et al.* (2013): $C = cV/m$ where, C = total phenolic/flavonoid compound in mg QE/g dry extract, c = concentration of gallic acid/quercetin established from calibration curve in mg/mL, V = volume of extract in mL, m = weight of extract in gram (Khanal *et al.* 2020). Data were recorded as a mean \pm standard deviation of three determination of Absorbance for each concentration, from which the linear correlation coefficient (R^2) value was calculated using MS Office Excel 2007. The linear regression equation for a straight line

is, $y = mx + c$ where, y = absorbance of extract, m = slope of the calibration curve, x = concentration of extract, c = intercept. Using this regression equation, concentrations of each extract, the phenolics and flavonoid content were calculated.

2.4 Biological Screening

2.4.1 Antioxidant activity using 2,2-Diphenyl-1-picrylhydrazyl (DPPH) radical scavenging assay

Antioxidant activity of the selected extracts was assessed using DPPH free radical (Jamuna *et al.* 2012; Pokhrel & Neupane 2021). DPPH solution (0.2 mM) was prepared by dissolving 7.8 mg of DPPH in 100 mL methanol. Different concentrations (20, 40, 60, 80 and 100 $\mu\text{g/mL}$) of methanolic solutions of each extract were prepared by serial dilution of the stock solution of the respective extract. To each 0.5 mL extract solution, 2.5 mL DPPH (0.2 mM) solution was added. A control was prepared by mixing 0.5 mL distilled water and 2.5 mL DPPH (0.2 mM) methanolic solution. These samples were shaken well, incubated at 37 °C for half an hour, and the absorbance was recorded at 517 nm by UV-Vis spectrophotometer (Hitachi U2800). The radical scavenging activity was expressed as the radical scavenging percentage using the following formula (Mishra *et al.* 2012; Dhital 2017):

$$\text{DPPH Inhibition (\%)} = [1 - A_1/A_0] \times 100$$

Where, A_1 = Absorbance of the sample, and A_0 = Absorbance of control

The assay was replicated thrice, and the result was taken as mean \pm standard deviation. The IC_{50} (50% inhibitory concentration) value is the effective concentration of the sample required to scavenge 50% of the DPPH free radicals. It was calculated using the inhibition curve by plotting extract concentration versus the corresponding scavenging effect (Pokhrel & Neupane 2021).

2.4.2 α -Amylase Antidiabetic Assay

α -amylase inhibition assay was carried out to determine the antidiabetic potential of plants. The undigested starch due to enzyme inhibition was detected through the blue starch iodine complex detected at 630 nm. The starch solution

was prepared by dissolving 0.2 g starch in 25 mL of NaOH (0.4 M), and pH was maintained to 7.0, and the final volume made up to 100 mL by adding distilled water. Acarbose was used as a standard. 400 μ L of starch was incubated at 37 $^{\circ}$ C for 5 min with 200 μ L of acarbose or plant extract at different concentrations (40, 60, 80, 160, 320 and 640 μ g/mL), followed by 200 μ L of α -amylase (20 mM phosphate buffer with 6.7 mM NaCl, pH 6.9), and incubation at 37 $^{\circ}$ C for 15 min. Termination of the reaction was done by adding 800 μ L of HCl (0.1 M). Then, 1000 μ L of iodine reagent (2.5 mM) was added, and absorbance was measured at 630 nm. The percentage of enzyme inhibition was calculated by using the following equation (Chakrabarti *et al.* 2014);

$$\% \text{ inhibition} = [1 - (\text{Abs}_2 - \text{Abs}_1 / \text{Abs}_4 - \text{Abs}_3)] \times 100$$

Where, Abs1 = Absorbance of an incubated mixture containing plant extract, starch and amylase, Abs2 = Absorbance of an incubated mixture containing plant extract and starch. Abs3 = Absorbance of an incubated mixture containing starch and amylase Abs4 = Absorbance of an incubated mixture containing starch only. A graph was plotted by taking the concentration on the X-axis and % inhibition on the Y-axis. From this graph, each extract's IC₅₀ value was calculated, and the values of different extracts were compared. The extract having the lowest IC₅₀ is considered to have the best inhibition property.

3. RESULTS AND DISCUSSION

3.1 Phytochemical Screening

The results of the phytochemical screening of *Carica papaya* leaves and seeds extracts are shown in Table 1. Phytochemical screening indicated alkaloids, phenols, glycoside, quinones, saponins, tannins, and steroids in *Carica papaya* seeds and leaves' extracts. Flavonoids are present only in leaves extract. Among phytochemicals, alkaloids are significant for the protection and survival of plants because they ensure their survival against microorganisms (antibacterial activities). Phenolic compounds are a large and complex group of chemical constituents found in the plant (de Beer *et al.* 2002). They have an essential role as defense compound. The best-described property of flavonoids is their capacity to act as antioxidants to protect the human body from free radicals and reactive oxygen species. Recently, tannins have attracted scientific interest due to increased AIDS and various cancers (Saxena *et al.* 2013). The phytochemical analysis revealed the presence of many phytoconstituents (Peter *et al.* 2014; Dwivedi *et al.* 2020) which indicated that it has some medicinal potential (Pokhrel & Chaulagain 2019). Phytochemical screening revealed that the methanol extract of *Carica papaya* flowers contains alkaloids, flavonoids, saponins, and tannins as major components; saponins and tannins were present in chloroform and n-hexane extracts; however, steroids and flavonoids were additionally found in n-hexane extract (Dwivedi *et al.* 2020).

Table 1. Qualitative screening of *Carica papaya* leaves and seeds

Leaves Extract (LE)						
Phytochemicals	Method	Hexane	Ethylacetate	Chloroform	Methanol	Water
	Mayer's Test	-	-	-	+	+
Alkaloids	Dragendorff's Test	-	-	-	+	+
Phenols		-	-	-	+	+
Polyphenols	FeCl ₃ Test	-	-	-	+	+
Flavonoids	Alkaline Reagent Test	+	+	+	+	+

Reducing sugar	Fehling's Test	+	+	+	+	+
Glycosides	Molisch's Test	+	+	+	+	+
Quinones	-	-	-	+	+	+
Saponins	Chloroform and H ₂ SO ₄ Test	+	+	+	+	+
Tannins	Lead Acetate Test	+	+	+	+	+
Steroids	Liebermann-Burchard Test	+	+	+	+	+
	Salkowski Test	+	+	+	+	+

Seeds Extract (SE)

Phytochemicals	Method	Hexane	Ethyl-acetate	Chloroform	Methanol	Water
	Mayer's Test	+	+	+	+	+
Alkaloids	Dragendorff's Test	+	+	+	+	+
Phenols		+	+	+	+	+
Polyphenols	FeCl ₃ Test	+	+	+	+	+
Flavonoids	Alkaline Reagent Test	+	+	-	+	-
	Lead Acetate Test	+	+	-	+	-
Reducing sugar	Fehling's Test	-	-	+	+	+
Glycosides	Molisch's Test	+	+	-	+	-
Quinones	-	-	-	+	+	+
Saponins	Chloroform and H ₂ SO ₄ Test	+	+	+	+	+
Tannins	Lead Acetate Test	+	+	+	+	+
	Liebermann Burchard Test	-	-	+	+	+
Steroids	Salkowski Test	-	-	+	+	+

3.2 Quantitative Analysis

3.2.1 Total Phenolic Content (TPC)

The quantitative assessment of total phenol was done using Folin-Ciocalteu reagent (FCR) in terms of gallic acid equivalent. The absorbance values

obtained at 760 nm for different concentrations of gallic acid were used to construct the calibration curve. The obtained TPC values of methanol leaves and seeds extracts are shown in Table 2. TPC contents of methanol leaf and seed extract showed 93.18±0.40 and 84.14±0.45 mgGAE/g,

respectively. So, the leaf extract has high phenolic content than that of seed extract. However, Asghar *et al.* (2016) reported the phenolic content of methanol extract of leaves and the seeds was 54.28 ± 0.10 and 38.86 ± 0.82 mgGAE/g respectively whereas, Vuong *et al.* (2013) and Gogna *et al.* (2015) reported 63.59 ± 0.62 and 97.08 ± 2.8 mgGAE/g TPC content in leaves and seed extracts of *C. papaya*, respectively. The total phenolic content in our sample is relatively

different when compared with other studies. This variation is due to different extraction parameters, including temperature and extraction duration (Vuong *et al.* 2013). The result also indicated that the leaves contain high phenolic content that may provide an excellent dietary antioxidant source. The phenolic compounds are the main micro constituents contributing to the antioxidant property of different parts of papaya (Addai *et al.* 2013).

Table 2. Total phenolic contents (TPC) in methanol extract of *Carica papaya* leaves and seeds (LME and SME)

SN.	Conc. ($\mu\text{g/mL}$)	Methanol leaves extract (LME)			Methanol seeds extract (SME)		
		Abs.	TPC (mgGAE/g)	Mean TPC	Abs	TPC (mgGAE/g)	Mean TPC
1	10	0.812	54.63	93.18 \pm 0.40	0.612	41.93	89.14 \pm 0.45
			74.31			76.14	
2	25	1.095	79.13	1.091	1.091	81.52	119.76
			126.04			126.36	
3	50	1.165	131.78	1.166	1.166	126.36	
4	100	1.74		1.75			
5	125	1.867		1.843			

3.2.2 Total Flavonoid Content (TFC)

The TFC of methanol leaf and seed extract of *Carica papaya* presented in Table 3. The results show that the total flavonoid content (TFC) was comparatively higher in leaf extract (18.85 ± 0.061) than methanol seed extract (16.64 ± 0.065). Similarly, Asghar *et al.* (2016) also reported higher TFC in leaves ($15.54 \pm$

0.12) mg catechin equivalent (CE)/g than in seeds (08.62 ± 0.16) mg CE/g ethanol extract. Therefore, *Carica papaya* leaves and seeds are good sources of flavonoid content. Dwivedi *et al.* (2020) also reported that *Carica papaya* flowers are good sources of total flavonoid content (1.53 ± 0.10 mg QE/g dry weight).

S.N	Concentration ($\mu\text{g/mL}$)	Leaves methanol extract (LME)			Seeds methanol extract (SME)		
		Abs.	TFC (mgQE/g)	Mean TFC	Abs.	TFC (mgQE/g)	Mean TFC
1	10	0.038	10.76		0.012	8.54	
2	25	0.106	17.18	18.85 \pm 0.061	0.109	16.69	16.64 \pm 0.065
3	50	0.149	21.09		0.12	18.02	
4	100	0.207	26.36		0.178	23.15	

3.3 Biological Screening

3.3.1 DPPH Free Radical Scavenging Potential for Antioxidant Activity

The recorded absorbance with different concentration of ascorbic acid is shown in Figure 1. The DPPH free radical scavenging assay results showed a significant difference in scavenging activity among seeds and leaves of *Carica papaya*. Free radicals act as triggers to several degenerative diseases; therefore, samples having free radical scavenging activity can be of potent medicinal importance.

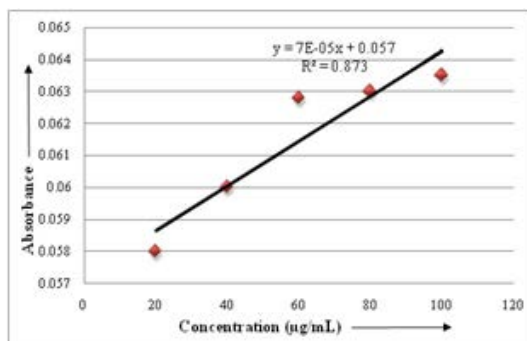


Fig. 1. Absorbance vs concentration of ascorbic acid.

The % radical scavenging activity of methanol extract of *Carica papaya* seeds and standard ascorbic acid was found almost similar and better than leaves extract (LME). The antioxidant potential has an inverse relation with IC_{50} value, lower value of IC_{50} represents high antioxidant ability. The IC_{50} value for seed methanolic extract (SME) was 15.48 ± 0.013 µg/mL which was comparatively lower than the IC_{50} (43.54 ± 0.007 µg/mL) of methanolic leaves extract (LME), showed that SME of *Carica papaya* is more effective as antioxidant compared to LME. The seed methanol extracts were observed as good antioxidants as their IC_{50} values were close to the standard ascorbic acid. Kumawat *et al.* (2012) reported 21.23 µg/mL IC_{50} values for ascorbic acid. Flavones and flavanols are the major compounds responsible for the antioxidant activity, which depends on the presence of free OH groups (Dwivedi *et al.* 2020). The present finding suggests that methanolic seed extract of *Carica papaya* could be a potential natural source of antioxidants and could have greater importance as

a therapeutic agent in preventing oxidative stress-related degenerative diseases (Kumawat *et al.* 2012). Dwivedi *et al.* (2020) studied the antioxidant and antibacterial properties of phytochemical extracts of *Carica papaya* flowers. Its leaves methanolic extract showed the presence of more phytochemicals and have phenolic, flavonoid content, and exhibited strongest antioxidant properties which can effectively scavenge reactive oxygen species compared to n-hexane, ethylacetate and ethanol solvent extracts (Nandini *et al.* 2020).

Here, the effective results were obtained from methanol extract of seeds of *Carica papaya* which showed the strongest DPPH radical scavenging activity as its IC_{50} value was near to standard ascorbic acid. So, this result show that the plant used for this study can play a significant role in the field of medicine based on the antioxidant property.

Table 4. Comparison of IC_{50} values of leaf and seed extracts with standard ascorbic acid.

S.N.	Sample	IC_{50} value (µg/mL)
1	Standard ascorbic acid	15.38 ± 0.013
2	Seed extract	15.48 ± 0.013
3	Leaf extract	43.54 ± 0.007

3.3.2 Antidiabetic Activity using α -amylase Inhibition Assay

Among five different extracts, the antidiabetic activity of methanol extracts of seeds and leaves *Carica papaya* were assessed using Acarbose as standard. An α -amylase inhibition assay was carried out to assess the antidiabetic activity of selected samples. The methanol extract of *Carica papaya* seed showed a lower IC_{50} value than leaf extract. The IC_{50} value of standard Acarbose was found as 56.12 ± 0.013 µg/mL. The IC_{50} value of seed extract was 46.99 ± 0.018 µg/mL, which showed higher antidiabetic activity than leaf extract. However, Cowan (1999) showed that the methanolic extract of *Carica papaya* leaves also possesses significant antidiabetic activity, which shows that leaves can develop drugs in combating drugs. The results provide useful information on pharmacological activities of the plant associated with people with diabetes (Gawli & Lakshmidevi 2015).

4. CONCLUSION

This study revealed that all extracts of leaves and seeds of *Carica papaya* consist of pharmacological substances. The leaf methanol extract contains more phenolic and flavonoid contents than seed methanol extract. These seeds methanol extracts exhibited good antioxidant and antidiabetic activity, whereas leaf methanol extract showed more antibacterial property in comparison to leaf chloroform extracts. The study thus reveals the potential value of *Carica papaya* in medicinal applications. However, further investigations are needed to identify the active compounds to confirm the antioxidant and antidiabetic seed papaya extract activity.

ACKNOWLEDGEMENT

The authors are grateful to Department of Chemistry, Tri-Chandra Multiple Campus, Tribhuvan University, Kathmandu, Nepal for providing laboratory facilities to conduct experimental work. We are thankful to Dr. Laxmi Prasad Thapa, Polyclinic Research Centre, Kathmandu, Nepal for antibacterial test.

REFERENCES

1. Addai, Z. R., A. Abdullah and S. A. Mutalib. 2013. Effect of extraction solvents on the phenolic content and antioxidant properties of two papaya cultivars. *Journal of Medicinal Plant and Research* 7(46):3354-3359.
2. Agarwal, A., S. Vyas and D. P. Agarwal. 2016. Therapeutic benefits of *Carica papaya* leaf extracts in dengue fever patients. *Scholars Journal of Applied Medical Sciences* 4(2A):299-302.
3. Aravind. G., D. Bhowmik, S. Duraivel and G. Harish. 2013. Traditional and medicinal uses of *Carica papaya*. *Journal of Medicinal Plants Studies* 1(1):7-15.
4. Ariffin, M. M., H. Y. Khong, N. Nyokat, G. M. Liew, A. S. Hamzah and K. Boonpisuttinant. 2021. In vitro antibacterial, antioxidant, and cytotoxicity evaluations of *Musa paradisiaca* cv. Sekaki florets from Sarawak, Malaysia. *Journal of Applied Pharmaceutical Science* 11(05):091-099.
4. Asghar, N., S. A. R. Naqvi, Z. Hussain, N. Rasool, Z. A. Khan, S. A. Shahzad, T. A. Sherazi, M. R. S. A. Janjua, S. A. Nagra, M. Zia-Ul-Haq and H. Z. Jaafar. 2016. Compositional difference in antioxidant and antibacterial activity of all parts of the *Carica papaya* using different solvents. *Chemistry Central Journal* 10(1):1-11. DOI: 10.1186/s13065-016-0149-0
6. Chakrabarti, R., B. Singh, P. V. N, L. Vanchhawng and K. Thirumurugan. 2014. Screening of nine herbal plants for in vitro α -amylase inhibition. *Asian Journal of Pharmaceutical and Clinical Research* 7(4):84-89.
7. Cowan, M.M., 1999. Plant products as antimicrobial agents. *Clinical Microbiology Reviews* 12(4):564-582.
8. de Beer, D., E. Joubert, W. C. A. Gelderblom and M. Manley. 2002. Phenolic compounds: A review of their possible role as in vivo antioxidants of wine. *South African Journal of Enology and Viticulture* 23(2):49-61.
9. Dhital, K. S. 2017. Phytochemical screening and antioxidant activities of *Mangifera indica* leaves grown in temperate region of the Nepal. *Journal of Pharmacognosy and Phytochemistry* 6(3):205-209.
10. Dwivedi M. K., S. Sonter, S. Mishra, D. K. Patel and P. K. Singh. 2020. Antioxidant, antibacterial activity, and phytochemical characterization of *Carica papaya* flowers. *Beni-Suef University Journal of Basic and Applied Sciences* 9(23):1-11. DOI: 10.1186/s43088-020-00048-w
11. Gawli, K. and N. Lakshmidivi. 2015. Antidiabetic and antioxidant potency evaluation of different fractions obtained from *Cucumis prophetarum* fruit. *Pharmaceutical Biology* 53(5):689-694.
12. Gogna, N., N Hamid and K. Dorai. 2015. Metabolomic profiling of the phytomedicinal constituents of *Carica papaya* L. leaves and seeds by ¹H NMR spectroscopy and multivariate

- statistical analysis. *Journal of Pharmaceutical and Biomedical Analysis* 115:74-85.
13. Jamuna, S., S. Paulsamy and K. Karthika. 2012. Screening of in vitro antioxidant activity of methanolic leaf and root extracts of *Hypochoeris radicata* L. (Asteraceae). *Journal of Applied Pharmaceutical Science* 2(7):149-154.
14. Khanal, S., D. P. Bhandari, L. Bhandari and A. Adhikari. 2020. Potent free-radical-scavenging activity of bark of *Poranopsis paniculata* (roxb.) Roberly. *American Journal of Essential Oils and Natural Products* 8(3):39-42.
15. Kumawat, B. K., M. Gupta, T. Chand and Y. Singh. 2012. Free radical scavenging effect of various extracts of leaves of *Balanites aegyptiaca* (L.) Delile by DPPH method. *Asian Journal of Plant Science and Research* 2(3):323-329.
16. Kousika. G. and A. D. Priyadharshini. 2020. Comparative study on antimicrobial activity of *Carica papaya* and *Nyctanthes Arbor-tristis* leaf extract and application in cosmetics formulation. *International Journal of Scientific Development and Research* 5(9):161-165.
17. Mishra, K., H. Ojha and N. K. Chaudhury. 2012. Estimation of antiradical properties of antioxidants using DPPH assay: A critical review and results. *Food Chemistry* 130(4):1036-1043.
18. Nna, P. J., O. J. Egbuje and D. C. Don-Lawson. 2019. Determination of phytoconstituents and antimicrobial analysis of the ethylacetate extract of *Carica Papaya* seed. *International Journal of Research and Innovation in Applied Science* 4(XII):1-7.
19. Nandini, G., T. S. Gopenath, P. Nagalambika, K. Murugesan, G. Ashok, M. S. Ranjith, P. Pradeep and M. B. Kanthesh, 2020. Phytochemical analysis and antioxidant properties of leaf extracts of *Carica papaya*. *Asian Journal of Pharmaceutical and Clinical Research*. 13(11):58-62. DOI: 10.22159/ajpcr.2020.v13i11.38956
20. Ogundele, A. V., K. O. Otun, A. Ajiboye and B. Eunice. 2017. Antidiabetic efficacy and phytochemical screening of methanolic leaf extract of Pawpaw (*Carica papaya*) grown in North Central Nigeria. *Journal of the Turkish Chemical Society* 4(1):99-114.
21. Pandey, B. and M. Rajbhandari. 2014. Estimation of total phenolic and flavonoid contents in some medicinal plants and their antioxidant activities. *Nepal Journal of Science and Technology* 15(1):53-60.
22. Panzarini, E., M. Dwikat, S. Mariano, C. Vergallo and L. Dini. 2014. Administration dependent antioxidant effect of *Carica papaya* seeds water extract. *Evidence-Based Complementary and Alternative Medicine*, 2014:1-13. DOI: 10.1155/2014/281508
23. Peter, J. K., Y. Kumar, P. Pandey and H. Masih. 2014. Antibacterial activity of seed and leaf extract of *Carica Papaya* var. Pusa dwarf Linn. *IOSR Journal of Pharmacy and Biological Sciences* 9(2):29-37.
24. Pokhrel, S., and K. Chaulagain. 2020. Phytoconstituents and biological analysis of *Acorus calamus* rhizome of Sindhupalchowk District, Nepal. *Bibechana* 17:104-109.
25. Pokhrel, S. and P. Neupane. 2021. Phytochemical analysis, antioxidant and antibacterial efficacy of methanol and hexane extract of *Centella asiatica*. *Bibechana* 18(2):18-25.
26. Saxena, M., J. Saxena, R. Nema, D. Singh and A. Gupta. 2013. Phytochemistry of medicinal plants. *Phytochemistry of Medicinal Plants* 1(6):168-182.
27. Udoh, P., I. Essien and F. Udoh. 2005. Effect of *Carica papaya* (Pawpaw) seeds extract on the morphology of male Wistar rats' pituitary-gonadal axis. *Phytotherapy Research* 19:1065-1068.

28. Vongsak, B., P. Sithisarn and W. Gritsanapan. 2013. Simultaneous determination of crypto-chlorogenic acid, isoquercetin, and astragalins contents in *Moringa oleifera* leaf extracts by TLC-densitometric method. *Evidence-Based Complementary and Alternative Medicine* 2013:1-7. DOI: 10.1155/2013/917609
29. Vuong, Q. V., S. Hiruna, P. D. Roacha, M. C. Bowyer, P. A. Phillips and C. J. Scarlett. 2013. Effect of extraction conditions on total phenolic compounds and antioxidant activities of *Carica papaya* leaf aqueous extracts. *Journal of Herbal Medicine* 3:104-111.
30. Waterhouse, A. 2002. Determination of total phenolics. In: *Current protocols in food analytical chemistry* (Ed Wrolstad, R. E). John Wiley and Sons, New York, Units 11.1.1-11.1.8.
31. Zuhair, R. A., A. Aminah, A. M. Sahilah and D. Eqbal. 2013. Antioxidant activity and physicochemical properties changes papaya (*Carica papaya* L. cv. Hongkong) during different ripening stages. *International Food Research Journal* 20(4):1653-1659.