

Bacteriological Profile of Neonatal Sepsis and Antibiogram of the Isolates

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Abstract

Introduction: Neonatal sepsis is a serious problem in developing countries like Nepal. The main objectives of this study were to determine the bacteriological profile of neonatal sepsis, to determine the antimicrobial susceptibility patterns of the causative agents and to evaluate the association between the neonatal sepsis and the different characteristics of the neonates. **Material and Methods:** A hospital based cross-sectional study was conducted among a total of 450 neonates suspected of suffering from sepsis. Blood culture was performed using standard microbiological techniques. The colonies grown were identified on the basis of colony morphology, Gram's stain and biochemical tests. The antimicrobial susceptibility testing was performed by Kirby Bauer disc diffusion method. **Results:** Out of total 450 blood samples, 92 (20.4%) were culture positive. Of which, 16 (17.4%) samples contained Gram negative bacilli and 76 (82.6%) samples contained Gram positive cocci. The most common bacterial pathogens isolated were *Staphylococcus epidermidis* (67.4%) followed by *Escherichia coli* (13%). All Gram positive cocci were susceptible to vancomycin, while all Gram negative bacilli were sensitive to amikacin. There was statistically significant relationship between neonatal sepsis and gestation age of neonates. **Conclusion:** Neonatal sepsis is still present as a serious problem in Nepal. *Staphylococcus epidermidis* was the most common cause of the neonatal sepsis. Prematurely delivered neonates are more prone to suffer from neonatal sepsis. Vancomycin and amikacin can be used as the drugs of choice for preliminary treatment of neonatal sepsis in our settings.

Key words: Neonatal sepsis, *Staphylococcus epidermidis*, *Escherichia coli*, Nepal

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Introduction

Neonatal sepsis is a clinical condition characterized by systemic signs and symptoms due to bacteremia in the first month of the life¹. It is a global problem and despite the development of highly effective antibiotics and implementation of the hygiene practices in the healthcare settings, neonatal sepsis has established itself as a

major cause of morbidity and mortality with high level of impact in low resource countries². The incidence of neonatal sepsis may vary not only from developed countries to developing countries but also from hospital to hospital even in the same country². Twenty percent of all neonates get neonatal sepsis and is the cause of 30-50% of total neonatal deaths³.

According to World Health Organization every year an estimated 1.6 million neonatal deaths occur globally with 40% of all neonatal deaths occurring in developing countries⁴. The risk factors those may be associated with neonatal sepsis are premature rupture of membrane, prolonged rupture, prematurity, urinary tract infection, poor maternal nutrition, low birth weight, birth asphyxia and congenital anomalies⁵. Neonatal sepsis may be divided into two types: early onset and late onset³. The infection acquired within 72 hrs of age is known as early onset neonatal sepsis and the common bacteria associated with it are group B *Streptococcus*, *Escherichia coli*, coagulase negative *Staphylococcus* spp., *Hemophilus influenzae* and *Listeria monocytogenes*³. Similarly, the infection acquired after 72 hrs of age is known as late onset neonatal sepsis and the most common causative agents are coagulase negative *Staphylococcus* spp., *S. aureus*, *Klebsiella pneumoniae*, *E. coli*, *Enterobacter* spp., *Pseudomonas aeruginosa* and *Acinetobacter* spp.³.

Neonatal sepsis can be life threatening if proper treatment is not given in time⁶. Blood culture for the isolation of the causative agent is gold standard for identification of the cases of neonatal sepsis and the antibiotic susceptibility pattern of the bacteria isolated is necessary for giving proper treatment². In Nepal, different studies have reported the high rates of neonatal sepsis with the bacteria showing different rates of resistance to commonly used antibiotics^{3,7}. The microbiological pattern and antimicrobial susceptibility patterns of the causative agents of neonatal sepsis may vary from hospital to hospital and their knowledge may be helpful in timely proper management of neonatal sepsis. So, in this study we determined the rate of neonatal sepsis, bacteriological profile of neonatal sepsis and antimicrobial susceptibility patterns of the causative agents in a tertiary care hospital in Kathmandu, Nepal. Further, we also determined the association between the neonatal sepsis and the different characteristics of the neonates.

Material and Methods

A hospital based cross-sectional study was conducted among a total of 450 neonates suspected

of suffering from sepsis (children with fever, breathing problem, low blood sugar, reduced sucking, low or high heart rate) at Paropakar Maternity and Women's hospital, Thapathali, Kathmandu, Nepal, a tertiary care hospital from April 2013 to September 2013. The hospital has well-equipped neonatal intensive care unit.

One ml of venous blood was collected using standard procedures and was inoculated into 9 ml of brain heart infusion broth (HiMedia, India). The blood culture bottles were immediately sent to the microbiology laboratory of the Paropakar Maternity and Women's hospital. All the blood culture bottles were incubated at 37°C for 24 hrs and subcultured on MacConkey agar, blood agar and chocolate agar (HiMedia, India) daily for 7 days. The inoculated MacConkey agar plates were incubated aerobically, where as blood agar and chocolate agar plates were incubated in CO₂ enriched humid atmosphere using candle jar, at 37°C for 24-48 hours. Blood culture bottles showing no growth on subculture done after incubation of 7 days were reported as negative⁸. The colonies grown were identified on the basis of colony morphology, Gram's stain and biochemical tests⁹. The necessary patient's informations were obtained from the neonatal ward, which were entered in excel.

The antimicrobial susceptibility testing was performed by Kirby Bauer disc diffusion method following standard guidelines and interpretive criteria of the Clinical and Laboratory Standards Institute¹⁰. For quality control of biochemical tests, purity plate was used. Similarly, for quality control of antimicrobial susceptibility testing, *Escherichia coli* ATCC 25922 and *Staphylococcus aureus* ATCC 25923 were used.

Data were analyzed using statistical package for social sciences version 16.0. Chi-square test was used and p-value<0.05 was considered as statistically significant.

Results

Out of total 450 blood samples, 92 (20.4%) were culture positive. Of all 92 bacterial isolates, 16 (17.4%) were Gram negative bacilli and 76 (82.6%) were Gram positive cocci.

Different bacterial pathogens isolated from the cases of neonatal sepsis: The bacterial pathogens isolated were *Staphylococcus epidermidis* (67.4%) followed by *Escherichia coli* (13%), *Staphylococcus aureus* (9.8%), *Klebsiella pneumoniae* (4.3%), *Staphylococcus saprophyticus* (3.3%) and *Enterococcus faecalis* (2.2%).

Association of sex of neonates with growth of bacteria in blood culture: Out of 450 neonates, 237 (52.7%) were females and 213 (47.3%) were males. Among males, 22.4% showed blood culture positive while 21.8% female neonates were blood culture positive. There was no statistically significant relationship between growth of organisms and sex of the neonates ($p>0.05$).

Association of mode of delivery of neonates with growth of bacteria in blood culture: Of the 450 neonates, normal delivery was found to be occurring in highest numbers 273(60.7%) followed by caesarean delivery 152(33.8%) and instrumental delivery 25(5.6%). It was found that neonates having normal delivery had maximum culture positive cases (n=55) followed by caesarean delivery (n=30) and instrumental delivery (n=7). However, there was no statistically significant relationship between growth of organisms and mode of delivery of neonates ($p>0.05$).

Association of neonatal weight with growth of bacteria in blood culture: Out of 450 neonates, 178(39.6%) neonates were born with very low birth weight (<1500g), 176(39.1%) with low birth weight (1500-2500g) and remaining 96(21.3%) with good birth weight (>2500g). Growth of organisms in blood was seen maximum in neonates with very low birth weight (24.2%) followed by neonates with low birth weight (19.3%) and neonates with good birth weight (15.6%). But there was no statistically significant relationship between neonatal weight and growth of organisms in blood culture ($p>0.05$).

Association of gestation age of neonates with growth of bacteria in blood culture: Out of 450 neonates, 181(40.2%) were pre-term babies (<37 weeks), 191(42.4%) were term babies (37-42 weeks) and 78(17.3%) were post-term babies (>42 weeks). The rate of growth of organisms was highest among pre-term babies 36.5% (66/181) followed by term babies 11% (21/191) and post-term babies 6.4% (5/78). There was statistically significant relationship between growth of organisms and gestation age of neonates ($p<0.05$).

Antibiotic susceptibility patterns of the Gram positive cocci: Among Gram positive cocci isolated, highest rate of susceptibility was seen toward vancomycin (100%) followed by amikacin (80.3%) (Table 1).

Antibiotic susceptibility patterns of Gram negative bacilli: Among Gram negative bacilli, the highest rate of susceptibility was seen toward amikacin (100%) followed by cefotaxime (75%) and ciprofloxacin (75%) (Table 2).

Table 1: Antimicrobial susceptibility patterns of the Gram positive cocci

| Antibiotics | Susceptibility (%) |
|---------------|--------------------|
| Erythromycin | 36 (47.4%) |
| Gentamicin | 39 (51.3%) |
| Vancomycin | 76 (100%) |
| Amikacin | 61 (80.3%) |
| Cefotaxime | 48 (63.2%) |
| Ciprofloxacin | 51 (67.1%) |
| Cloxacillin | 44 (57.9%) |

Table 2: Antimicrobial susceptibility patterns of the Gram negative bacilli

| Antibiotics | Susceptibility (%) |
|---------------|--------------------|
| Ampicillin | 3 (18.8%) |
| Gentamicin | 10 (62.5%) |
| Tobramycin | 7 (43.8%) |
| Amikacin | 16 (100%) |
| Cefotaxime | 12 (75%) |
| Ciprofloxacin | 12 (75%) |
| Ceftazidime | 7 (43.8%) |

Discussion

In the developing countries like Nepal, neonatal sepsis is a serious problem³. In our study, the rate of neonatal sepsis was 20.4%, which was similar to the finding by Samaga and Sumangala (21.9%)¹¹. However, higher rates were reported by Muley et al. (26.6%)¹², Jain et al. (28.3%)⁶, Shrestha et al. (30.8%)³, Bhatt et al. (55.6%)¹³, Malla et al. (57.22%)⁷ and Premalatha et al. (82.35%)¹⁴, while lower rates were noted by Ansari et al. (12.6%)¹⁵ and Gyawali and Sanjana (15.13%)¹⁶. Use of antibiotics just after birth, effective control of nosocomial infections and infection by anaerobes may be reason for the difference in the results reported by different authors³. The patients might have got infections either from the mothers during birth or from hospital environment after birth.

As in our study, in a study by Ansari et al., among all bacterial isolates 63.8% were Gram positive isolates and 36.2% were Gram negative isolates with the commonest bacteria being coagulase negative *Staphylococcus* spp. followed by *S. aureus*¹⁵. Similarly, in another study by Shrestha et al. the most common organisms isolated from the cases of neonatal sepsis were *S. aureus* followed by *Klebsiella pneumoniae*³. Further, Gyawali and Sanjana noted the incidence of Gram positive and Gram negative organisms to be 44.1% and 55.9% respectively with *Staphylococcus aureus* being the predominant isolate followed by *Klebsiella* spp.¹⁶. However, Muley et al. showed the *Klebsiella pneumoniae* to be the most

predominant pathogen followed by *Staphylococcus aureus*¹². Accordingly, Kumaravel and Rameshbabu found the Gram negative bacteria to be predominant (88%) with *Klebsiella* spp. being commonest followed by *E. coli*¹⁷. The causative agents of neonatal sepsis have changed over time and may vary from place to place¹⁵.

We found the statistically significant association between the gestational age and rate of neonatal sepsis with prematurely delivered neonates being at higher risk of sepsis. Similarly, Premalatha et al. found the low birth weight and prematurity to be the risk factors for neonatal sepsis¹⁴. But in our study, though highest rate of neonatal sepsis was found in the neonates with low birth weight, statistically there was no significant correlation. This may be due to the small sample size taken in our study. Premature babies have low immunity and are more prone to infection⁷. The risk factors associated with neonatal sepsis are premature rupture of membranes, prolonged rupture, prematurity, urinary tract infections, poor maternal nutrition, low birth weight, birth asphyxia and congenital anomalies⁵.

As we have reported, Kumaravel and Rameshbabu showed the highest rates of susceptibility of Gram negative and Gram positive bacteria toward amikacin and vancomycin respectively¹⁷. In addition, as in our study Gyawali and Sanjana found the third generation cephalosporins and aminoglycosides to be more satisfactory for Gram negative bacteria in comparison to Gram positive bacteria¹⁶. Further, Muley et al. reported the maximum susceptibility of both Gram negative and Gram positive bacteria to ciprofloxacin and amikacin¹².

And suggested to use these antibiotics in empirical therapy of neonatal sepsis¹². The difference in patterns of antibiotic usage in different hospitals is the main reason for the difference in antibiotic susceptibility reported by different authors.

We did not try to find out the source of infection in neonates, which is a major limitation of our study.

Conclusion

Staphylococcus epidermidis followed by *Escherichia coli* were the most common causes of the neonatal sepsis. Prematurely delivered neonates are more prone to suffer from neonatal sepsis. Vancomycin and amikacin can be used as the drugs of choice for preliminary treatment of neonatal sepsis in our settings. Further, cefotaxime and ciprofloxacin may be good options for treatment of neonatal sepsis caused by Gram negative bacteria.

Recommendations

Prematurely delivered neonates should be given more care, as they are more prone to suffering from neonatal sepsis. Effective infection control program including the strict hand hygiene policy should be implemented in the neonatal ward to prevent the neonatal sepsis. Further, the vaginal swab of the pregnant woman should be cultured for the detection of the colonization by the possible pathogens before delivery, so that the necessary steps can be taken to prevent the infection in neonate during birth.

References

- Saranghi KK, Pattnaik D, Mishra SN, Nayak MK, Jena J. Bacteriological profile and antibiogram of blood culture isolates done by automated culture and sensitivity method in a neonatal intensive care unit in a tertiary care hospital in Odisha, India. *Int J Advances Med* 2015;2(4):387-92. DOI: <http://dx.doi.org/10.18203/2349-3933.ijam20151015>
- Pius S, Bello M, Galadima GB, Ibrahim HA, Yerima ST, Ambe JP. Neonatal septicaemia, bacterial isolates and antibiogram sensitivity in Maiduguri North-Eastern Nigeria. *Niger Postgrad Med J* 2016;23:146-51. DOI: 10.4103/1117-1936.190340
- Shrestha RK, Rai SK, Khanal LK, Mandal PK. Bacteriological study of neonatal sepsis and antibiotic susceptibility pattern of isolates in Kathmandu, Nepal. *Nepal Med Coll J* 2013;15(1):71-3.
- Sawhney N, Shynu P, Singh VA. Bacteriological Profile and Antibiotic Susceptibility Pattern of Neonatal Septicaemia in a Tertiary Care Hospital. *Int J Curr Microbiol App Sci* 2015;4(10):977-84.
- Prabhu K, Bhat S, Rao S. Bacteriologic profile and antibiogram of blood culture isolates in a pediatric care unit. *J Lab Physicians* 2010;2:85-8. DOI: 10.4103/0974-2727.72156.
- Jain NK, Jain VM, Maheshwari S. Clinical profile of neonatal sepsis. *Kathmandu Univ Med J* 2003;1(2):117-20.
- Malla KK, Malla T, Rao KS. Bacteriological Profile of Sepsis Outbreak in the NICU of a Tertiary Care Hospital in Western Nepal. *J Nepal Paediatr Soc* 2013;33(1):8-14. DOI: <http://dx.doi.org/10.3126/jnps.v33i1.7016>
- Cheesbrough M. District laboratory practice in tropical countries, part II. 2nd edition. New York: Cambridge University Press; 2006.
- Holt JG, Krieg NR, Sneath PHA, Staley JT, Williams ST. Bergey's manual of determinative bacteriology. Williamsons and Wilkins, Baltimore. 1994.

10. Clinical and Laboratory Standards Institute. CLSI document M100-S23. Performance standards for antimicrobial susceptibility testing; Twenty third informational supplement edition. Wayne: CLSI; 2013.
11. Samaga MP, Sumangala B. Bacteriological profile of neonatal Septicaemia in MIMS, Mandya, India. *Int J Curr Microbiol App Sci* 2016;5(3):495-501.
12. Muley VA, Ghadage DP, Bhore AV. Bacteriological profile of neonatal septicemia in a tertiary care hospital from Western India. *J Glob Infect Dis* 2015;7(2):75-7. doi: 10.4103/0974-777X.154444.
13. Bhatt SK, Patel DA, Gupta P, Patel K, Joshi G. Bacteriological profile and antibiogram of neonatal septicemia. *National J Com Med* 2012;3(2):238-41.
14. Premalatha DE, Koppad M, Halesh LH, Siddesh KC, Prakash N. The Bacterial Profile and Antibiogram of Neonatal Septicaemia in a Tertiary Care Hospital. *Int J Rec Trends Sci Tech* 2014;10(3):451-5.
15. Ansari S, Nepal HP, Gautam R, Shrestha S, Neopane P, Chapagain ML. Neonatal Septicemia in Nepal: Early-Onset versus Late-Onset. *Int J Pediatr* 2015;2015:379806. DOI:<http://dx.doi.org/10.1155/2015/379806>.
16. Gyawali N, Sanjana RK. Bacteriological profile and antibiogram of neonatal septicemia. *Indian J Pediatr* 2013;80(5):371-4. DOI:10.1007/s12098-012-0911-9.
17. Kumaravel KS, Rameshbabu B. A study of the bacteriological profile and antibiotic sensitivity in neonatal septicemia. *Int J Cont Med Res* 2016;3(6):1830-1.