

Current understanding and the potential future of spinal fusion

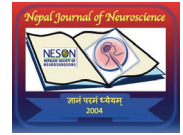
Krishna Sharma¹ 

¹Consultant Neurosurgeon Annapurna Neurological Institute and Allied Sciences, Maitighar, Kathmandu, Nepal

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Introduction

Trauma, infection, cancer, and congenital abnormalities are examples of spinal pathologies that can compromise structural stability. The spinal cord and nerve roots, which the spine is meant to safeguard, may become compressed and damaged as a result. To stop further damage in such cases, the spine needs to be decompressed, re-aligned, and fused. Spinal fusion is a normal bone-healing procedure that helps to regain structural stability. Although this unassisted process may accomplish alignment and strength, it may do so slowly and insufficiently, causing ongoing compression on neural structures.^{1,2} Therefore, grafts and instrumentation may be required to supplement the natural process of spine fusion. Pharmacological agents, graft expanders, and graft enhancers may need to be added to the process in order to achieve sufficient and quicker fusion.

For Pott's spine, Albee published the first American account on assisted spinal fusion in 1911.^{3,4} Since then, there has been a sharp rise in spinal fusion procedures. The increase in annual procedures done in the USA, from 174,223 in 1998 to 413,171 in 2008, is evidence of this.⁵ Fusion is increasingly being included in all major spine surgeries. Time has also demonstrated the related effects and difficulties of spine fusion which could be reduced

mobility, pseudoarthrosis, diseases of the neighboring segment, revision surgeries, rising costs, and other related morbidities. Continued study is necessary with an emphasis on appropriate case selection, risk reduction, fusion techniques, and postoperative spinal fusion optimization in order to maximize value and keep complications at a minimum level.

Hibbs treated a 9-year-old child with a kyphotic deformity at the turn of the 20th century by removing the spinous processes, repositioning them over the interspinous space to encourage fusion, and repairing the periosteum over the fusion mass.⁴ During the same time frame, Albee suggested using bone grafts to improve spinal fusion in Pott's disease patients.^{3,6} Since then, the procedure for fusing the spine has progressed from fusing it alone to fusing it with instruments and the addition of medication.

Pathogenesis of fusion

Normal progression of spine fusion process includes inflammatory response, osteogenesis, angiogenesis, and remodelling. The spine does not fuse in the same way as the other bones, with minimal callus formation, for unknown causes. Bone fusion takes between three and six months. The oxygen tension, force application trajectory, and motion at the location of fusion all have a significant impact on bone fusion. In nearly a year, the bony callus matures, remodels in accordance with the strain applied⁷, and completes the fusion process.

Factors influencing fusion

Co-morbidities of patients, major modifiable risk factors, surgical techniques, postoperative care, the use of grafts and implants, as well as the type of fusion carried out, are just a few of the many variables that can affect fusion.⁸ To reach the ideal spinal fusion state, these variables should be recognized and optimized. They can be broadly separated into systemic and local variables. The specifics of each of these factors are outside the purview of this review and can be researched further using the references given.

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Address for correspondence:

Dr. Krishna Sharma MBBS, MS, DNB

Associate Professor

Consultant Neurosurgeon Annapurna Neurological Institute and Allied Sciences, Maitighar, Kathmandu, Nepal

E-mail: Krishnasharma@yahoo.com

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Systematic factors

- Due to the reduced availability and delayed mobilization of growth factors, osteogenic factors, and anti-inflammatory factors in systemic diseases like metabolic bone diseases and immune compromised states like diabetes, renal failure, malignancy, and rheumatoid arthritis, the fusion process is slowed down.^{9,10}
- Osteogenesis, overall bone mass, and bone metabolism are all significantly influenced by hormones. Therefore, changes in hormone levels, particularly those of growth hormone, thyroid hormones, parathyroid hormone, leptin, adiponectin, angiotensin, cortisol, erythropoietin, insulin, oxytocin, and calcitriol, as well as estrogen and androgen, have a detrimental impact on osteogenesis.^{11,12,13}
- Low albumin levels, iron deficiency anemia, leucopenia and a negative nitrogen balance are indicators of poor nutritional condition.^{14,15,16} These reduce healing process, and fusion. Patients who have hemoglobin A1c levels under 8% have a poor chance of fusion.¹⁷
- Osteoporosis and low bone mineral density (BMD), which is defined as a BMD 2.5 or more below the young adult mean or a T-score at or below -2.5, as well as a lack of nutrients like calcium, iron, and magnesium, all essential for bone formation, can delay fusion.^{18,19,20,21,22,23} If BMD is less than 0.3 g/cm², internal fixation devices ought to be avoided.²⁴
- Drugs that inhibit bone development and healing include corticosteroids, methotrexate, adriamycin, H2 blockers, and NSAIDs. Additionally, these medications decrease bone fusion by causing mineral losses that are necessary for fusion.^{25,26,27}
- Smoking decreases bone fusion by as much as 56% due to the effects of nicotine.^{25,28,29,30,31}
- Alcohol intake on a regular basis slows down fusion as well.¹⁷
- Spinal fusion is delayed in obesity with greater body mass index (BMI).^{32,33}
- Lack of exercise hinders union.³⁴
- Genetic factors: It has been demonstrated that alterations in the genes for the vitamin D receptor (VDR), the oestrogen receptor (ER), and collagen type II (COLIA1) affect bone fusion.^{35,36}

Local factors

- During fusion, maintaining equilibrium is crucial, including sagittal balance. Sagittal balance problems result in deformity, pain, neurological compression and impairments, as well as a changed force

distribution that slows the rate of fusion.^{37,38} Sagittal balance and normal spine biomechanics must be attained in order to improve fusion and re-establish mechanical stability.³⁷

- The donor graft can fuse more quickly if there is sufficient compressive force applied to it, which encourages the ingrowth of vascular branches and proliferating mesenchymal cells from the cancellous host bone into the donor graft.³⁹
- Multiple level fusion and junctional area involvement have a detrimental effect on fusion and increase the chance of non-union.⁴⁰
- There are seven biological variables that affect fusion.⁷
 - Sufficient local blood flow, particularly in the union bed vascularity.
 - The use of grafts with good osteogenic potential and osteo-progenitor cell supply.
 - Proper receiver site setting, such as decortication, to enhance fusion.
 - If radiotherapy is administered within three weeks of operation, it slows fusion.⁴¹
- Diseases of the bone, such as tumors, fibrous dysplasia, and Paget's disease, inhibit union.^{42,43}

Radiologically, instability that requires fusion is defined as having at least 4mm of antero-posterior translation above the L1-L5 levels, 5mm of translation at the L5-S1 levels, or 11 degrees or more of end plate angular shift at a single level compared to an adjacent level.¹⁷

Standard radiography, dynamic radiography, radio-stereometric analysis (RSA), CT, and MRI can all be used to determine the fusion state. The most popular way to evaluate spinal fusion is with plain static spine radiographs, which look for bridging trabecular bone across the section.⁴⁴ The existence of deformity under physiologic load and graft resorption, implant subsidence or migration, implant integrity, and non-union, on the other hand, are signs of non-union.⁴⁵ However, there are some concerns about the accuracy of X-rays in identifying small voids connected to pseudoarthrosis, especially in the thoracic and lumbar spine. It was found that almost 25% of those who had been labelled as fused on plain radiographs had not actually fused.⁴⁶

Fusion can be divided into three phases radiologically.⁴⁷ (Table 1)

With a specificity of 89% and a sensitivity of 91%, dynamic X-rays improve the efficiency of fusion detection. Pseudoarthrosis has been identified on lateral dynamic radiographs by movement of more than 2 mm across fused segments between spinous processes and a Cobb angle greater than 2°. For pseudoarthrosis, a Cobb angle of at least 4° had a 100% positive predictive value (PPV).^{48,49}

Table 1

Classification of Interbody Fusion Success: Brantigan, Steffee, Fraser (BSF)

BSF-1: Radiographical pseudarthrosis is indicated by collapse of the construct, loss of the disk height, vertebral slip, broken screws, displacement of the carbon cage, or significant resorption of the bone graft, or lucency visible around the periphery of the graft or cage.

BSF-2: Radiographical locked pseudarthrosis is indicated by lucency visible in the middle of the cages with solid bone growing into the cage from each vertebral end plate.

BSF-3: Radiographical fusion: bone bridges at least half of the fusion area with at least the density originally achieved at surgery. Radiographical fusion through one cage (half of the fusion area) is considered to be mechanically solid fusion even if there is lucency on the opposite side.

Reproduced with permission from Fogel GR, Toohy JS, Neidre A, Brantigan JW: Fusion assessment of posterior lumbar interbody fusion using radiolucent cages: X-ray films and helical computed tomography scans compared with surgical exploration of fusion. *Spine J* 2008;8(4):570-577.

The ability of computed tomography (CT) to clearly display the bridging trabecular bone, a sign of arthrodesis, is a unique advantage.⁵⁰ An accurate CT scan can reliably show a posterolateral fusion 89% of the time.⁵¹ A spinal segment’s motion and stability are evaluated using quantitative motion analysis (QMA) software both before and after operation⁵², as well as for real-time feedback during fusion and instrumentation. The combination of CT scans and dynamic radiograph QMA, which has a positive predictive value (PPV) of 100% and a negative predictive value of 73%, is fast becoming the gold standard of treatment.

By converting two-dimensional data to three-dimensional data through radio-stereometric analysis,⁵³ it is possible to calculate the three-dimensional motions between grafts. The invasive nature of this methodology makes it largely unsuitable for regular clinical use. Similar to that, MRI is not the best technique for evaluating union. It is only applicable to instances where non-metallic cages have been used and inter-body fusion has been assessed.⁵⁴

Indications of a spine fusion keep on evolving as we gain a better knowledge of natural history, spinal fusion biomechanics, pathophysiology, socioeconomic factors, patient factors, surgeon factors, resource availability, and regional beliefs and customs. The primary indication is the spinal instability, both static and dynamic. Other important indications include:

- Complex spinal canal stenosis, which is a stenosis of the spinal canal accompanied by grade II and III spondylolisthesis as well as serious, static, or progressive deformity and pain.^{55,56,57}
- Recurrent disc herniation, typically with more than two recurrences and instability.^{58,59}
- Iatrogenic instability is a state of instability brought on directly by surgical and/or medical intervention. They are more likely to need revision surgery.^{60,61,62}
- Progressive congenital spine deformities that result in instability and compression of the neural tissue
- Kyphotic flat back condition

- Laminectomy of more than three levels
- Osteoporotic vertebral fracture not responding to conservative therapy⁶³

A scoring system has been developed by Kulkarni et al. to help determine who would most likely benefit from spinal fusion. Fusion surgery would be beneficial if the result is higher than 5.5.⁶⁴

TABLE 1. The Scoring System

Factors	Score
Clinical factors	
Mechanical back pain (bothersome pain, present on loading the spine and decreasing on lying supine)	2
Age <70 y	1
High-demand activity	1
Radiologic factors	
Segmental kyphosis (change in angle from lordosis to kyphosis)	1.5
Segmental dynamic spondylolisthesis (> 2 mm translation on lateral dynamic films)	1
Disk height (50% more than adjacent cephalad level)	1
Bilateral facet effusion (bilateral full facets on T2-weighted axial magnetic resonance imaging > 1 mm fluid)	1
Sagittal facets (measured by calculating the angle generated by connecting the 2 endpoints of each facet on a preoperative axial lumbar magnetic resonance imaging (midcut through the disk) and a line connecting the 2 ventral points of each facet joint. > 50 degrees = sagittal facets)	1
Technical factor	
Feasibility to decompress (anatomy unfavorable for complete decompression via a laminotomy that is both facet and midline sparing)	1.5

Complications of fusion

Up to 25% of fusion surgeries can have complications.^{65,66} The main complications are:

1. Adjacent segment disease (ASD)

Adjacent segment disease (ASD) is a significant issue following fusion surgeries, with a prevalence of nearly 25% within 10 years of the initial fusion and an incidence of symptomatic ASD of 2.9%.^{67,68,69} Fusion modifies the

spine's biomechanics, increasing the mechanical load on the nearby disc space.⁷⁰ Fusion can result in rapid disc degeneration, pain, deformity, facet arthropathy, and increasing stenosis with neural compression. Taken together, these conditions lead to ASD. This risk has been greatly decreased by recent developments in taking into account and correcting sagittal balance as well as attention to limiting disruption of adjacent segment tissues.⁷¹ The result of adjacent segment disease is progressive deformity, especially junctional kyphosis.⁷²

2. Non-union or pseudoarthrosis

Pseudoarthrosis, also known as non-union, is a surgical fusion failure. It is the primary reason for revision surgery and has a prevalence between 5 and 42%.^{73,74,75} Smoking, patient co-morbidities, multilevel arthrodesis^{76,77}, the use of medications that affect osteogenesis, and ineffective fusion techniques are a few of the many variables that delay fusion.

Other complications include

3. Mobility restrictions, primarily flexion and extension
4. Osteomyelitis of the fused section^{78, 79, 80}
5. Vascular and neural compression associated with displacement for transplant^{81, 82}
6. Biomechanical collapse⁸³
7. Failed back condition and persistent pain syndrome.⁸⁴

To minimize these complications, the following steps could be taken,

- Managing the patient's comorbid conditions, such as osteoporosis, diabetes, renal failures, cancer, etc., as well as addressing nutritional deficits and other issues.
- Preoperative analysis of clinical features and imaging and planning accordingly.
- Correcting and maintaining biomechanics like sagittal alignment^{85,86} and lumbar lordosis (LL) in relation to a patient's-age-adjusted pelvic incidence (PI)^{86, 87}
- Carefully choosing fusion levels^{88, 89}, preventing multilevel fusion, and excluding hypermobile adjacent segments⁹⁰ are important for maintaining motion segments.
- Avoid including adjacent degenerative levels.
- Use grafts that are of right sort, size, and shape.
- Interbody placement of the graft completes a circumferential (360°) fusion, lessens stress on the pedicle screws, and corrects lordosis, and thus may improve fusion rates from 83.3% to 95.9%.^{91, 92, 93}
- The shortest plates and implants should be used.
- Multiple rods that decrease motion at L5-S1 have been shown to reduce lumbosacral junction stress.^{94,95,96}

- Stiffer rods (cobalt-chromium or steel) can reduce rod motion and strain and thus improve strength and resistance to fatigue better than less stiff rods (titanium). However, proximal junctional kyphosis (PJK) may occur more commonly with a stiffer rod.⁹⁷
- In-depth understanding of surgical technique, and preservation of critical structures like the adjacent annulus, cranial and caudal anterior longitudinal ligament (ALL), and longus coli muscle help to lessen fusion complications.^{98, 99}
- Meticulous prior planning with the goal of eradicating all sources of infection and use of suitable and sufficient antibiotics when necessary.
- Supervised physical therapy before and after surgery, helps to keep the graft in position, prevent stiffness or contractures, maintain alignment, and gradually build up the associated muscle strength.

Osteoporosis

Osteoporosis is a co-morbidity that remains a significant risk factor for unsuccessful outcome of assisted fusion surgery due to fusion construct failure, interbody cage subsidence, compression fractures and pseudoarthrosis^{100, 101, 102}. The NIH Consensus Statement predicted that osteoporosis would affect 12% of the population overall, 4.2% of males, and 18.8% of women, particularly after the age of 50.^{103, 20} Women have a 29% lifetime risk and males have a 14% lifetime risk of developing an osteoporotic spine fracture.¹⁰⁴ There is slowed osteogenesis and poor grip to hold the implants in place.^{105, 106, 20, 21, 22, 107} One must identify those who have or at risk for osteoporosis by determining metabolic bone health panels (vitamin D, parathyroid hormone, thyroid-stimulating hormone, albumin, and pre-albumin levels) and dual-energy X-ray absorptiometry (DEXA) scans to evaluate bone mineral density and initiate medical optimization before surgery.^{108, 109, 110, 111, 22}

Pharmacotherapeutic approaches, such as the supplementation of calcium, vitamin D^{108, 112}, anti-osteoporotic medications like bisphosphonates, hormonal therapy, calcitonin, and teriparatide, depending on the severity and therapeutic reaction, are beneficial during the perioperative phase. These have been demonstrated to lower the chance of bone-implant failure while increasing fusion mass and rates.^{113, 114} The surgical approach must be modified to preserve the endplates,¹¹⁵ use longer constructs, concurrently use more anchors, perform additional interbody fusion, do under-tapping the pedicle and use longer, bigger, cement-augmented and growth factor-coated screws.¹¹⁶

Assisted bone fusion

The natural fusing process is further aided and enhanced by assisted bone fusion, which also lessens complications. In order to accomplish a better, faster, and stronger union, bone graft substitutes, fusion enhancers, and implants are being used to assist fusion. Broadly, assisted bone fusion can be divided as either structural graft or fusion substrate.

1. During the process of fusion, structural grafts provide the spine with instant physical support. These include,
 - a. Bone grafts
 - i. Autologous bone grafts
 - ii. Allograft
 - b. Artificial Grafts
2. Fusion substrates are those that enhances the process of fusion itself. The fusion substrates can be
 - a. Autograft
 - b. Allograft
 - c. BMP (Bone morphogenic protein)
 - d. Other synthetic products

A. Bone grafts

Bone grafts, either on-lay or inlay, are used to achieve better fusion. The appropriateness of graft depends on its properties, namely osteo-induction, osteo-conduction, and osteogenesis. Among the bone grafts, autologous (autograft) bone graft is the time tested and gold standard living graft obtained directly from the index host. It has very high osteogenic, osteo-inductive and osteoconductive properties.¹¹⁷ At no extra cost, it is readily available from adjacent exposed spinous processes or either anterior or posterior iliac crest.¹¹⁸ These have up to 100% fusion in instrumented spinal fusion procedures.^{119, 120, 121, 122} It became less popular due to donor site complications like pain, neurovascular injury, anterior–superior iliac spine avulsion fracture, hematoma, and infection.^{123, 124, 125, 126} These limitations have led spine surgeons to look for other potential substitutes.

Allograft is a popular commercially available substitute of autologous graft^{125, 127, 128} with fusion rate up to 94.3%¹²⁹ and similar clinical and radiologic out-comes.¹³⁰ It is a cadaveric bone, sterilized to remove infectious agents and processed to contain only inert material. They have only osteoconductive properties acting as a scaffold for the bone from adjacent bone to grow. Allografts take longer time to fuse but give an immediate additional structural support.¹³¹ While available in large amounts, allograft carries a theoretical risk for disease transmission, including hepatitis B or C and HIV though the risk is extremely low: less than 1 in 1,000,000.^{132, 127} Allograft also has the potential to incite a host immune reaction.¹³³ Allograft is

available in a variety of forms, including cancellous bone, cortical bone, and demineralized bone matrix (DBM). With absent donor site morbidity, equivalent outcomes, and increased availability, allograft has become a popular choice for many surgeons.

Xenograft are the grafts obtained from other species and are not a popular source of bone graft.

B. Artificial Bone graft

Bone graft substitutes or artificial bone grafts have been created to address the drawbacks of organic bone grafts. They can be resorbable or non-resorbable. It is necessary to take into account the unique mechanical, chemical, and immunological characteristics of each class of bone graft alternatives. Currently, there is no single substitute material that contains all the ideal properties for a bone graft substitute¹³⁴, i.e. a three-dimensional structure strong enough to mimic the mechanical and biological properties of natural bone, allowing osteo-induction by having surface proteins necessary for osteoblast attachment and containing cells and signalling factors to promote osteogenesis.¹³⁵ It is immunologically inert and prevents the formation of fibrous tissue, which can lead to aseptic loosening.¹³⁴ Bone graft substitutes can be supplemented with natural bone, bone marrow aspirate, bone graft expanders, bone growth factors, or stem cells to improve fusion and achieve a fusion rate that is similar to autogenous bone graft. There are no complications or morbidities at the donor location, a shorter operating time, less blood loss, and a quicker and more powerful fusion. With in-lay grafts, there is less sinking and the height of the intervertebral disc is maintained.¹³⁵ The various artificial graft replacements include:

1. Metallic structural grafts

Metals are powerful in both compression and strain. However, they can cause an unspecific immune reaction and do not offer a natural substrate for cell adhesion.¹³⁶ The surface is customized and made rougher with the introduction of additive fabrication. Computer design is used to manage the layering of three-dimensional structures. New surface properties are being developed in metals to enable improved osteointegration.¹³⁷ They are sprayed with bone growth factors and promote bone fusion in the metal-bone interface. Titanium metallic implants are the most frequently used.

2. Polyetheretherketone (PEEK) structural grafts

PEEK cages are made of plastic and have rigidity characteristics comparable to those of normal bones. It is possible for radiological fusion study because it is radio-opaque. When used alone, PEEK causes fibrosis and inflammation, which may lead to implant separation. It results in a similar fusion when impregnated with titanium and bone growth stimulators.^{138, 139}

3. Ceramic structural grafts and bone graft substitutes

Ceramics are inert, calcium-derived materials and, one of the most widely studied groups of bone substitutes in spinal fusion. It includes bioactive glass, calcium phosphates, and corals. It is an attractive graft option as they demonstrate good biocompatibility and osteo-conduction.^{136,137} It is biodegradable by osteoclast-mediated resorption. They have limited osteo-induction potential. These synthetic grafts are easily manufactured, have porous structure resembling cancellous bone that enhances ingrowth of bone while offering scaffolding with immediate and significant mechanical strength. They have limited immunogenicity and have no risk for disease transmission.^{125,127,121} The implants have demonstrated satisfactory outcomes and good efficacy compared to autologous bone grafts.^{140,141} They have shown successful outcomes and high fusion rates even in multilevel and revision fusions.^{142,143}

4. β -tricalcium phosphate (TCP) bone graft substitutes

β -tricalcium phosphate (β -TCP) is one the most used and potent synthetic bone graft substitute. It is not only osteoconductive, but also osteo-inductive. These properties, combined with its cell-mediated resorption, allow full bone defects regeneration. They can be moulded or cut, allowing great versatility in surgery, and can act as carriers for demineralized bone matrix (DBM) or other growth factors. β -TCP has shown fusion rates up to 85% when used alone or up to 96% when used in conjunction with iliac crest bone graft (ICBG).^{140,144}

One unique subset of ceramic bone substitutes is silicate substituted calcium phosphate, which have both osteoconductive and osteoinductive properties. Its osteoinductive ability originates from silicate's negative charge, which attracts osteoblasts to the ceramic implant.¹⁴⁵ Despite their cost-effectiveness and fusion efficacy, ceramics are brittle and have poor resistance to tensile forces, making them susceptible to fracture. Additionally, ceramic resorption rates vary widely, with β -TCP absorbed over a period of months, while hydroxyapatite may remain latent in the body for up to a decade.^{127,146} Tricalcium phosphate has been associated with soft tissue inflammation¹⁴⁷ and calcium sulphate has been associated with serous drainage.¹⁴⁸

5. Polymers

Polymers include a vast array of materials, ranging from natural (collagen, chitosan, silk, hyaluronic acid, and peptides) to synthetic compounds (polyglycolic acid and polylactic acid). Naturally derived polymer scaffolds, such as collagen or chitosan, have the ability to resorb and also contain signalling factors for cell migration. They

lack the ideal mechanical properties of bone and have to be integrated with other harder materials.¹³⁶

6. Peptide hydrogels

Peptide hydrogels are a new bone graft substitute that have shown promise in the regeneration of tissues, with some reparative potential of cartilaginous, neuronal, and cardiac tissues.¹⁴⁹ Hydrogels are synthesized from the molecular self-assembly of amphiphilic peptides into an entangled nanofiber structure, which is similar to the extracellular matrix of native tissues.¹⁵⁰ Moreover, they can be engineered to contain epitopes such as the $\alpha 5 \beta 1$ integrin receptor that promote cell migration and adhesion.^{151, 136} Hydrogel materials can be combined with osteogenic cells and assembled into a matrix that allows osteoid formation and can be tuned to degrade at an appropriate time.^{136, 152} Hydrogels can also act as a delivery system that maintains and releases rhBMP-2 from microporous tri-calciumphosphate in controlled fashion at the surgical site while preventing systemic diffusion.¹⁵³ Thus, they represent a new horizon in bone grafting, offering the panacea of osteoinductive, osteoconductive, and osteogenic properties.

C. Bone graft enhancers

Bone graft enhancers can be used in conjunction with bone grafts and bone graft substitutes, to help with bone fusion. Most of these bone graft enhancers are still in trial form and need more experience before a firm commitment can be made for clinical use. The popular ones are:

1. Growth Factors and Gene Therapy

The common available growth factors include transforming growth factor beta (TGF- β) and platelet-derived growth factor (PDGF). These are signalling proteins that induce cellular division and/or differentiation and bone matrix synthesis thus allowing bone growth. The growth factors being used are mainly derived from platelet rich plasma.¹³⁶

Transforming growth factor beta (TGF- β)

The most widely researched member of TGF- β is Bone Morphogenetic Protein (BMP).^{154, 155} BMP was first extracted by Marshall Urist in 1980 from demineralized rabbit bone and were shown to be able to induce bone morphogenesis across species.¹⁵⁶ It contains many crucial factors in bone formation^{125,127,155} Molecularly, they function by binding to a cell surface serine-threonine kinase receptor, which then transduces the signal through SMAD and ras/raf proteins to activate the gene expression necessary for bone production.¹⁵⁷ BMPs also potentiate cell differentiation, causing mesenchymal cells to become osteoblasts and stimulate osteo-induction.¹⁵⁷ At lower concentrations, BMPs induce endochondral ossification¹⁵⁸

and at higher concentrations, through trans-membranous bone formation, it directly forms bone without a cartilage intermediary. BMPs have been shown to be safe and effective promoters of local bone healing in multiple animal studies^{159, 160, 161, 162} BMP has been shown in humans to increase the rate of fusion, especially in cases of refractory non-union.⁷

BMP is incorporated in carrier material like collagen sponge or ceramic, such as calcium phosphate, and delivered to the fusion site to enhance fusion. This vehicle both serves as an osteoconductive agent for bone and also improves the tissue retention of BMP. Clinical trials involving BMP showed improvement in Oswestry Disability Index (ODI) scores at 24 months compared with iliac crest harvest, as well as a 12% higher fusion rate.¹⁶³ There are two preparations of BMPs available for clinical use: recombinant human BMP-2 (rhBMP-2) and rhBMP-7.¹⁶⁴ rhBMP-2 is genetically produced with recombinant technology, and is highly osteo-inductive, inducing bone formation by stimulating the differentiation of mesenchymal cells into chondroblasts and osteoblasts.^{165, 166} It has the ability to stimulate patient's own cells to make more bone and has been suggested as an innovative material to increase the fusion rate. rhBMP-2 helps with accelerated bone formation to achieve fusion.⁹² Several prospective trials have demonstrated equivalent fusion rates between^{167, 168} 60 to 100%, in instrumented fusion.¹⁶⁹ It provides a strong protective effect against pseudarthrosis and is safe and effective for grafting, with no significant complications other than radiculitis.^{170, 171} RhBMP-2 has action on both osteoclast and osteoblast function, thus it enhances bone growth and it also induces transient bone resorption.¹⁷² It is associated with complications like postoperative edema, dysphagia, cancer.^{173, 174} ectopic bone formation, heterotopic ossification, end-plate resorption¹⁷⁵ retrograde ejaculation, osteolysis, post-operative radiculitis, and seroma formation^{176, 177} especially in cervical spine.^{178, 179, 180, 181, 182, 183, 184} rhBMP-2 is expensive.¹⁸⁵ However, studies evaluating costs have shown that the cost at 2 years post-operatively is actually less with BMP than ICBG due to decreased revision surgeries.¹⁸⁶ BMP is available in several forms, including putty, sheets, and within a glycerol carrier.

Mesenchymal stem cells (MSCs)

Regenerative medicine has investigated the role of mesenchymal stem cells (MSCs), a renewable population of undifferentiated multipotent cells, derived from bone marrow or bone marrow aspirate. It can give rise to the various types of mature cells like muscle, bone, tendons, fat, and other stromal tissues.¹⁸⁷ It was first described by Friedenstein.¹⁸⁸ Bone can be derived from mesenchymal tissues, and therefore MSCs are clinically useful in the context of bone healing and bone formation. They can

be harvested from the host with minimal morbidity and even be modified to secrete osteo-inductive factors, which are implanted on an osteoconductive scaffold.¹⁸⁹ They have high potential of both osteogenic and osteoinductive properties within the fusion bed. Both local bone and distant site autologous bone grafts can serve as a source of stem cells at the fusion site, and many surgeons incorporate MSCs directly into grafting material with hopes of improving fusion rates.

Gene therapy is an additional endeavour to enhance fusion. When BMP gene is delivered to a host cell by a vector, it has been shown in animal models to induce fusion even in non-osteoid tissue^{190, 191}. When compared with locally delivered BMP, the gene therapy has shown to increase the bioavailability of BMP by producing ongoing osteogenic expression locally¹⁹² Vectors to deliver the BMP gene range from viral (adenovirus or herpes virus) to nonviral (liposomes, electroporation) media. Delivery to a host may be either ex vivo (implantation of transfected cells into the host) or in vivo (injection of genes directly into host cells).¹⁹¹ Riew et al. were able to show that ex vivo BMP-transfected bone marrow cells, replanted in rabbits, were able to produce spinal fusion.¹⁹³ Currently, gene therapy is limited by the massive immune response against viral vectors.¹⁹⁴

2. Electrical stimulation

Electrical stimulation is one of the therapies available to enhance spinal fusion. This therapy has been used for more than 30 years^{195, 196, 197}. Normally, when bone is mechanically strained, electrical potentials are generated; electronegative potentials are found in areas of compression and electropositive potentials in areas of tension. Bone is formed in the electro-negative regions and resorbed in the electro-positive regions. These electric fields may form the basis by which bone remodels in response to mechanical stimuli (Wolff's Law).¹⁹⁸ Three types of electrical stimulation are available clinically: direct current (DC), capacitive coupling (CC), and inductive coupling (IC) such as pulsed electromagnetic fields (PEMF) and combined magnetic fields (CMF). The mechanisms of action of each of the three electrical stimulation therapies differ. Broadly, they upregulate mRNA for growth factors^{199, 200, 201, 202, 203, 204} like BMP-2, -4, -6, -7, FGF-2, and VEGF^{199, 200}, resulting into upregulation of several synergistic growth factors and promote bone healing²⁰⁵ Various electrical stimulation devices have been designed to deliver these fields to enhance bone formation.^{7, 205} The DC technology requires surgical implantation of the device whereas IC and CC technologies are non-invasive methods of producing electric fields at the fusion site. All of these technologies can also be utilized as adjuncts to surgical procedures using bone grafts. Treatment usually lasts for a minimum of 6 months post-implantation, after which the procedure can

be discontinued at the discretion of the surgeon. A large multicentre randomized double-blind clinical studies by Ken WJ²⁰⁶, using the above mentioned methodologies of electrical stimulation to enhance radiographic and clinical spinal fusions showed a statistically significant higher success rate of 85 to 91.5% compared to 65 to 80.5% in the control groups. Electrical stimulation has been accepted as an established cost effective adjunct to spinal surgery, improving the outcomes of spinal fusion²⁰⁷ typically in high risk patients with fractures such as uncontrolled diabetes, untreated osteoporosis or continued nicotine use.

D. Instrumentation and implants

Spinal instrumentation has become an integral part of spinal fusion as it allows to achieve immediate stability, enhance fusion, allow early mobility, correct deformities and maintain alignment till fusion occurs. The role of postoperative external arthrosis is lessening. The chemical composition, hydrophilicity, topographical nature and overall roughness of the surface of an implant play role in bony fusion.^{208,209,210,211}

After placement of an implant in patients, the surface of the implant is coated with proteins from the blood and serum. This protein layer facilitates the migration of mesenchymal progenitor cells into the implant surface via the $\alpha 2\beta 1$ integrin receptor, a major collagen type 1 receptor.²⁰⁹ These cells then differentiate into an osteoblastic lineage to form new bone. The roughness of implants is important for osteoblastic differentiation²¹² as more progenitor cells can get attached to the surface to induce bone fusion.²¹³

The latest achievements in implant development are:

1. Nanoscale surface technology

Application of nanoscale surface technology in the implant and instrumentation produces roughened titanium and generates an osteoblastic environment.^{209,210,214,215} The increase in surface area and roughness of the implant surface allows host cells to attach on a molecular level via cellular membrane receptors. This interaction can trigger osteoblastic-lineage differentiation and improve fusion results.²¹⁶ There are two primary types of manufacturing of Nanoscale surface technology in the production of spine implants, additive and subtractive. In the subtractive manufacturing, surface features are generated through the removal of material. Acid etching and grit blasting are two forms of subtractive manufacturing.²¹⁷ Although subtractive manufacturing is much more commonly used, these techniques waste material substrate, and the physical process itself limits the types of designs that can be created. Interbody grafts are produced from treated pieces of titanium, and subtractive technologies are used to produce submicron surface textures.²¹⁵ Nano-roughened titanium surfaces induce greater differentiation

of osteoblasts from mesenchymal stem cells, as compared with PEEK-treated surfaces. Roughened titanium also increases osteoblast maturation and produce an osteogenic environment that contains bone morphogenetic proteins (BMPs), as compared with smooth titanium and PEEK. Similar studies have shown that nanoengineered implants increase stimulation of local growth factors, including BMPs, VEGF, and TGF- β .²¹⁵

Hardware infection can be a life-threatening sequela of spinal fusion surgery.²¹⁸ Nanotechnology is also used to fight infection by coating the implants with bactericidal antibiotics or chemical like silver. Bacteria adhere to implants via the formation of a complex glycocalyx that protects them from antibiotics, making eradication very difficult.²¹⁹ The surface features of an implant can decrease bacterial adhesion.²²⁰ Nano-roughened surfaces have been shown to significantly decrease rates of bacterial adhesion, specifically of *Staphylococcus aureus*, *S. epidermidis*, and *Pseudomonas aeruginosa*.²²¹ Moreover, silver nanoparticles have been shown to have a bactericidal effect while still being biocompatible with bone. They achieve this through the release of silver ions from soluble complexes, which then generate reactive oxygen species that break down bacterial components. Silver nanoparticles can be applied to an implant via silver plasma ion immersion or by vapor deposition. Nanoparticles have also been shown to inhibit bacterial biofilm formation in animal studies. In particular, titanium pedicle screws coated in silver-based nanoparticles have been shown to be bactericidal in rabbits because of their release of silver ions.^{222,223, 219}

2. Three-dimensional (3D) printing

Additive manufacturing, known as three-dimensional (3D) printing²²⁴ involves customised layer-by-layer construction of complex 3D objects using computer-aided design software or the deposition of a material coating on the implant itself.²¹⁷ These provide more surface area for adhesions of macrophages and growth factors, thus enhancing fusion and strong bone-implant interface fusion.

Use of PEEK in implants is common, primarily because of its radiolucency and modulus of elasticity that closely resembles that of native bone. However, it produces fibrous encapsulation because of its induction of an inflammatory environment, and this can result in nonunion.²¹¹ Porous surface may also be applied to PEEK by extruding it through a bed of sodium chloride crystals, which has been shown to improve osteoconductivity but only in the presence of osteogenic mediators.^{225,226} To overcome this limitation, PEEK implants are sprayed with titanium spray (post-processing) which improves the surface properties of PEEK, but this method has been associated with increased generation of wear debris.

3. Bio-absorbable implants

Bioabsorbable interbody fusion is a new addition in the science of fusion where the graft is resorbed over time and replaced by host bone. The issue is the ability to preserve and maintain postoperative distraction, biomechanical stability and histological characteristics of intervertebral bone matrix formation. These implants create the extracellular matrix of bone.²²⁷ They generate an inflammatory response and have poor osteo-conductivity.²²⁸ To improve the osteo-conductivity, nano-sized β -tricalcium phosphate (β -TCP) has been incorporated into PLA cages.²²⁹ Poly (D,L-Lactide-co-Glycolide) (PLDLLA) Cage and Polymer Calcium phosphate-composite (PCC) Cage have been used in animal models as absorbable intervertebral implants and have given promising results.²³⁰

4. Self-Assembly of Peptide Amphiphiles

This is a new addition of bio-nanotechnology where peptide amphiphiles, a class of molecules that combine the structural features of amphiphilic surfactants with the functions of bioactive peptides, assemble into a variety of nanostructures. A specific type of peptide amphiphiles are known to self-assemble into one-dimensional (1D) nanostructures under physiological conditions, predominantly nanofibers with a cylindrical geometry. The resultant nanostructures could be highly bioactive and are of great interest in many biomedical applications, including tissue engineering, regenerative medicine and drug delivery.^{He B 2014} Reversible intramolecular disulfide bonds allow for cross-linking of nanofibers, resulting in a robust network that directs the mineralization of hydroxyapatite. The alignment of hydroxyapatite in the resulting composite material was found to be identical to the alignment observed between hydroxyapatite crystals and collagen fibrils in bone.²³¹ Using this foundation, phosphorylated serine segments within the PA molecules were incorporated, which allows for the generation of a self-supporting, bioactive gel matrix that mimics bone sialoprotein, further augmenting mineralization.¹⁵¹

5. Artificial intelligence (AI) and machine learning (ML)

Access to big, high fidelity clinical databases and the development of machine learning algorithms are making analysis and prediction a reality today. New technology using the newest advancements in machine learning and predictive analytics may offer significant clinical advantages in determining unique goals of correction to reduce the rates of pseudarthrosis, revision surgery, and proximal junctional failure.²³² Development of a validated computer-based preoperative predictive model for pseudarthrosis with 91% accuracy in adult spinal deformity is a step forward to decrease non-union. Clinical

oversight of “black box” algorithms to determine real-world practical application and interpretations in clinical settings is one of the limitations of machine learning. These tools hold the potential of aiding with improved diagnosis, surgical planning and risk optimization.

Conclusions

Spinal fusion has become an integral part of spine surgery. The evolution of fusion over the last 100 years has dramatically increased the safety and efficacy of this treatment. Much of this was driven by the advancement in instrumented and assisted fusion. Improvements in fusion rates, faster recovery times and reduction in complications are also due to better patient selection and peri-operative optimization of modifiable risk factors. Advances in material and design of fixation instrumentations, nanoscale surface technologies of structural grafts as well as use of biological agents for supporting and accelerating fusion are contributing to highly reliable rates of arthrodesis. All these advancement has made bone fusion almost a guaranteed event with very low complication rates. Artificial intelligence and machine learning aims to make fusion a more predictive procedure with a better prognosis helping patients to get a better quality of life.²³³ These technological advancement and results come with a huge financial burden to the community and the patients, and it is the surgeons' added responsibility to strike a good balance.

References

1. Don, A. S., & Carragee, E. A brief overview of evidence-informed management of chronic low back pain with surgery. *The Spine Journal*. 2008; 8(1), 258–265. doi:10.1016/j.spinee.2007.10.027
2. Last AR, Hulbert K. Chronic low back pain: evaluation and management. *Am Fam Phys*. 2009;79:1067–1074. DOI:10.1080/20786204.2010.10873969
3. Albee FH: Transplantation of a portion of the tibia into the spine for Pott's disease: A preliminary report 1911. *Clin Orthop Relat Res*. 2007;460:14-16.
4. Tarpada SP, Morris MT, Burton DA. Spinal fusion surgery: a historical perspective. *J Orthop*. 2016;14:134–136. doi: 10.1016/j.jor.2016.10.029
5. Rajae SS, Bae HW, Kanim LE, Delamarter RB. Spinal fusion in the United States: analysis of trends from 1998 to 2008. *Spine (Phila Pa 1976)*. 2012;37:67–76. doi:10.1097/BRS.0b013e31820cccfb.
6. Lange F, Peltier LF. Support for the spondylitic spine by means of buried steel bars, attached to the vertebrae. *Clin Orthop Relat Res*. 1986;203:3–6.

7. Boden, S. D., Schimandle, J. H., Hutton, W. C. An Experimental Lumbar Intertransverse Process Spinal Fusion Model. *Spine*. 1995; 20(supplement), 412420. doi:10.1097/00007632-199502001-00003
8. Marie-Jacqueline Reisener, Matthias Pumberger, Jennifer Shue, Federico P. Girardi, and Alexander P. Hughes. Trends in lumbar spinal fusion—a literature review. *J Spine Surg*. 2020 Dec; 6(4): 752–761. doi: 10.21037/jss-20-492 PMID: 33447679
9. Aghaloo, T., Pi-Anfruns, J., Moshaverinia, A., Sim, D., Grogan, T., Hadaya, D. The Effects of Systemic Diseases and Medications on Implant Osseointegration: A Systematic Review. *The International Journal of Oral & Maxillofacial Implants*. 2019; 34: s35–s49. doi:10.11607/jomi.19suppl.g3
10. Chen, H., Liu, N., Xu, X., Qu, X., & Lu, E. Smoking, Radiotherapy, Diabetes and Osteoporosis as Risk Factors for Dental Implant Failure: A Meta-Analysis. *PLoS ONE*. 2013; 8(8):e71955. doi:10.1371/journal.pone.00719
11. Robson H, Siebler T, Shalet SM, Williams GR. Interactions between GH, IGF-I, glucocorticoids and thyroid hormones during skeletal growth. *Pediatr Res*. 2002;52(2):137-147.
12. L G Raisz. Hormonal regulation of bone growth and remodelling. *Ciba Found Symp*. 1988;136:226-38. doi: 10.1002/9780470513637.ch14.
13. Slemenda CW, Reister TK, Hui SL, Miller JZ, Christian JC, Johnston CC Jr. Influences on skeletal mineralization in children and adolescents: evidence for varying effects of sexual maturation and physical activity. *J Pediatr*. 1994;125(2):201-207.
14. Park EA. The imprinting of nutritional disturbances on the growing bone. *Pediatrics*. 1964;33(suppl):815-862.
15. Golden, N. H., & Abrams, S. A. Optimizing Bone Health in Children and Adolescents. *Pediatrics*. 2014; 134(4):e1229–e1243. doi:10.1542/peds.2014-2173
16. Price, T C. Essential Nutrients for Bone Health and a Review of their Availability in the Average North American Diet. *The Open Orthopaedics Journal*. 2012;6(1):143–149. doi:10.2174/187432500120601014
17. <https://www.qualityhealth.org/bree/wp-content/uploads/sites/8/2019/01/Lumbar-Fusion-Bundle-and-Warranty-Final-2018.pdf>
18. Bonjour JP, Carrie AL, Ferrari S, Clavien H, Slosman D, Theintz G, Rizzoli R. Calcium-enriched foods and bone mass growth in prepubertal girls: A randomized, double-blind, placebo-controlled trial. *J Clin Invest*. 1997;99:1287–1294.
19. Johnston CC Jr, Miller JZ, Slemenda CW, Reister TK, Hui S, Christian JC, Peacock M. Calcium supplementation and increases in bone mineral density in children. *N Engl J Med*. 1992;327:82– 87.
20. NIH Consensus Development Panel on Osteoporosis Prevention, Diagnosis, and Therapy. *JAMA*. 2001;285(6):785–795. doi:10.1001/jama.285.6.785
21. Kohno M, Iwamura Y, Inasaka R, Kaneko K, Tomioka M, Kawai T, Inaba Y. Surgical Intervention for Osteoporotic Vertebral Burst Fractures in Middle-low Lumbar Spine with Special Reference to Postoperative Complications Affecting Surgical Outcomes. *Neurologia Medico-Chirurgica*. 2019;59(3):1-8. doi:10.2176/nmc.oa.2018-0232
22. Nakajima, H., Uchida, K., Honjoh, K., Sakamoto, T., Kitade, M., & Baba, H. Surgical treatment of low lumbar osteoporotic vertebral collapse: a single-institution experience. *Journal of Neurosurgery: Spine*. 2016; 24(1):39-47. doi:10.3171/2015.4.spine14847
23. Khosla, S., & Hofbauer, L. C. Osteoporosis treatment: recent developments and ongoing challenges. *The Lancet Diabetes & Endocrinology*. 2017;5(11):898–907. doi:10.1016/s2213-8587(17)30188-2
24. World Health Organization. Assessment of fracture risk and its application to screening for postmenopausal osteoporosis. Technical report series. Geneva: WHO; 1994.
25. Sai Chilakapati MS, Cody Eldridge BS, Syed I, Khalid MD, Owoicho Adogwa MD, MPH. In opioid naïve patients undergoing thoracolumbar spinal fusions, smoking increases odds for postoperative opioid use disorder. *The Spine Journal*. 2022;22(9) Supplement:2022, S82.
26. Dumont Aaron S, Verma, Subodh, Dumont, Randall J, Hurlbert, R. John. Nonsteroidal anti-inflammatory drugs and bone metabolism in spinal fusion. *Journal of pharmacological and toxicological methods*. *J Pharmacol Toxicol Methods*. 2000 Jan-Feb;43(1):31-9. doi: 10.1016/s1056-8719(00)00077-0.
27. Reuben, S S. Effect of nonsteroidal anti-inflammatory drugs on osteogenesis and spinal fusion. *Regional anesthesia and pain medicine*. *Reg Anesth Pain Med*. 2001 Nov-Dec;26(6):590-1. doi: 10.1053/rapm.2001.25927
28. Crandall, D. G., Wilson, M. R., Revella, J., Revella, D., McLemore, R. Smoking and Spinal Fusion with rhBMP-2: Long-Term Clinical, Functional and Occupational Outcomes. *The Spine Journal*. 2012;12(9), S92. doi:10.1016/j.spinee.2012.08.2
29. Lin YH, Wu CC, Huang YH, Hsu HC, Huang KC, Chang CH, Lin YC. The association between smoking and bone healing in orthopedic trauma patients.

- A systemic review and meta-analysis. *Journal of Orthopaedic Surgery and Research*.2021;16(1):160. doi:10.1186/s13018-021-02336-6
30. Thompson, L., Pardee, N., Stewart, J., & Scharf, J. Impact of smoking on fracture healing: a systematic review. *Journal of the American Academy of Orthopaedic Surgeons*. 2021;29(19):e954-e960. doi: 10.5435/JAAOS-D-21-00203
 31. Wu, J. P., Wu, L. J., Cui, J. Q., & Zhao, J. The effect of smoking on bone healing in spinal fusion surgery: a meta-analysis. *Journal of Orthopaedic Surgery and Research*. 2022;17(1):30. doi: 10.1186/s13018-022-02941-6
 32. Jackson KL 2nd, Devine JG. The effects of obesity on spine surgery: a systematic review of the literature. *Global Spine J*. 2016;6(4):394–400. doi: 10.1055/s-0035-1570750.
 33. V.T. Truong, T. Sunna, F. Al-Shakfa, M. Mc, Graw, G. Boubez, D. Shedid, S.-J. Yuh, Z. Wang. Impact of obesity on complications and surgical outcomes of adult degenerative scoliosis with long-segment spinal fusion. *Neurochirurgie*. 2022;68(3): 289-292.
 34. Welten DC, Kemper HC, Post GB, van Mechelen W, Twisk J, Lips P, Teule GJ. Weight-bearing activity during youth is a more important factor for peak bone mass than calcium intake. *J Bone Miner Res*. 1994;9:1089–1096.
 35. McGuigan FE, Murray L, Gallagher A, et al. Genetic and environmental determinants of peak bone mass in young men and women. *J Bone Miner Res*. 2002;17(7):1273-1279.
 36. Pocock NA, Eisman JA, Hopper JL, Yeates MG, Sambrook PN, Eberl S. Genetic determinants of bone mass in adults. A twin study. *J Clin Invest*. 1987;80:706–710.
 37. Ebrahimkhani, M., Arjmand, N., & Shirazi-Adl, A. Biomechanical effects of lumbar fusion surgery on adjacent segments using musculoskeletal models of the intact, degenerated and fused spine. *Scientific Reports*. 2021;11(1). doi:10.1038/s41598-021-97288-2
 38. Motta, M. M. da, Pratali, R. R., Coutinho, M. A. C., Hoffman, C. B., Barsotti, C. E. G., Santos, F. P. E. dos, & Oliveira, C. E. A. S. de. Correlation between obesity, sagittal balance and clinical outcome in spinal fusion. *Coluna/Columna*. 2015;14(3):186–189. doi:10.1590/s1808-185120151403140254
 39. Bolger, C., Bourlion, M., Leroy, X., Petit, D., Vanacker, G., McEvoy, L., & Nagaria, J. Maintenance of graft compression in the adult cervical spine. *European Spine Journal*. 2006;15(8):1204–1209. doi:10.1007/s00586-005-0054-z
 40. Alexander Spiessberger, Nicholas Dietz, Basil Erwin Gruter, Justin Virojanapa, Peter Hollis, and Ahmad Latefi. Junctional kyphosis and junctional failure after multi-segmental posterior cervicothoracic fusion – A retrospective analysis of 64 patients. *J Craniovertebr Junction Spine*. 2020 Oct-Dec;11(4):310–315. doi: 10.4103/jcvjs.JCVJS_177_20
 41. Tae-Kyum Kim, Wonik Cho, Sang Min Youn, Ung-Kyu Chang. The Effect of Perioperative Radiation Therapy on Spinal Bone Fusion Following Spine Tumor Surgery. *J Korean Neurosurg Soc*.2016;59(6):597-603. 10.3340/jkns.2016.59.6.597
 42. Elder, B. D., Ishida, W., Goodwin, C. R., Bydon, A., Gokaslan, Z. L., Sciubba, D. M. Bone graft options for spinal fusion following resection of spinal column tumors: systematic review and meta-analysis. *Neurosurgical Focus*.2017;42(1),E16. doi:10.3171/2016.8.focus16112
 43. Wallach, S. Paget’s disease and fibrous dysplasia. *Current Opinion in Rheumatology*. 1991;3(3):472–480. doi:10.1097/00002281-199106000-00022
 44. Tuli SK, Chen P, Eichler ME, Woodard EJ: Reliability of radiologic assessment of fusion: Cervical fibular allograft model. *Spine (Phila Pa 1976)*. 2004; 29(8):856-860. doi: 10.1097/00007632-200404150-00007.
 45. Farey ID, McAfee PC, Davis RF, Long DM. Pseudarthrosis of the cervical spine after anterior arthrodesis: Treatment by posterior nerve-root decompression, stabilization, and arthrodesis. *J Bone Joint Surg Am*. 1990;72(8):1171-1177. PMID: 2398087
 46. Kant AP, Daum WJ, Dean SM, Uchida T. Evaluation of lumbar spine fusion: Plain radiographs versus direct surgical exploration and observation. *Spine (Phila Pa 1976)*.1995;20(21):2313-2317.
 47. Fogel GR, Toohey JS, Neidre A, Brantigan JW. Fusion assessment of posterior lumbar interbody fusion using radiolucent cages: X-ray films and helical computed tomography scans compared with surgical exploration of fusion. *Spine J*. 2008;8(4):570-577. DOI:10.1016/J.SPINEE.2007.03.013
 48. Cannada LK, Scherping SC, Yoo JU, Jones PK, Emery SE. Pseudoarthrosis of the cervical spine: A comparison of radiographic diagnostic measures. *Spine (Phila Pa 1976)* 2003;28(1):46-51. doi: 10.1097/00007632-200301010-00012.
 49. Ghiselli G, Wharton N, Hipp JA, Wong DA, Jatana S. Prospective analysis of 33. imaging prediction of pseudoarthrosis after anterior cervical discectomy and fusion: Computed tomography versus flexion-extension motion analysis with intraoperative correlation. *Spine (Phila Pa 1976)* 2011;36(6):463-468. doi: 10.1097/BRS.0b013e3181d7a81a.
 50. Chafetz N, Cann CE, Morris JM, Steinbach LS, Goldberg HI, Ax L. Pseudoarthrosis following lumbar

- fusion: Detection by direct coronal CT scanning. *Radiology*.1987;162(3):803-805. doi: 10.1148/radiology.162.3.3809497.
51. Carreon LY, Glassman SD, Djurasovic M. Reliability and agreement between fine-cut CT scans and plain radiography in the evaluation of posterolateral fusions. *Spine J*.2007;7(1):39-43. doi: 10.1016/j.spinee.2006.04.005.
 52. Taylor M, Hipp JA, Gertzbein SD, Gopinath S, Reitman CA. Observer agreement in assessing flexion-extension X-rays of the cervical spine, with and without the use of quantitative measurements of intervertebral motion. *Spine J* 2007;7(6):654-658. doi: 10.1016/j.spinee.2006.10.017.
 53. Humadi, A., Dawood, S., Halldin, K., & Freeman, B. RSA in Spine: A Review. *Global Spine Journal*. 2017;7(8):811–820. doi:10.1177/2192568217701722
 54. Selby, M. D., Clark, S. R., Hall, D. J., & Freeman, B. J. C. Radiologic Assessment of Spinal Fusion. *Journal of the American Academy of Orthopaedic Surgeons*. 2012; 20(11): 694–703. doi:10.5435/jaaos-20-11-694.
 55. Auerbach, J. D., Ong, K. L., Lau, E., & Schmier, J. Perioperative Outcomes, Complications and Costs Associated with Lumbar Spinal Fusion in Older Patients with Spinal Stenosis and Spondylolisthesis: Analysis of the United States Medicare Claims Database. *The Spine Journal*. 2012;12(9), S3. doi:10.1016/j.spinee.2012.08.030
 56. Richter, A., Koutsoumbelis, S. A., Essig, D. A., & Silber, J. S. Comparison between Instrumented and Uninstrumented Posterolateral Fusion for Lumbar Spinal Stenosis and Spondylolisthesis. *The Spine Journal*. 2014;14(11), S52–S53. doi:10.1016/j.spinee.2014.08.138
 57. Sigmundsson, F. G., Jönsson, B., & Strömqvist, B. Outcome of decompression with and without fusion in spinal stenosis with degenerative spondylolisthesis in relation to preoperative pain pattern: a register study of 1,624 patients. *The Spine Journal*. 2015;15(4), 638–646. doi:10.1016/j.spinee.2014.11.020
 58. Sabry, I. H., Kabil, M. S., Mostafa, H. N., Ahmed, O. E., & Elshafei, K. M. M. Stand-Alone Open Disectomy versus Fixation in Management of Recurrent Lumbar Disc Prolapse. *QJM: An International Journal of Medicine*. 2020; 113(Supplement_1). <https://doi.org/10.1093/qjmed/hcaa054.019>
 59. Kamrul Ahsan, Shahidul Islam Khan, Naznin Zaman, Nazmin Ahmed, Nicola Montemurro, and Bipin Chaurasia. Fusion versus non-fusion treatment for recurrent lumbar disc herniation *J Craniovertebr Junction Spine*. 2021 Jan-Mar; 12(1): 44–53. doi: 10.4103/jcvjs.JCVJS_153_20, PMID: PMC8035587, PMID: 33850381
 60. Nooraie, H., Taghipour, M., & Arasteh, M. M. Lumbar spine fusion and pedicle fixation with C-D screws for lumbar iatrogenic instability. *Archives of Orthopaedic and Trauma Surgery*. 1997;116(4), 236–238. doi:10.1007/bf00393719
 61. Bevevino, A. J., Helgeson, M. D., & Albert, T. J. Iatrogenic spinal instability: Cervical and thoracic spine. *Seminars in Spine Surgery*. 2013; 25(2):119-130. doi:10.1053/j.semss.2013.03.006
 62. Kothe, R., Panjabi, M. M., & Liu, W. Multidirectional Instability of the Thoracic Spine Due to Iatrogenic Pedicle Injuries During Transpedicular Fixation. *Spine*. 1997;22(16):1836–1842. doi:10.1097/00007632-199708150-00008
 63. WU, H., YU, W.-D., JIANG, R., & GAO, Z.-L. Treatment of multilevel degenerative lumbar spinal stenosis with spondylolisthesis using a combination of microendoscopic discectomy and minimally invasive transforaminal lumbar interbody fusion. *Experimental and Therapeutic Medicine*. 2012; 5(2) 567–571. doi:10.3892/etm.2012.812
 64. Kulkarni, A. G. Degenerative Spondylolisthesis. *Clin Spine Surg*. 2020 Oct;33(8):E391-E400. doi: 10.1097/BSD.0000000000000970.
 65. DeWald, C. J., & Stanley, T. Instrumentation-Related Complications of Multilevel Fusions for Adult Spinal Deformity Patients Over Age 65. *Spine*. 2006; 31(Suppl):S144–S151. doi:10.1097/01.brs.0000236893.65878.39
 66. Lehman RA Jr, Kang DG, Wagner SC. Management of osteoporosis in spine surgery. *J Am Acad Orthop Surg*.2015;23(4):253– 263. doi: 10.5435/JAAOS-D-14-00042.
 67. Hilibrand AS, et al. Radiculopathy and myelopathy at segments adjacent to the site of a previous anterior cervical arthrodesis. *J Bone Joint Surg Am*. 1999;81(4):519–528. doi: 10.2106/00004623-199904000-00009.
 68. Khan NR, Clark AJ, Lee SL, Venable GT, Rossi NB, Foley KT. Surgical outcomes for minimally invasive vs open transforaminal lumbar interbody fusion: an updated systematic review and meta- analysis. *Neurosurgery*.2015;77(6):847-874. doi: doi.org/10.1227/NEU.0000000000000913
 69. Ou C-Y, Lee T-C, Lee T-H, Huang Y-H. Impact of body mass index on adjacent segment disease after lumbar fusion for degenerative spine disease. *Neurosurgery*. 2015;76(4):396–402. doi.org/10.1227/NEU.0000000000000627
 70. Oda I, et al. Does spinal kyphotic deformity influence the bio- mechanical characteristics of the adjacent motion segments? An in vivo animal model. *Spine (Phila Pa 1976)*. 1999;24(20):2139– 2146. doi: 10.1097/00007632-199910150-00014.

71. Phan, K., Nazareth, A., Hussain, A. K., Dmytriw, A. A., Nambiar, M., Nguyen, D., ... Mobbs, R. J. Relationship between sagittal balance and adjacent segment disease in surgical treatment of degenerative lumbar spine disease: meta-analysis and implications for choice of fusion technique. *European Spine Journal*. 2018;27(8),1981–1991. doi:10.1007/s00586-018-5629-6
72. Kim HJ, Iyer S. Proximal junctional kyphosis. *J Am Acad Orthop Surg*. 2016;24:318–326. DOI: 10.5435/JAAOS-D-14-00393
73. Kim YJ, Bridwell KH, Lenke LG, Cho KJ, Edwards CC 2nd, Rinella AS. Pseudarthrosis in adult spinal deformity following multisegmental instrumentation and arthrodesis. *J Bone Joint Surg Am*. 2006;88:721–728. doi: 10.2106/JBJS.E.00550.
74. Kelly MP, Savage JW, Bentzen SM, Hsu WK, Ellison SA, Anderson PA. Cancer risk from bone morphogenetic protein exposure in spinal arthrodesis. *J Bone Joint Surg Am*. 2014;96:1417–1422. doi: 10.2106/JBJS.M.01190.
75. Nasser R, Yadla S, Maltenfort MG., Complications in spine surgery. *J Neurosurg Spine*. 2010;13:144–157. doi: 10.3171/2010.3.SPINE09369.
76. O’Shaughnessy BA, Bridwell KH, Lenke LG, et al. Does a long- fusion “T3-sacrum” portend a worse outcome than a short-fusion “T10-sacrum” in primary surgery for adult scoliosis? *Spine (Phila Pa 1976)*. 2012;37:884–890. doi: 10.1097/BRS.0b013e3182376414.
77. Shriver MF, Lewis DJ, Kshetry VR, Rosenbaum BP, Benzel EC, Mroz TE. Pseudoarthrosis rates in anterior cervical discectomy and fusion: a meta-analysis. *Spine J*. 2015;15(9):2016–2027. doi: 10.1016/j.spinee.2015.05.010.
78. Casper, D. S., Zmistowski, B., Hollern, D. A., Hilibrand, A. S., Vaccaro, A. R., Schroeder, G. D., & Kepler, C. K. The Effect of Postoperative Spinal Infections on Patient Mortality. *Spine*. 2018; 43(3), 223–227. doi:10.1097/brs.0000000000002277
79. Qin H, Cao H, Zhao Y, et al. In vitro and in vivo anti-biofilm effects of silver nanoparticles immobilized on titanium. *Biomaterials*. 2014;35(33):9114–9125. doi: 10.1016/j.biomaterials.2014.07.040.
80. Colon, G., Ward, B. C., & Webster, T. J. Increased osteoblast and decreased Staphylococcus epidermidis functions on nanophase ZnO and TiO₂. *Journal of Biomedical Materials Research Part A*. 2006; 78A(3), 595–604. doi:10.1002/jbm.a.30789
81. Wang, J. C., Hart, R. A., Emery, S. E., & Bohlman, H. H. Graft Migration or Displacement After Multilevel Cervical Corpectomy and Strut Grafting. *Spine*. 2003;28(10), 1016-1021. doi:10.1097/01.brs.0000061998.62204.d7
82. Adam P. Myhre, Todd J. Jarosz, John C. Hunter, Michael L. Richardson. Postoperative Bone Graft Displacement: An Unusual Sign of Infection Following Posterior Spinal Fusion. *Radiology Case Reports*.2006; 1(1): 21-23 <https://doi.org/10.2484/rcr.v1i1.9>
83. Park Y, et al. Comparison of anterior cervical fusion after two- level discectomy or single-level corpectomy: sagittal alignment, cervical lordosis, graft collapse, and adjacent-level ossification. *Spine J*. 2010;10(3):193–199. doi: 10.1016/j.spinee.2009.09.006.
84. Findlay, G. F. G. The failed back syndrome: Etiology and therapy. *Current Orthopaedics*. 1994;8(1), 22. doi:10.1016/02680890(94)90007-8
85. Glassman SD, Bridwell K, Dimar JR, Horton W, Berven S, Schwab F. The impact of positive sagittal balance in adult spinal deformity. *Spine (Phila Pa 1976)*. 2005;30:2024–2029. doi: 10.1097/01.brs.0000179086.30449.96.
86. Schwab FJ, Blondel B, Bess S, et al. Radiographical spinopelvic parameters and disability in the setting of adult spinal deformity: a prospective multicenter analysis. *Spine (Phila Pa 1976)*. 2013;38:e803–e812. doi: 10.1097/BRS.0b013e318292b7b9.
87. Lafage R, Schwab F, Glassman S, et al. Age-adjusted alignment goals have the potential to reduce PJK. *Spine (Phila Pa 1976)*. 2017;42:1275–1282. doi: 10.1097/BRS.0000000000002146.
88. Hilibrand AS, et al. Radiculopathy and myelopathy at segments adjacent to the site of a previous anterior cervical arthrodesis. *J Bone Joint Surg Am*. 1999;81(4):519–528. doi: 10.2106/00004623-199904000-00009.
89. Komura S, et al. Lower incidence of adjacent segment degeneration after anterior cervical fusion found with those fusing C5-6 and C6-7 than those leaving C5-6 or C6-7 as an adjacent level. *J Spinal Disord Tech*. 2012;25(1):23–29. doi: 10.1097/BSD.0b013e31820bb1f8.
90. Arutyunyan, G. G., Angevine, P. D., & Berven, S. Cost-Effectiveness in Adult Spinal Deformity Surgery. *Neurosurgery*. 2018; Oct 1;83(4):597-601. doi: 10.1093/neuros/nyx575.
91. Tsuchiya K, Bridwell KH, Kuklo TR, Lenke LG, Baldus C. Minimum 5-year analysis of L5-S1 fusion using sacropelvic fixation (bilateral S1 and iliac screws) for spinal deformity. *Spine (Phila Pa 1976)*. 2006;31:303–308. *Spine (Phila Pa 1976)* doi: 10.1097/01.brs.0000197193.81296.fl.
92. Harimaya K, Mishiro T, Lenke LG, Bridwell KH, Koester LA, Sides BA. Etiology and revision surgical strategies in failed lumbosacral fixation of adult spinal deformity constructs. *Spine (Phila*

- Pa 1976). 2011;36:1701–1710. doi: 10.1097/BRS.0b013e3182257eaf.
93. Kim YJ, Bridwell KH, Lenke LG, Rinella AS, Edwards C 2nd. Pseudarthrosis in primary fusions for adult idiopathic scoliosis: incidence, risk factors, and outcome analysis. *Spine (Phila Pa 1976)*. 2005;30:468–474. doi: 10.1097/01.brs.0000153392.74639.ea.
 94. Shen FH, Harper M, Foster WC, Marks I, Arlet V. A novel “four-rod technique” for lumbo-pelvic reconstruction: theory and technical considerations. *Spine (Phila Pa 1976)*. 2006;31:1395–1401
 95. Hyun SJ, Lenke LG, Kim YC, Koester LA, Blanke KM. Comparison of standard 2-rod constructs to multiple-rod constructs for fixation across 3-column spinal osteotomies. *Spine (Phila Pa 1976)* 2014;39:1899–1904. doi: [10.1177/2192568217699392](https://doi.org/10.1177/2192568217699392)
 96. Merrill RK, Kim JS, Leven DM, Kim JH, Cho SK. Multi-rod constructs can prevent rod breakage and pseudarthrosis at the lumbosacral junction in adult spinal deformity. *Global Spine J*. 2017;7:514–520. doi: [10.1177/2192568217699392](https://doi.org/10.1177/2192568217699392).
 97. Han S, Hyun SJ, Kim KJ, Jahng TA, Lee S, Rhim SC. Rod stiffness as a risk factor of proximal junctional kyphosis after adult spinal deformity surgery: comparative study between cobalt chrome multiple-rod constructs and titanium alloy two-rod constructs. *Spine J*. 2017;17:962–968. doi: [10.1016/j.spinee.2017.02.005](https://doi.org/10.1016/j.spinee.2017.02.005).
 98. Goffin J, et al. Long-term results after anterior cervical fusion and osteosynthetic stabilization for fractures and/or dislocations of the cervical spine. *J Spinal Disord*. 1995;8(6):500–508. PMID: 8605425.
 99. Oda I, et al. Does spinal kyphotic deformity influence the bio-mechanical characteristics of the adjacent motion segments? An in vivo animal model. *Spine (Phila Pa 1976)*. 1999;24(20):2139–2146. doi: [10.1097/00007632-199910150-00014](https://doi.org/10.1097/00007632-199910150-00014).
 100. Guzman JZ, et al. Osteoporosis in cervical spine surgery. *Spine (Phila Pa 1976)*. 2016;41(8):662–668. doi: [10.1097/BRS.0000000000001347](https://doi.org/10.1097/BRS.0000000000001347).
 101. DeWald, C. J., & Stanley, T. Instrumentation-Related Complications of Multilevel Fusions for Adult Spinal Deformity Patients Over Age 65. *Spine*. 2006; 31(Suppl), S144–S151. doi: [10.1097/01.brs.0000236893.65878.39](https://doi.org/10.1097/01.brs.0000236893.65878.39)
 102. Lehman RA Jr, Kang DG, Wagner SC. Management of osteoporosis in spine surgery. *J Am Acad Orthop Surg*. 2015;23(4):253–263. doi: [10.5435/JAAOS-D-14-00042](https://doi.org/10.5435/JAAOS-D-14-00042).
 103. Cheng, H., Gary, L. C., Curtis, J. R., Saag, K. G., Kilgore, M. L., Morrissey, M. A., ... Delzell, E. Estimated prevalence and patterns of presumed osteoporosis among older Americans based on Medicare data. *Osteoporosis International*. 2009; 20(9), 1507–1515. doi: [10.1007/s00198-009-0835-z](https://doi.org/10.1007/s00198-009-0835-z)
 104. Clynes, M. A., Harvey, N. C., Curtis, E. M., Fuggle, N. R., Dennison, E. M., & Cooper, C. The epidemiology of osteoporosis. *British Medical Bulletin*. 2020. doi: [10.1093/bmb/ldaa005](https://doi.org/10.1093/bmb/ldaa005)
 105. Bonjour JP, Carrie AL, Ferrari S, Clavien H, Slosman D, Theintz G, Rizzoli R. Calcium-enriched foods and bone mass growth in prepubertal girls: A randomized, double-blind, placebo-controlled trial. *J Clin Invest*. 1997; 99:1287–1294.
 106. Johnston CC Jr, Miller JZ, Slemenda CW, Reister TK, Hui S, Christian JC, Peacock M. Calcium supplementation and increases in bone mineral density in children. *N Engl J Med*. 1992; 327:82–87.
 107. Khosla, S., & Hofbauer, L. C. Osteoporosis treatment: recent developments and ongoing challenges. *The Lancet Diabetes & Endocrinology*. 2017; 5(11), 898–907. doi: [10.1016/s2213-8587\(17\)30188-2](https://doi.org/10.1016/s2213-8587(17)30188-2)
 108. Holick MF. Vitamin D deficiency. *N Engl J Med*. 2007;357(3):266–281. doi: [10.1056/NEJMra070553](https://doi.org/10.1056/NEJMra070553).
 109. DeWald, C. J., & Stanley, T. Instrumentation-Related Complications of Multilevel Fusions for Adult Spinal Deformity Patients Over Age 65. *Spine*. 2006; 31(Suppl), S144–S151. doi: [10.1097/01.brs.0000236893.65878.39](https://doi.org/10.1097/01.brs.0000236893.65878.39)
 110. Carlson BC, Robinson WA, Wanderman NR, et al. A review and clinical perspective of the impact of osteoporosis on the spine. *Geriatr Orthop Surg Rehabil*. 2019;10:1-8. doi: <https://doi.org/10.1177/2151459319861591>
 111. Andreassen, T. T., Fledelius, C., Ejersted, C., & Oxlund, H. Increases in callus formation and mechanical strength of healing fractures in old rats treated with parathyroid hormone. *Acta Orthopaedica Scandinavica*. 2001; 72(3), 304–307. doi: [10.1080/00016470152846673](https://doi.org/10.1080/00016470152846673)
 112. Rosen CJ. Clinical practice. Vitamin D insufficiency. *N Engl J Med*. 2011;364(3):248–254. doi: [10.1056/NEJMc1009570](https://doi.org/10.1056/NEJMc1009570).
 113. Fischer CR, et al. A systematic review of treatment strategies for degenerative lumbar spine fusion surgery in patients with osteoporosis. *Geriatr Orthop Surg Rehabil*. 2016;7(4):188–196. doi: [10.1177/2151458516669204](https://doi.org/10.1177/2151458516669204).
 114. Kleerekoper M, et al. Assessing the effects of teriparatide treatment on bone mineral density, bone microarchitecture, and bone strength. *J Bone Joint Surg Am*. 2014;96(11):e90. doi: [10.2106/JBJS.L.01757](https://doi.org/10.2106/JBJS.L.01757).
 115. Polikeit A, et al. The importance of the endplate for interbody cages in the lumbar spine. *Eur Spine J*. 2003;12(6):556–561. doi: [10.1007/s00586-003-0556-5](https://doi.org/10.1007/s00586-003-0556-5)

116. Ohe M, Moridaira H, Inami S, Takeuchi D, Nohara Y, Taneichi H. Pedicle screws with a thin hydroxyapatite coating for improving fixation at the bone-implant interface in the osteoporotic spine: experimental study in a porcine model. *J Neurosurg Spine*. 2018;28(6):679–687. doi: 10.3171/2017.10.SPINE17702.
117. Samartzis D, Shen FH, Matthews DK, Yoon ST, Goldberg EJ, An HS. Comparison of allograft to autograft in multilevel anterior cervical discectomy and fusion with rigid plate fixation. *Spine J*. 2003;3:451–459. doi: 10.1016/s1529-9430(03)00173-6.
118. Roberts TT, Rosenbaum AJ. Bone grafts, bone substitutes and orthobiologics: the bridge between basic science and clinical advancements in fracture healing. *Organogenesis*. 2012;8:114–124. doi: 10.4161/org.23306.
119. Buser, Z., Brodke, D. S., Youssef, J. A., Meisel, H.-J., Myhre, S. L., Hashimoto, R., ... Wang, J. C. Synthetic bone graft versus autograft or allograft for spinal fusion: a systematic review. *Journal of Neurosurgery. Spine*. 2016; 25(4), 509–516. doi:10.3171/2016.1.spine151005
120. Dimar, J. R., Glassman, S. D., Burkus, J. K., Pryor, P. W., Hardacker, J. W., & Carreon, L. Y. Two-year fusion and clinical outcomes in 224 patients treated with a single-level instrumented posterolateral fusion with iliac crest bone graft. *The Spine Journal*. 2009; 9(11), 880–885. doi:10.1016/j.spinee.2009.03.013
121. Grabowski G, Cornett CA. Bone graft and bone graft substitutes in spine surgery: current concepts and controversies. *J Am Acad Orthop Surg*. 2013;21:51–60. doi: 10.5435/JAAOS-21-01-51.
122. Rihn JA, Kirkpatrick K, Albert TJ. Graft options in posterolateral and posterior interbody lumbar fusion. *Spine (Phila Pa 1976)*. 2010;35:1629–1639. doi: 10.1097/BRS.0b013e3181d25803.
123. Ahlmann E, Patzakis M, Roidis N, Shepherd L, Holtom P. Comparison of anterior and posterior iliac crest bone grafts in terms of harvest-site morbidity and functional outcomes. *J Bone Joint Surg Am*. 2002; 84:716–720. Doi: 10.2106/00004623-200205000-00003.
124. Ebraheim NA, Elgafy H, Xu R. Bone-graft harvesting from iliac and fibular donor sites: techniques and complications. *J Am Acad Orthop Surg*. 2001;9:210–218. doi: 10.5435/00124635-200105000-00007.
125. Khan SN, Tomin E, Lane JM. Clinical applications of bone graft substitutes. *Orthop Clin North Am*. 2000;31:389–398. doi: 10.1016/s0030-5898(05)70158-9.
126. Schnee CL, Freese A, Weil RJ, Marcotte PJ. Analysis of harvest morbidity and radiographic outcome using autograft for anterior cervical fusion. *Spine (Phila Pa 1976)*. 1997;22:2222–2227. doi: 10.1097/00007632-199710010-00005.
127. Khan SN, Cammisa FP, Jr., Sandhu HS, Diwan AD, Girardi FP, Lane JM. The biology of bone grafting. *J Am Acad Orthop Surg*. 2005;13:77–86. PMID: 15712985
128. Tomford WW. Bone allografts: past, present and future. *Cell Tissue Bank*. 2000;1:105-109. doi: 10.1023/A:1010158731885.
129. Samartzis D, Shen FH, Matthews DK, Yoon ST, Goldberg EJ, An HS. Comparison of allograft to autograft in multilevel anterior cervical discectomy and fusion with rigid plate fixation. *Spine J*. 2003;3:451–459. doi: 10.1016/s1529-9430(03)00173-6.
130. Miller LE, Block JE. Safety and effectiveness of bone allografts in anterior cervical discectomy and fusion surgery. *Spine (Phila Pa 1976)*. 2011;36:2045–2050. doi: 10.1097/BRS.0b013e3181ff37eb.
131. Smith KA, Russo GS, Vaccaro AR, Arnold PM. Scientific, Clinical, regulatory, and economic aspects of choosing bone graft/ biological options in spine surgery. *Neurosurgery*. 2018;84:827–835. doi: 10.1093/neuros/nyy322.
132. He B, Ou Y, Zhou A, et al. Functionalized D-form self-assembling peptide hydrogels for bone regeneration. *Drug Des Devel Ther*. 2016;10:1379–1388. doi: 10.2147/DDDT.S97530
133. Skovrlj B, Guzman JZ, Al Maaieh M, Cho SK, Iatridis JC, Qureshi SA. Cellular bone matrices: viable stem cell-containing bone graft substitutes. *Spine J*. 2014;14:2763-2772. doi: 10.1016/j.spinee.2014.05.024
134. O'Loughlin PF, Morr S, Bogunovic L, Kim AD, Park B, Lane JM. Selection and development of preclinical models in fracture-healing research. *J Bone Joint Surg Am*. 2008;90 Suppl 1:79–84. doi: 10.1097/BRS.0000000000002441.
135. Gupta A, Kukkar N, Sharif K, Main BJ, Albers CE, El-Amin III SF. Bone graft substitutes for spine fusion: A brief review. *World J Orthop*. 2015;6:449–456. doi: 10.5312/wjo.v6.i6.449.
136. Polo-Corrales L, Latorre-Esteves M, Ramirez-Vick JE. Scaffold design for bone regeneration. *J Nanosci Nanotechnol*. 2014;14:15–56. doi: 10.1166/jnn.2014.9127.
137. Henkel J, Woodruff MA, Epari DR, et al. Bone regeneration based on tissue engineering conceptions—a 21st century perspective. *Bone Res*. 2013;1:216–248. doi: 10.4248/BR201303002.
138. Tsai, PI., Wu, MH., Li, YY. et al. Additive-manufactured Ti-6Al-4V/Polyetheretherketone composite porous cage for Interbody fusion: bone

- growth and biocompatibility evaluation in a porcine model. *BMC Musculoskelet Disord.* 2021; 22: 171. <https://doi.org/10.1186/s12891-021-04022-0>
139. Kingsley R Chin, Nishant N Gohel, Daniel M Aloise, Jason A Seale, Deepak K Pandey, Fabio J Pencle. Effectiveness of a Fully Impregnated Hydroxyapatite Polyetheretherketone Cage on Fusion in Anterior Cervical Spine Surgery. *Cureus.* 2021 Aug 26;13(8):e17457. doi: 10.7759/cureus.17457.
 140. Epstein NE. Efficacy of posterior cervical fusions utilizing an artificial bone graft expander, beta tricalciumphosphate. *Surg Neurol Int.* 2011;2:15. doi: 10.4103/2152-7806.76458
 141. Epstein NE. Preliminary documentation of the comparable efficacy of vitoss versus NanOss bioactive as bone graft expanders for posterior cervical fusion. *Surg Neurol Int.* 2015;6(Suppl 4): S164–S171. doi: 10.4103/2152-7806.156559.
 142. Dai, L.-Y., & Jiang, L.-S. Anterior cervical fusion with interbody cage containing β -tricalcium phosphate augmented with plate fixation: a prospective randomized study with 2-year follow-up. *European Spine Journal.* 2008; 17(5): 698–705. doi:10.1007/s00586-008-0643-8
 143. Sugawara T, et al. β -tricalcium phosphate promotes bony fusion after anterior cervical discectomy and fusion using titanium cages. *Spine (Phila Pa 1976).* 2011; 36(23): E1509–E1514. doi: 10.1097/BRS.0b013e31820e60d9.
 144. LeGeros RZ. Properties of osteoconductive biomaterials: calcium phosphates. *Clin Orthop Relat Res.* 2002;395:81–98. doi: 10.1097/00003086-200202000-00009.
 145. Khan AF, et al. Bioactive behaviour of silicon substituted calcium phosphate based bioceramics for bone regeneration. *Mater Sci Eng C Mater Biol Appl.* 2014;35:245–252. doi: 10.1016/j.msec.2013.11.013.
 146. Gazdag AR, Lane JM, Glaser D, Forster RA. Alternatives to Autogenous Bone Graft: Efficacy and Indications. *J Am Acad Orthop Surg.* 1995;3(1):1–8. doi: 10.5435/00124635-199501000-00001.
 147. Friesenbichler J, Maurer-Ertl W, Sadoghi P, Pirker-Fruehauf U, Bodo K, Leithner A. Adverse reactions of artificial bone graft substitutes: lessons learned from using tricalcium phosphate geneX[®]. *Clin Orthop Relat Res.* 2014;472:976–982. doi: 10.1007/s11999-013-3305-z.
 148. Kumar CY, Nalini KB, Menon J, Patro DK, Banerji BH. Calcium sulfate as bone graft substitute in the treatment of osseous bone defects, a prospective study. *J Clin Diagn Res.* 2013;7:2926–2928. doi: 10.7860/JCDR/2013/6404.3791
 149. Helm G, Anderson DG, Andersson GB, et al. Summary statement: bone morphogenetic proteins: basic science. *Spine (Phila Pa 1976).* 2002;27:S9. doi: 10.2106/00004623-200100001-00001.
 150. He B, Ou Y, Zhou A, et al. Functionalized D-form self-assembling peptide hydrogels for bone regeneration. *Drug Des Devel Ther.* 2016;10:1379–1388. doi: 10.2147/DDDT.S97530
 151. Mata A, Geng Y, Henrikson KJ, Aparicio C, Stock SR, Satcher RL, Stupp SIJB. Bone regeneration mediated by biomimetic mineralization of a nanofiber matrix. *Biomaterials.* 2010;31:6004–6012. doi: 10.1016/j.biomaterials.2010.04.013.
 152. Vo TN, Shah SR, Lu S, Tataru AM, Lee EJ, Roh TT, Tabata Y, Mikos AG. Injectable dual-gelling cell-laden composite hydrogels for bone tissue engineering. *Biomaterials.* 2016;83:1–11. doi: 10.1016/j.biomaterials.2015.12.026.
 153. Kissling S, Seidenstuecker M, Pilz IH, Suedkamp NP, Mayr HO, Bernstein A. Sustained release of rhBMP-2 from microporous tricalciumphosphate using hydrogels as a carrier. *BMC Biotechnol.* 2016;16(1):44. DOI:10.1186/s12896-016-0275-8
 154. Ozkaynak E, Rueger DC, Drier EA, et al. OP-1 cDNA encodes an osteogenic protein in the TGF-beta family. *EMBO J.* 1990;9:2085–2093 Park JH, Bae YK, Suh SW, Yang JH, Hong JY. Efficacy of cortico/cancellous composite allograft in treatment of cervical spondylosis. *Medicine (Baltimore).* 2017;96:e7803. doi: 10.1002/j.1460-2075.1990.tb07376.x.
 155. Wozney JM, Rosen V, Celeste AJ, et al. Novel regulators of bone formation: molecular clones and activities. *Science.* 1988;242:1528–1534. doi: 10.1126/science.3201241.
 156. Urist MR, Mikulski A, Lietze A. Solubilized and insolubilized bone morphogenetic protein. *Proc Natl Acad Sci U S A.* 1979;76:1828–1832. doi: 10.1073/pnas.76.4.1828.
 157. Ebara S, Nakayama K. Mechanism for the action of bone morphogenetic proteins and regulation of their activity. *Spine (Phila Pa 1976).* 2002;27:S10–15. doi: 10.1097/00007632-200208151-00004.
 158. Helm G, Anderson DG, Andersson GB, et al. Summary statement: bone morphogenetic proteins: basic science. *Spine (Phila Pa 1976).* 2002;27:S9. doi: 10.2106/00004623-200100001-00001.
 159. Boden, S. D., Zdeblick, T. A., Sandhu, H. S., & Heim, S. E. The Use of rhBMP-2 in Interbody Fusion Cages. *Spine.* 2000; 25(3), 376–381. doi:10.1097/00007632-200002010-00020
 160. Cunningham, B. W., Kanayama, M., Parker, L. M., Weis, J. C., Seftor, J. C., Fedder, I. L., & McAfee, P. C. Osteogenic Protein Versus Autologous Interbody Arthrodesis in the Sheep Thoracic Spine. *Spine.* 1999; 24(6), 509–518. doi:10.1097/00007632-

- 199903150-00002
161. David, S. M., Gruber, H. E., Meyer, R. A., Murakami, T., Tabor, O. B., Howard, B. A., ... Hanley, E. N. Lumbar Spinal Fusion Using Recombinant Human Bone Morphogenetic Protein in the Canine. *Spine*. 1999; 24(19), 1973. doi:10.1097/00007632-199910010-00002
 162. Silcox DH, 3rd, Boden SD, Schimandle JH, Johnson P, Whitesides TE, Hutton WC. Reversing the inhibitory effect of nicotine on spinal fusion using an osteoinductive protein extract. *Spine (Phila Pa 1976)* 1998;23:291–296; discussion 297. doi: 10.1097/00007632-199802010-00001.
 163. Simmonds MC, Brown JV, Heirs MK, et al. Safety and effectiveness of recombinant human bone morphogenetic protein-2 for spinal fusion: a meta-analysis of individual-participant data. *Ann Intern Med*. 2013;158:877–889. doi: 10.7326/0003-4819-158-12-201306180-00005.
 164. Wozney JM. Overview of bone morphogenetic proteins. *Spine (Phila Pa 1976)*. 2002;27:S2-S8. doi: 10.1097/00007632-200208151-00002.
 165. Wang JC, Alanay A, Mark D, Kanim LEA, Campbell PA, Dawson EG, Lieberman JR. A comparison of commercially available demineralized bone matrix for spinal fusion. *Euro Spine J*. 2007;16:1233–1240. doi: 10.1007/s00586-006-0282-x
 166. Wang M, Lam RW, Abbah SA, et al. Heparin-based poly-electrolyte complex enhances the therapeutic efficacy of bone morphogenetic protein-2 for posterolateral fusion in a large animal model. *Spine (Phila Pa 1976)*. 2016;41:1199–1207. doi: 10.1097/BRS.0000000000001543.
 167. Cammisa, F. P., Lowery, G., Garfin, S. R., Geisler, F. H., Klara, P. M., McGuire, R. A., Block, J. E. Two-Year Fusion Rate Equivalency Between Grafton® DBM Gel and Autograft in Posterolateral Spine Fusion. *Spine*. 2004; 29(6):660–666. doi:10.1097/01.brs.0000116588.17129.b9
 168. Vaccaro AR, Stubbs HA, Block JE. Demineralized bone matrix composite grafting for posterolateral spinal fusion. *Orthopedics*. 2007;30:567–570. doi: 10.3928/01477447-20070701-06.
 169. Buser, Z., Brodke, D. S., Youssef, J. A., Rometsch, E., Park, J.-B., Yoon, S. T., Meisel, H.-J. Allograft Versus Demineralized Bone Matrix in Instrumented and Noninstrumented Lumbar Fusion: A Systematic Review. *Global Spine Journal*. 2017; 8(4): 396–412. doi:10.1177/2192568217735342
 170. Poorman GW, Jalai CM, Boniello A, Worley N, McClelland S, 3rd, Passias PG. Bone morphogenetic protein in adult spinal deformity surgery: a meta-analysis. *European Spine J*. 2017;26:2094–2102. doi: 10.1007/s00586-016-4841-5.
 171. Kim HJ, Buchowski JM, Zebala LP, Dickson DD, Koester L, Bridwell KH. rhBMP-2 is superior to iliac crest bone graft for long fusions to the sacrum in adult spinal deformity: 4- to 14-year follow-up. *Spine (Phila Pa 1976)*. 2013;38:1209–1215. doi: 10.1097/BRS.0b013e31828b656d.
 172. Seeherman HJ, Li XJ, Bouxsein ML, Wozney JM. rhBMP-2 induces transient bone resorption followed by bone formation in a nonhuman primate core-defect model. *J Bone Joint Surg Am*. 2010;92:411–426. doi: 10.2106/JBJS.H.01732.
 173. Carragee, E. J., Hurwitz, E. L., & Weiner, B. K. A critical review of recombinant human bone morphogenetic protein-2 trials in spinal surgery: emerging safety concerns and lessons learned. *The Spine Journal*. 2011; 11(6), 471–491. doi:10.1016/j.spinee.2011.04.023
 174. Simmonds MC, Brown JV, Heirs MK, et al. Safety and effectiveness of recombinant human bone morphogenetic protein-2 for spinal fusion: a meta-analysis of individual-participant data. *Ann Intern Med*. 2013;158:877–889. doi: 10.7326/0003-4819-158-12-201306180-00005.
 175. Vaidya R, Carp J, Sethi A, Bartol S, Craig J, Les CM. Complications of anterior cervical discectomy and fusion using recombinant human bone morphogenetic protein-2. *Euro Spine J*. 2007;16:1257–1265. doi: 10.1007/s00586-007-0351-9.
 176. McClellan JW, Mulconrey DS, Forbes RJ, Fullmer N. Vertebral bone resorption after transforaminal lumbar interbody fusion with bone morphogenetic protein (rhBMP-2). *J Spinal Disord Tech*. 2006;19:483–486. doi: 10.1097/01.bsd.0000211231.83716.4b.
 177. McKay B, Sandhu HS. Use of recombinant human bone morphogenetic protein-2 in spinal fusion applications. *Spine (Phila Pa 1976)*. 2002;27:s66–s85. doi: 10.1097/00007632-200208151-00014.
 178. Benglis, D., Wang, M. Y., & Levi, A. D. A comprehensive review of the safety profile of bone morphogenetic protein in spine surgery. *Operative Neurosurgery*. 2008; 62, ONS423–ONS431. doi:10.1227/01.neu.0000326030.24220.d8
 179. Burkus, J. K. Use of rhBMP-2 in Combination with Structural Cortical Allografts: Clinical and Radiographic Outcomes in Anterior Lumbar Spinal Surgery. *The Journal of Bone and Joint Surgery (American)*. 2005; 87(6), 1205. doi:10.2106/jbjs.d.02532.
 180. Cahill, K. S. Prevalence, Complications, and Hospital Charges Associated With Use of Bone-Morphogenetic Proteins in Spinal Fusion Procedures. *JAMA*. 2009; 302(1), 58. doi:10.1001/jama.2009.956
 181. Wang JC, Alanay A, Mark D, Kanim LEA, Campbell PA, Dawson EG, Lieberman JR. A comparison of

- commercially available demineralized bone matrix for spinal fusion. *Euro Spine J.* 2007;16:1233–1240. doi: 10.1007/s00586-006-0282-x
182. Wang M, Lam RW, Abbah SA, et al. Heparin-based poly- electrolyte complex enhances the therapeutic efficacy of bone morphogenetic protein-2 for posterolateral fusion in a large animal model. *Spine (Phila Pa 1976)*. 2016;41:1199–1207. doi: 10.1097/BRS.0000000000001543.
 183. Carragee, E. J., Hurwitz, E. L., & Weiner, B. K. A critical review of recombinant human bone morphogenetic protein-2 trials in spinal surgery: emerging safety concerns and lessons learned. *The Spine Journal*. 2011; 11(6), 471–491. doi:10.1016/j.spinee.2011.04.023
 184. Poynton AR, Lane JM. Safety profile for the clinical use of bone morphogenetic proteins in the spine. *Spine (Phila Pa 1976)*. 2002;27:S40–48. doi: 10.1097/00007632-200208151-00010.
 185. Glassman SD, Carreon LY, Campbell MJ, et al. The perioperative cost of Infuse bone graft in posterolateral lumbar spine fusion. *Spine J.* 2008;8:443–448. doi: 10.1016/j.spinee.2007.03.004.
 186. Ackerman, S. J., Mafilios, M. S., & Polly, D. W. Economic Evaluation of Bone Morphogenetic Protein Versus Autogenous Iliac Crest Bone Graft in Single-Level Anterior Lumbar Fusion. *Spine*. 2002; 27(Supplement), S94–S99. doi:10.1097/00007632-200208151-00017
 187. Robbins, M. A., Haudenschild, D. R., Wegner, A. M., & Klineberg, E. O. Stem Cells in Spinal Fusion. *Global Spine Journal*. 2017;7(8): 801–810. doi:10.1177/2192568217701102
 188. Friedenstein AJ, Chailakhjan RK, Lalykina KS. The development of fibroblast colonies in monolayer cultures of guinea- pig bone marrow and spleen cells. *Cell Tissue Kinet.* 1970 Oct;3(4): 393-403. doi: 10.1111/j.1365-2184.1970.tb00347.x.
 189. Fu TS, Chang YH, Wong CB, et al. Mesenchymal stem cells expressing baculovirus-engineered BMP-2 and VEGF enhance posterolateral spine fusion in a rabbit model. *Spine J.* 2015;15(9): 2036-2044. doi: 10.1016/j.spinee.2014.11.002.
 190. Alden, T. D., Varady, P., Kallmes, D. F., Jane, J. A., & Helm, G. A. Bone Morphogenetic Protein Gene Therapy. *Spine*. 2002; 27(Supplement), S87–S93. doi:10.1097/00007632-200208151-00016
 191. Musgrave DS, Bosch P, Ghivizzani S, Robbins PD, Evans CH, Huard J. Adenovirus-mediated direct gene therapy with bone morphogenetic protein-2 produces bone. *Bone*. 1999;24:541–547. doi: 10.1016/s8756-3282(99)00086-1.
 192. Bosch, P., Musgrave, D., Ghivizzani, S., Latterman, C., Day, C. S., & Huard, J. The Efficiency of Muscle-Derived Cell-Mediated Bone Formation. *Cell Transplantation*. 2000; 9(4), 463–470. doi:10.1177/096368970000900403
 193. Riew KD, Wright NM, Cheng S, Avioli LV, Lou J. Induction of bone formation using a recombinant adenoviral vector carrying the human BMP-2 gene in a rabbit spinal fusion model. *Calcif Tissue Int.* 1998;63:357–360. doi: 10.1007/s002239900540.
 194. Bjerke, B. T., Zarrabian, M., Aleem, I. S., Fogelson, J. L., Currier, B. L., Freedman, B. A., Nassr, A. Incidence of Osteoporosis-Related Complications Following Posterior Lumbar Fusion. *Global Spine Journal*. 2017; 8(6): 563–569. doi:10.1177/2192568217743727
 195. Dwyer AF, G G Wickham. Direct current stimulation in spinal fusion. *Med J Aust.* 1974 Jan 19;1(3):73-5. doi: 10.5694/j.1326-5377.1974.tb50762.x.
 196. Bassett, C. A. L., & Becker, R. O. Generation of Electric Potentials by Bone in Response to Mechanical Stress. *Science*. 1962; 137(3535): 1063–1064. doi:10.1126/science.137.3535.1063
 197. Yasuda I. Fundamental problems in the treatment of fracture. *J Kyoto Med Soc.* 1953; 4:395–406. PMID: 340088
 198. Rubinacci A, De Ponti A, Shipley A, Samaja M, Karplus E, Jaffe LF Bicarbonate dependence of ion current in damaged bone. *Calcif Tissue Int.* 1996; 58:423–428. doi: 10.1007/BF02509442.
 199. Fredericks DC, Petersen EB, Bobst JA, Gan JC, Simon BJ, Glazer P, Nepola JV. Effects of direct current electrical stimulation on gene expression of osteopromotive factors in a posterolateral spinal fusion model. *Spine (Phila Pa 1976)*. 2007; Jan 15;32(2):174-81. doi: 10.1097/01.brs.0000251363.77027.49.
 200. Gan JC, Fredericks DC, Glazer PA. Direct current and capacitive coupling electrical stimulation upregulates osteopromotive factors for spinal fusions. *Orthop J Harvard Med School*. 2004;6:57–59.
 201. Bushinsky, D. A. Metabolic alkalosis decreases bone calcium efflux by suppressing osteoclasts and stimulating osteoblasts. *American Journal of Physiology-Renal Physiology*. 1996; 271(1): F216–F222. doi:10.1152/ajprenal.1996.271.
 202. Fitzsimmons RJ, Ryaby JT, Magee FP, Baylink DJ. IGF-II receptor number is increased in TE-85 osteosarcoma cells by combined magnetic fields. *J Bone Miner Res.* 1995; 10:812–819. doi: 10.1002/jbmr.5650100519.
 203. Zhuang H, Wang W, Seldes RM, Ta- hernia AD, Fan H, Brighton CT. Electrical stimulation induces the level of TGF- β 1 mRNA in osteoblastic cells by a mechanism involving calcium/cal- modulin pathway. *Biochem Biophys Res Commun*. 1997; 237:225–229. doi: 10.1006/bbrc.1997.7118.
 204. Lorch DG, Brighton CT, Gupta R, Corsetti

- JR, Levine SE, Gelb ID, Seldes R, Pollack SR. Biochemical pathway mediating the response of bone cells to capacitive coupling. *Clin Orthop*. 1998; 350:246–256. DOI:10.1097/00003086-199805000-00033
205. Goodwin CB, Brighton CT, Guyer RD, Johnson JR, Light KI, Yuan HA. A double-blind study of capacitively coupled electrical stimulation as an adjunct to lumbar spinal fusions. *Spine*. 1999; 24:1349–1357. doi: 10.1097/00007632-199907010-00013.
206. Kane WJ. Direct current electrical bone growth stimulation for spinal fusion. *Spine*. 1988; 24:363–365
207. Kahanovitz N. Electrical stimulation of spinal fusion: a scientific and clinical update. *Spine J*. 2002;2(2):145–50. doi: 10.1016/S1529-9430(02)00177-8.
208. Linder L. Osseointegration of metallic implants. I. Light micros-copy in the rabbit. *Acta Orthop Scand*. 1989;60(2):129–134. doi: 10.3109/17453678909149239.
209. Olivares-Navarrete R, Hyzy SL, Hutton DL, et al. Direct and indirect effects of microstructured titanium substrates on the induction of mesenchymal stem cell differentiation towards the osteoblast lineage. *Biomaterials*. 2010;31(10):2728–2735. doi: 10.1016/j.biomaterials.2009.12.029.
210. Olivares-Navarrete R, Hyzy SL, Gittens RA, et al. Rough titanium alloys regulate osteoblast production of angiogenic factors. *Spine J*. 2013;13(11):1563–1570. doi: 10.1016/j.spinee.2013.03.047.
211. Olivares-Navarrete R, Hyzy SL, Slosar PJ, Schneider JM, Schwartz Z, Boyan BD. Implant materials generate different peri-implant inflammatory factors: poly-ether-ether-ketone promotes fibrosis and microtextured titanium promotes osteogenic factors. *Spine (Phila Pa 1976)*. 2015;40(6):399–404. doi: 10.1097/BRS.0000000000000778.
212. Boyan, B. D., Batzer, R., Kieswetter, K., Liu, Y., Cochran, D. L., Szmuckler-Moncler, S., Schwartz, Z. Titanium surface roughness alters responsiveness of MG63 osteoblast-like cells to 1 α ,25-(OH) $_2$ D $_3$. *Journal of Biomedical Materials Research*. 1998; 39(1): 77–85. doi:10.1002/(sici)1097-4636(199801)39:1<77::aid-jbm10>3.0.co;2-l
213. Yang Y, Tian J, Deng L, Ong JL. Morphological behavior of osteoblast-like cells on surface-modified titanium in vitro. *Biomaterials*. 2002;23(5):1383–1389. doi: 10.1016/s0142-9612(01)00259-9.
214. De Leonardis D, Garg AK, Pecora GE. Osseointegration of rough acid-etched titanium implants: 5-year follow-up of 100 minimatic implants. *Int J Oral Maxillofac Implants*. 1999;14(3):384–391. PMID: 10379112
215. Hwang HH, Zhu W, Victorine G, Lawrence N, Chen S. 3D- printing of functional biomedical microdevices via light- and extrusion-based approaches. *Small Methods*. 2017;2(2). doi: 10.1002/smt.201700277.
216. Gittens RA, Olivares-Navarrete R, McLachlan T, et al. Differential responses of osteoblast lineage cells to nanotopographically-modified, microroughened titanium–aluminum–vanadium alloy surfaces. *Biomaterials*. 2012;33:8986– 8994. doi: 10.1016/j.biomaterials.2012.08.059.
217. Vyatskikh A, Delalande S, Kudo A, Zhang X, Portela CM, Greer JR. Additive manufacturing of 3D nano-architected metals. *Nat Commun*. 2018;9(1):593. DOI:10.1038/s41467-018-03071-9
218. Casper, D. S., Zmistowski, B., Hollern, D. A., Hilibrand, A. S., Vaccaro, A. R., Schroeder, G. D., & Kepler, C. K. The Effect of Postoperative Spinal Infections on Patient Mortality. *Spine*. 2018; 43(3), 223–227. doi:10.1097/brs.0000000000002277
219. Qin H, Cao H, Zhao Y, et al. In vitro and in vivo anti-biofilm effects of silver nanoparticles immobilized on titanium. *Biomaterials*. 2014;35(33):9114–9125. doi: 10.1016/j.biomaterials.2014.07.040.
220. Colon, G., Ward, B. C., & Webster, T. J. Increased osteoblast and decreased Staphylococcus epidermidis functions on nanophase ZnO and TiO $_2$. *Journal of Biomedical Materials Research Part A*. 2006; 78A(3), 595–604. doi:10.1002/jbm.a.30789
221. Puckett SD, Taylor E, Raimondo T, Webster TJ. The relationship between the nanostructure of titanium surfaces and bacterial attachment. *Biomaterials*. 2010;31(4):706–713. doi: 10.1016/j.biomaterials.2009.09.081.
222. Hazer DB, Sakar M, Dere Y, Altinkanat G, Ziyal MI, Hazer B. Antimicrobial effect of polymer-based silver nanoparticle coated pedicle screws: experimental research on biofilm inhibition in rabbits. *Spine (Phila Pa 1976)*. 2016;41(6):E323–E329. doi: 10.1097/BRS.0000000000001223.
223. Li W, Xu D, Hu Y, Cai K, Lin Y. Surface modification of titanium substrates with silver nanoparticles embedded sulfhydrylated chitosan/gelatin polyelectrolyte multilayer films for antibacterial application. *J Mater Sci Mater Med*. 2014;25(6):1435–1448. doi: 10.1007/s10856-014-5190-8.
224. Slosar PJ. Spine implant surface technology state of the art: separating fact from fiction. *Spine (Phila Pa 1976)*. 2018;43:S10–S11. DOI: 10.1097/BRS.0000000000002549
225. Torstrick FB, Evans NT, Stevens HY, Gall K, Guldberg RE. Do surface porosity and pore size influence mechanical properties and cellular response to PEEK? *Clin Orthop Relat Res*. 2016;474(11):2373–

2383. doi: 10.1007/s11999-016-4833-0.
226. Katsuura, Y., Wright-Chisem, J., Wright-Chisem, A., Virk, S., & McAnany, S. (2020). The Importance of Surface Technology in Spinal Fusion. *HSS Journal*. 2020;16(2):113–116. doi:10.1007/s11420-020-09752-w
227. Polini A, Pisignano D, Parodi M, Quarto R, Scaglione S. Osteoinduction of human mesenchymal stem cells by bioactive composite scaffolds without supplemental osteogenic growth factors. *PLoS One*. 2011;6(10):e26211. doi: 10.1371/journal.pone.0026211.
228. Kandziora, F., Pflugmacher, R., Scholz, M., Eindorf, T., Schnake, K. J., & Haas, N. P. Bioabsorbable Interbody Cages in a Sheep Cervical Spine Fusion Model. *Spine*. 2004; 29(17):1845–1855. doi: 10.1097/01.brs.0000137060.79732.78.
229. Cao, L., Duan, Li, Yuan, Zhao, M.-D., Wu, Dong. Biomechanical stability of a bioabsorbable self-retaining polylactic acid/nano-sized β -tricalcium phosphate cervical spine interbody fusion device in single-level anterior cervical discectomy and fusion sheep models. *International Journal of Nanomedicine*. 2012;7:5875-80. doi:10.2147/ijn.s38288
230. Steinberger, J., York, P., Virk, S., & Kim, H. J. Advances in Spinal Fusion Strategies in Adult Deformity Surgery. *HSS Journal*. 2020. doi:10.1007/s11420-020-09751-x.
231. He B, Yuan X, Zhou A, Zhang H, Jiang D. Designer functionalised self-assembling peptide nanofibre scaffolds for cartilage tissue engineering. *Expert Rev Mol Med*. 2014;16:e12. doi: 10.1017/erm.2014.13.
232. Scheer, J. K., Osorio, J. A., Smith, J. S., Schwab, F., Lafage, V., Hart, R. A., Ames, C. P. Development of Validated Computer-based Preoperative Predictive Model for Proximal Junction Failure (PJF) or Clinically Significant PJK With 86% Accuracy Based on 510 ASD Patients With 2-year Follow-up. *SPINE*. 2016;41(22): E1328 E1335. doi:10.1097/brs.0000000000001598
233. Ames, C. P., Scheer, J. K., Lafage, V., Smith, J. S., Bess, S., Berven, S. H., Daubs, M. D. Adult Spinal Deformity: Epidemiology, Health Impact, Evaluation, and Management. *Spine Deformity*. 2016; 4(4):310–322. doi:10.1016/j.jspd.2015.12.009