

Evaluation of an enzyme immunoassay and TPPA for detection of antibodies against *Treponema pallidum*

Maharian A^a, Karki S^b, Shrestha S^a and Raikarnikar M^a

Abstract

Introduction An Enzyme Linked Immunosorbent Assay (ELISA) technique is used to screen the *Treponema pallidum* antibodies in donated blood by many blood banks of all around the world, where as their sensitivity and specificity has been reported to be variable in different population. ELISA techniques have been reported to give less equivocal results and can easily be automatized, making them suitable to be used in Blood Banks, where large number of blood samples need to be screened.

Objectives To assess the suitability of an ELISA (Enzygnost Syphilis) test to screen the specific *Treponema pallidum* antibodies in the units of blood collected from Nepalese Blood donors.

Methods This research was conducted in Central Blood Transfusion Service, Nepal Red Cross Society (Exhibition Road). A total of 760 blood samples were randomly collected from blood donors and tested for the presence of specific *Treponema pallidum* antibodies using ELISA (Enzygnost Syphilis) in an automated ELISA Processor (BEP-III) and TPPA (Serodia TP-PA), strictly following the protocol described in the kit inserts.

Results The study evaluated an Enzyme Immunoassay (Enzygnost Syphilis) test for qualitative detection of specific *Treponema pallidum* antibodies in 760 sera of blood donors. There was a strong association of test results between the Enzygnost Syphilis and Serodia TP-PA (P-Value <0.000, Fisher's Exact Test). For sera of 9 blood donors the Enzygnost Syphilis was reactive (Sensitivity, 100% in relation to TP-PA) and for sera of 751 blood donors the Enzygnost Syphilis was non-reactive (Specificity, 100% in relation to TP-PA), the overall agreement of the test results was 100 percent (760 of 760 sera).

Conclusion The result suggests that the ELISA test evaluated in this study could be used to screen specific *Treponema pallidum* antibodies in blood units collected from blood donors.

Keywords Screening *Treponema pallidum* antibodies, *Treponema pallidum*, ELISA, TP-PA

Introduction

Treponema pallidum is the dominant pathogen among the spirochetes that causes the venereal syphilis. The incubation period varies from 3-90 days, with a mean of three weeks. The disease encompasses through the stages of primary syphilis, secondary syphilis, latent syphilis and late syphilis¹.

Syphilis can be transmitted by only a few routes: sexual contact², direct introduction into vascular system by shared needles or transfusions³, direct cutaneous contact with infectious lesions, or

transplacental transfer of spirochetes. It has been estimated epidemiologically that as many as 50 percent of sexual contacts of infectious persons escape infection¹.

Transfusion transmitted syphilis is not a major hazard of modern blood transfusion therapy. *Treponema pallidum*, the infectious agent causing syphilis survives at the most for 5 days in blood stored at 4°C⁴. The transmission of syphilis in itself is not a big problem because cure is available for it. However, the presence of a sexually transmitted disease

Corresponding Author: Anil Maharian, MPH. **E-mail:** anilaabhas@hotmail.com. ^aCentral blood transfusion service, Nepal Red Cross Society, Exhibition Road, Kathmandu, Nepal. ^bCentral Department of Microbiology, TU Kirtipur.

Table 1: Characteristics of serologic tests for syphilis.

TEST	TYPE	% POSITIVE AT INFECTIOUS STAGES		
		Primary	Secondary	Late
VDRL	Nontreponemal	70	99	1
RPR	Nontreponemal	80	99	0
FTA-ABS	Treponemal	85	100	98
TPHA	Treponemal	65	100	95
TPI	Treponemal	50	97	95

Source: Tramont EC. et al. 1990

indicates toward donor's indulgence in "high risk" behavior and consequent higher risk of exposure to infections like HIV and hepatitis which have no cure.

The serologic tests for syphilis can be divided into two groups: non-treponemal tests and treponemal tests. The non-treponemal tests take advantage of antibodies to a tissue lipid, called cardiolipin that are produced as a byproduct of treponemal infection. The procedures in current use are flocculation tests. They use a form of cardiolipin that is complexed with a cholesterol and lecithin. The most commonly used procedures are Venereal Disease Research Laboratory (VDRL) and the rapid plasma reagin (RPR) tests. The RPR card tests have achieved popularity because the VDRL test is technically demanding and unforgiving. Other tests that have been used are the reagin screen tests (RST), the unheated serum reagin (USR) test and the toluidine red unheated serum test (TRUST). The non-treponemal tests have a sensitivity of 70-99 percent, depending on the stage of disease¹.

The treponemal tests use specific treponemal antigens into the system. The traditional gold standard was the Treponema pallidum immobilization (TPI) test. This expensive and cumbersome test has been replaced by FTA-ABS test. As this test also requires fluorescent microscope, other easier approaches have been developed. These include Enzyme immunoassays and Particle agglutination test detecting specific treponemal antibodies^{5,6,7,8,9,10,11}.

Materials and Methods

A total of 760 blood samples were collected in July to August 2004 from blood donors. All the samples were tested individually for screening of syphilis by using two commercially available screening kits based on different principles viz. Enzygnost Syphilis (Behring, Marburg, Germany) and Serodia TP-PA (Fujiirebio, Japan).

The Enzygnost Syphilis is a competitive enzyme immunoassay for the qualitative detection of specific antibodies to Treponema pallidum in human serum or human plasma.

Serodia-TP-PA is a qualitative gelatin particle agglutination assay intended to be used for the detection of Treponema pallidum antibodies in human serum or plasma as an aid in the diagnosis of syphilis.

The principle of Enzygnost Syphilis test is a competitive one stage enzyme immunoassay for the in vitro determination of antibodies to Treponema pallidum. Treponema pallidum specific antibodies (IgG and/or IgM) contained in the sample and the POD labeled antibodies (anti-T. pallidum/POD conjugate) compete for binding to the Treponema pallidum antigens coated into the wells of the microtitration plate. Unbound serum antibodies and conjugate antibodies are washed out and the enzyme activity of the bound conjugate is then determined. The enzyme component of the conjugate reacts with the working chromogen solution (TMB plus hydrogen peroxide), thereby producing a blue colour. The reaction is terminated by the addition of stopping solution POD, resulting in a colour change to yellow. The intensity of the resultant yellow colour is inversely proportional to the concentration of the Treponema pallidum antibodies in the sample. This test was performed according to the recommendation of the manufacturer using the Behring ELISA processor (BEP III) that included incubation, washing, dispensing and reading the result except specimen loading.

The principle of Serodia TP-PA test is based on the agglutination of colored gelatin particle carriers sensitized with T. pallidum (Nichols Strain) antigen. Serum or plasma samples are serially diluted in sample diluent in microplate wells. Sensitized Gelatin Particles are added to respective wells and the contents of the plate mixed by hand or on a tray mixer. The mixture is incubated stationary for 2 hours at room

Table 2: Qualitative results obtained by Enzvgnost Svophilis (ELISA) and Serodia TP-PA.

		Serodia TP-PA		
		Reactive	Non-reactive	Total
Enzvgnost	Reactive	9	0	9
Svophilis	Non-reactive	0	751	751
Total		9	751	760

temperature. Serum or plasma containing specific antibodies reacts with the antigen-sensitized colored gelatin particles to form a smooth mat of agglutinated particles in the microtitration tray. A compact button formed by the settling of the non-agglutinated particles characterizes negative reactions. The test is designed to be used exclusively with microtitration techniques. The agglutination patterns and interpretation of the test are clear cut and easy to read visually or with the aid of a tray viewer.

The data of the study was analyzed by Fisher's Exact Test using the statistical software "WinPeri" version 3.8.

Results

Among the 760 samples tested, 9(1.2%) were reactive both by Serodia TPPA and Enzvgnost Svophilis and 751(98.8%) were non-reactive by both the techniques (table 1 and 2). There was a good association of the test result between these two tests during screening of the specific treponemal antibodies in blood donors (P-Value<0.000, Fisher's Exact Test). The samples were also used for evaluation of the sensitivity and specificity of the Enzvgnost svophilis (ELISA technique) in relation to the result of TP-PA regarding the TP-PA as "Gold standard." The sensitivity and specificity of the Enzvgnost Svophilis (ELISA) was 100 percent compared with the result of Serodia TP-PA. Sensitivity (100%), Specificity (100%), Negative Predictive Value (100%), Positive predictive value (100 %) compared to Serodia TP-PA.

Discussion

The possibility of using an ELISA technique as an alternative to other formats of treponemal tests has been evaluated in various studies. But there was no reliable data that support the use of an ELISA technique, as a screening test to detect the specific treponemal antibodies among Nepalese blood donor population. So, in this study, only the sera of blood donors, undergoing stringent donor screening and selection criteria was used. The Serodia TP-PA was used as Bench-mark to compare the ELISA (Enzvgnost Svophilis) and to evaluate the relative

sensitivity and specificity because of a number of reasons like, the Serodia TP-PA has been described to be produced using the same antigens, the high performance liquid chromatography purified sonicate of *Treponema pallidum*, that are used in a reliable and established confirmatory test for Svophilis, the MHA-TP(Serodia TP-PA, Kit insert). The Serodia TP-PA has been shown to be more sensitive and specific than many ELISA techniques in a number of studies particularly during the secondary and latent stage of the disease¹². The Serodia TP-PA has also been used by Center for Disease Control and prevention (CDC) as a confirmatory test to diagnose Svophilis¹³. In this study, the ELISA (Enzvgnost Svophilis) showed a sensitivity and specificity of 100 percent, which is slightly higher than in the study published by Virivatavikul R. et al¹⁴, (i.e. sensitivity of 99.1 and specificity of 98.91 percent when the equivocal value was considered negative and sensitivity of 100 percent and specificity of 97.89 when the equivocal value was considered positive) which may be due to testing of higher number of samples in that study (i.e. 2882), some of which were reported to be previously known positive samples by VDRL, TPHA or FTA-ABS tests, probably representing the more diverse population and stage of disease. Similarly, slightly less sensitivity of Enzvgnost Svophilis was reported by Maidment C. et al. (i.e. 99.5%) compared with the result of TPHA¹⁶; this was probably due to the use of sera which were recognized as causing problems with enzyme immunoassays. But the test specificity observed in our study was totally in concordance with the study conducted by Microbiological Diagnostics Assessment Service (MiDAS)¹⁵ and as claimed by the Dade Behring Company (kit insert) while testing the samples of Blood donors. Thus, on the basis of present study and similar studies^{11,15,16}, ELISA (Enzvgnost Svophilis) could be used as a screening test to detect specific treponemal antibodies in sera from healthy blood donors, where large number of donated blood need to be tested.

Conclusion

The current study has demonstrated that the ELISA (Enzvgnost svophilis) is highly sensitive and specific

screening method to screen specific treponemal antibodies in donated units of blood, while comparing it with the test results by Serodia TP-PA. Since, the ELISA tests can easily be automated, human errors would be minimized and test reports can be timely released, improving the quality of service where large numbers of blood units have to be screened. Moreover, ELISA technique is relatively cheaper than TP-PA for screening specific treponemal antibodies.

Acknowledgement

We are grateful to National AIDS and STD control center, Teku, for providing sufficient Serodia TP-PA kit to conduct this study.

References

1. Winn W, Stephen A, Janda W, Koneman E, Procop G, Schreckenberger P, et al. Koneman's Color Atlas and Textbook of Diagnostic Microbiology. *Lippincott's Williams and Wilkins*. 6th ed.. 2006: 1126-8.
2. Rolfs RT, Goldberg M, Sharrar RG. Risk factors for syphilis: Cocaine use and prostitution. *American journal of public health* 1990; 80: 853-8
3. Chambers RW, Foley HT, Schmidt PJ. Transmission of syphilis by fresh blood components. *Transfusion* 1969; 9: 32-4.
4. Van der Sluis JJ, Ten kate FJ, Vujeviski VD, Kothé FC, Albers GM, van Eijk RV. Transfusion syphilis, survival of *Treponema pallidum* in donor blood II. Dose dependence of experimentally determined survival times. *Vox Sang* 1985; 49: 390-9.
5. Ebel A, Bachelart L, Aionso M. Evaluation of a New Competitive Immunoassay (BioElisa Syphilis) for Screening for *Treponema pallidum* Antibodies at Various Stages of Syphilis. *J. Clin. Microbiol* 1998; 36: 358-61.
6. Fears MB, Pope V. Syphilis fast latex agglutination test, a rapid confirmatory test. *Clin Diagn Lab Immunology* 2001; 8: 841-2.
7. Hagedorn HJ, Kraminer-Hagedorn A, De Bosschere K, Hulstaert F, Pottel H, Zrein M. Evaluation of INNO-LIA Syphilis assay as a confirmatory test for syphilis. *J. Clin. Microbiol* 2002; 40: 973-8.
8. Pope V, Fears MB, Morrill WE, Castro A, Kikkert SE. Comparison of the Serodia *Treponema pallidum* particle agglutination, Cantia Syphilis-G, and SpiroTek Reagin II tests with standard test techniques for diagnosis of syphilis. *J Clin Microbiol* 2000; 38: 2543-5.
9. Schmidt BL, Edilalipour M, Luger A. Comparative evaluation of nine different enzyme-linked immunosorbent assays for determination of antibodies against *Treponema pallidum* in patients with primary syphilis. *J. Clin. Microbiol.* 2000; 38:1279-82
10. Zarakolu P, Buchanan I, Tam M, Smith K, Hook III EW. Preliminary evaluation of an immunochromatographic strip test for specific *Treponema pallidum* antibodies. *J Clinical Microbiology* 2002; 40: 3064-5.
11. Zrein M, Maure I, Boursier F, Soufflet L. Recombinant antigen-based enzyme immunoassay for screening of *Treponema pallidum* antibodies in blood bank routine. *J. Clin. Microbiol* 1995; 33: 525-7.
12. Schmidt BL. Evaluation of a new particle gel immunoassay for determination of antibodies against *Treponema pallidum*. *J Clin Microbiol* 2004; 42: 2833-5.
13. Pope V, Fears MB. Serodia *Treponema pallidum* passive particle agglutination (TP-PA) test. In: Larsen SA, Pope V, Johnson RE, Kennedy E Jr., editors. A manual of tests for syphilis, supplement. Washington, DC: *American Public Health Association* 2000: 363-78.
14. Virivatavikul R, Laodee N, Potprasat S, Pivonhiranong S. Comparative evaluation of three different enzyme immunoassays for syphilis. *J Med Assoc Thai* 2006; 89: 773-8.
15. Colle M, Perry K. Ten Syphilis EIAs. Microbiological diagnostics assessment service. 2004. For more information visit www.midas-evaluation.org.uk
16. Maidment C, Woods A, Chan R. An evaluation of the Behring Diagnostics Enzygnost® Syphilis enzyme immunoassay. *Pathology* 1998; 32: 177-8.