Intensity Revival of Weak Symmetric Infrared Band is Possible in Ubiquinone Molecules through the Asymmetric Site-specific Isotope Labeling

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Abstract: Modifications of Infrared (IR) spectral bands of ubiquinone molecule (UQ_p) upon site-specific ^{13}C labeling at the C_s or C_6 position are studied in CCl_4 using Gaussian 03. Polarizable continuum model (PCM) has been used to optimize the UQ1 molecule in solvent. The unlabeled neutral ubiquinone molecule consists of three intense IR bands in the frequency region between 1700 cm^{-1} to 1550 cm^{-1} . The symmetric fourth band in this spectral region does not appear in the spectrum because of very weak intensity. However, site-specific ^{13}C labeling at C_s or C_6 position removes the molecular symmetry and hence there appear four equivalent IR bands in the spectral region thus considered. This observation explains why there appears an extra spectral band in the experimental spectra observed by Brudler, R. et al [1].

Keywords: Quinone, ubiquinone, symmetric mode, asymmetric mode, IR spectrum, isotope labeling, vibration, modes of vibration.

1. INTRODUCTION:

Infrared spectroscopy is a tool which deals with the molecular vibrations in which corresponding molecule changes its dipole moment [2, 3]. A molecule is chemically influenced in a material from the atoms or group of atoms around it. Its bonding strength depends on the type of bonding, charge on the molecule and electronic environment around the molecule. Studying vibrational frequency and intensity of spectral bands of the infrared spectra digs up state of the molecule and its environment [3]. However, due to the interaction with neighbors the composite spectrum of molecules in vivo is very complex. Isotope labeling is one of the methods of separating contribution of a particular group of vibration in the composite spectrum [1-5]. Frequency of vibration is generally decreases upon isotope labeling of some atom which takes part in the vibration when it is replaced with a heavier isotope. The decrease of frequency upon isotope labeling also changes the extent and type of coupling between two functional groups which are vibrating with nearly equal frequencies. It is of physical importance to study what happens to the infrared weak symmetric band upon asymmetric isotope labeling. Ubiquinone is one of the most studied and useful molecules in this field of research.

Ubiquinones (UQ: 2,3-dimethoxy-5-methyl-6polyprenyl-1,4-benzoquinone) which important role in biological electron and proton transfer processes that occur in both respiration and photosynthesis [6] have intense IR spectral bands in the frequency region between 1700 to 1550 cm⁻¹ in the neutral state in solvent and in the purple bacteria reaction centers[1, 4, 5, 7-15]. These IR bands are originated from the stretching vibration of C=O and C=C bonds of ubiquinone. Experimental spectra of ubiquinone in the solvent or Q_A binding site of purple bacteria were taken from ubiquinone having long isoprene tail (10 units) [1, 4]. However, calculated spectra were generated from the ubiquinone with single isoprene unit [10, 16] because of negligible contribution of C=C groups of the tail and also the computational economy. To explore the environmental effect on the ubiquinone in the purple bacteria reaction center, isotope edited IR double difference spectra were measured in vivo [1, 4]. All of the experiments conform three major neutral ubiquinone IR bands which are down shifted in the purple bacteria reaction center compared to the band positions in the solvents [1, 4, 7]. However, IR spectra of neutral ubiquinones which are sitespecific ¹³C labeled at the C₅ or C₆ positions (C₃ or C₂ positions in author's numbering) contains four distinct bands in CCl₄ [1].

It is claimed that the extra band at 1638 cm⁻¹ is due to the band shifting from the 1650 cm⁻¹ band on site-specific ¹³C labeled at the C_5 or C_6 positions. However, origin of the extra band at 1638 cm⁻¹ has not clearly been discussed yet [1]. We have observed that the new band at 1638 cm⁻¹ of site-specific ¹³C labeled neutral ubiquinone in CCl_4 is due to the infrared intensity redistribution among C=C and C=O bands up on asymmetric isotope labeling of the molecule. This finding may also be valuable to explore the origin of various bands of ubiquinone difference spectrum in the purple bacteria reaction centers [1, 4].

2. MATERIALS AND METHODS UQ structure and numbering

Figure 1 shows the structure and numbering scheme for UQ_n . The subscript, n, refers to the number of isoprene units in the chain at position 6. The ubiquinone molecule has two carbonyl groups at C_1 and C_4 positions, two methoxy groups at positions at C_2 and C_3 , a methyl group at C_5 position and an isoprene unit at C_6 position. In this work we have removed the part of the tail after the first isoprene unit.

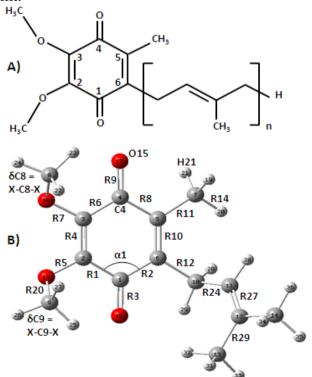


Figure 1 (A) Structure and numbering scheme for UQ_n . The subscript n refers to the number of isoprene units associated with the chain attached at position 6. (B) Energy minimized UQ_1 model used in calculations. Atom numbering scheme and brief representations of internal coordinates are also shown. R, α and δ represent for bond stretching, angle bending and combination of angle bending at a vertex atom respectively.

3. MOLECULAR OPTIMIZATION

Molecular geometry optimizations and harmonic vibrational frequency calculations of ubiquinone molecules were performed using hybrid DFT methods, employing the B3LYP functional and the 6-31+G(d) method within Gaussian 03 [17]. Minimum energy conformations of ubiquinone molecules were explored by calculating single point energy for 10° steps of 2- and 3- methoxy dihedral angles. The constrained minimum energy conformers were further optimized in dichloromethane using PCM method [18] without any constrained.

4. SPECTRAL CALCULATIONS OF UNLABELED AND SITE SPECIFIC ISOTOPE LABELED UBIQUINONE MOLECULE

Vibrational frequency calculations of optimized ubiquinone molecule were performed at DFT/B3LYP/6-31+G(d) level for both unlabeled molecule and ¹³C labeled molecule at C₅ or C₆ carbon position of ubiquinone molecule. Infrared spectra were generated from the Gaussian output files of frequency calculations using Gauss View 5.0 at 4 cm⁻¹ resolution. Potential energy distribution is calculated using gar2ped program [19].

5. RESULTS

Ubiquinone has two flexible methoxy groups and a long isoprene tail. Length and orientation of isoprene units does not affect on the vibrational frequency provided that the molecule contains at least one isoprene [10, 15]. However, orientations of methoxy groups have vital roles on the vibrational frequency, modes of vibration, and hence the shape of spectrum. Thus, to extract realistic absorption spectra from possible structures at room temperature, all possible minimum energy conformers of neutral ubiquinone molecules should be considered. The minimum

energy conformers of neutral ubiquinone molecules in CCl_4 have been explored by calculating single point energies at the step of 10° dihedral angles of the methoxy groups with respect to the ubiquinone head. Eight minimum energy conformers labeled A, B, C, D, E, F, G and H have been observed [10]. The Boltzmann weight factors [20] for conformers in the composite spectrum were calculated from the calculated energy difference of the neutral ubiquinone molecules at room temperature.

The calculated ground state energy, energy difference, Boltzmann factors and dihedral angles of methoxy groups of the conformers at room temperature are shown in Table 1. Table 1 shows that conformers A, B, C and D have relatively lesser energies than that of the remaining conformers E, F, G and H. Among conformers four low energy conformers C has the lowest energy at room temperature. Thus, the energy differences were calculated with respect to the energy of conformer C. Boltzmann factors were calculated at room temperature 298 K.

Table 1: Calculated optimized ground state energies, dihedral angles of methoxy groups and corresponding Boltzmann factors of 8 neutral UQ₁ conformers in CCl₄ At 298 K, $k_{\rm R}T \sim 0.592$ kcal/mol.

Ubiquinone Conformers	2-methoxy dihedralangle	2-methoxy dihedralangle	Energy (kcal/ Mole)	ΔE (kcal/ Mole)	Boltzmann Factors
A	-9.3	120.9	-530364.8585	0.0788	0.2082
В	10.7	-121.7	-530364.8912	0.0461	0.2201
С	-122.5	10.4	-530364.9373	0	0.2379
D	117.2	-5.2	-530364.8081	0.1292	0.1912
Е	118.4	123.3	-530363.887	1.0502	0.0404
F	-144.5	141.9	-530363.7736	1.1637	0.0333
G	-123.8	-122.7	-530364.0127	0.9245	0.0499
Н	139.4	-142.3	-530363.4424	1.4949	0.0190

Calculated vibrational frequencies, infrared intensities, and potential energy distribution (ped) of the C=O and C=C stretching modes of vibration of unlabeled, ¹³C₅ and ¹³C₆ isotope labeled neutral ubiquinone one (UQ₁) conformers in CCl₄ are given in Table 2. The tail C=C stretching mode is not considered here because this band has very weak IR intensity and also it is not coupled with other modes of vibrations. Corresponding composite spectra generated from weighing spectra of eight conformers with Boltzmann factors of unlabeled, ¹³C₅ and ¹³C₆ isotope labeled neutral UQ₁ in CCl₄ are shown in Figure 2. Figure 2 shows that the unlabeled neutral UQ₁ has three intense IR bands at 1676, 1650 and 1604 cm^{-1} . The 1676 cm^{-1} band is mainly ($\sim 80\%$ ped) due to C₁=O stretching vibration in conformers A and B and C₄=O stretching vibration in conformers C and D. The 1650 cm⁻¹ band is due to remaining C=O stretching modes of vibrations coupled with C=C stretching modes of vibration. The third band at 1604 cm⁻¹ is due to the asymmetric stretching modes of vibration coupled with C=O modes which give rise to 1650 cm⁻¹ band. The symmetric C=C stretching modes nearly at 1653 cm⁻¹ have very weak IR intensities and is not seen in the spectrum. Because of higher energies and lesser Boltzmann factors, conformers E, F, G and H have very small contribution to the composite spectrum and hence they are not considered in the discussions here.

Table 2. Calculated vibrational frequencies (cm⁻¹), infrared intensities (km/mole) in parenthesis and potential energy distribution (%) of unlabeled, ¹³C_c and ¹³C_c isotope labeled neutral ubiquinone one in CCl_c.

potentiai e	otential energy distribution (%) of unlabeled, ${}^{13}C_5$ and ${}^{13}C_6$ isotope labeled neutral ubiquinone one in CCl_4 . Calculated Ubiquinone Conformers and IR Bands in CCl_4					
Conf.				IV		
Com.	Unlabeled Ubiquinone					
A	1678(224)	1650(414)	1653(31)	1604(379)		
A	$C_1 = O(83\%)$	$C_4 = O(69\%), C_5 = C_6$ (7%)	$C_2 = C_3 (22\%), C_5 = C_6 (42\%)$	$C_2 = C_3 (41\%), - C_5 = C_6 (19\%), C_4 = O (7\%)$		
В	1679(220) C ₁ =O (80%)	1650(418) C ₄ =O (68%), C ₅ =C ₆ (8%)	$C_2 = C_3 (22\%), C_5 = C_6 (41\%)$	1604(377) C ₂ =C ₃ (41%), - C ₅ =C ₆ (19%), C ₄ =O (6%)		
С	1676(308) C ₄ =O (80%)	1649(316) C ₁ =O (51%), C ₅ =C ₆ (20%)	1654(69) C ₂ =C ₃ (21%), C ₅ =C ₆ (29%), - C ₁ =O (24%)	1604(392) C ₂ =C ₃ (42%), - C ₅ =C ₆ (18%)		
D	1675(309) C ₄ =O (80%)	$ \begin{array}{c} 1648(309) \\ C_1 = O (48\%), C_5 = C_6 \\ (21\%) \end{array} $	1654(74) C ₂ =C ₃ (22%), C ₅ =C ₆ (27%), - C ₁ =O (26%)	$ \begin{array}{c} 1605(400) \\ C_2 = C_3 (41\%), - C_5 = C_6 \\ (19\%) \end{array} $		
Е	1664(7) C ₁ =O (47%), C ₄ =O (31%)	1657(653) C ₁ =O (35%), - C ₄ =O (43%)	$ \begin{array}{c} 1651(65) \\ C_2 = C_3 (28\%), C_5 = C_6 \\ (36\%) \end{array} $	$ \begin{array}{c} 1607(211) \\ C_2 = C_3 (40\%), - C_5 = C_6 \\ (29\%) \end{array} $		
F	1661(14) C ₁ =O (36%), C ₄ =O (23%), C ₅ =C ₆ (12%)	1650(639) C ₁ =O (18%), - C ₄ =O (58%)	$ \begin{array}{c} 1653(111) \\ C_2 = C_3 (16\%), C_5 = C_6 \\ (33\%), - C_1 = O (28\%) \end{array} $	$ \begin{array}{c} 1607(251) \\ C_2 = C_3 (46\%), - C_5 = C_6 \\ (20\%) \end{array} $		
G	1664(7) C ₁ =O (46%), C ₄ =O (29%)	1656(654) C ₁ =O (35%), - C ₄ =O (44%)	$ \begin{array}{c} 1651(51) \\ C_2 = C_3 (26\%), C_5 = C_6 \\ (38\%), - C_4 = O (8\%) \end{array} $	$ \begin{array}{c} 1606(216) \\ C_2 = C_3 (41\%), - C_5 = C_6 \\ (27\%) \end{array} $		
Н	1660(7) $C_1 = O(28\%), C_4 = O(34\%), C_5 = C_6(10\%)$	1649(573) -C ₁ =O (20%), C ₄ =O (46%), - C ₅ =C ₆ (10%)	$ \begin{array}{c} 1652(177) \\ C_2 = C_3 (15\%), C_5 = C_6 \\ (27\%), - C_1 = O (28\%) \end{array} $	$ \begin{array}{c} 1602(252) \\ C_2 = C_3 (45\%), - C_5 = C_6 \\ (22\%) \end{array} $		
Calc.	1676	1650	-	1604		
Expt.	1666	1650	-	1611		
¹³ C labele	d at C ₅ position					
A	1678(226) C ₁ =O (81%)	1650(357) C ₄ =O (71%), -C ₂ =C ₃ (11%)	1635(200) C ₂ =C ₃ (28%),C ₅ =C ₆ (24%), C ₄ =O (11%)	1592(266) C ₂ =C ₃ (24%), - C ₅ =C ₆ (41%)		
В	1679(221) C ₁ =O (80%)	1651(365) C ₄ =O (71%), -C ₂ =C ₃ (11%)	1635(197) C ₂ =C ₃ (29%),C ₅ =C ₆ (24%), C ₄ =O (10%)	1592(266) C ₂ =C ₃ (24%), - C ₅ =C ₆ (41%)		
С	1675(315) C ₄ =O (82%)	1651(315) C ₁ =O (74%), -C ₂ =C ₃ (8%)	$ \begin{vmatrix} 1635(163) \\ C_2 = C_3 (30\%), C_5 = C_6 \\ (25\%) \end{vmatrix} $	$ \begin{array}{c c} 1591(294) \\ C_2 = C_3 (24\%), - C_5 = C_6 \\ (40\%) \end{array} $		

	1	T	T	1	
D	1675(315)	1651(296)	1635(187)	1591(295)	
	$C_4 = O(82\%)$	$C_1 = O(72\%), -C_2 = C_3$	$C_2 = C_3 (29\%), C_5 = C_6$	$C_2 = C_3 (23\%), - C_5 = C_6$	
		(10%)	$(24\%), C_1 = O(9\%)$	(41%)	
Е	1664(12)	1657(694)	1639(74)	1590(157)	
	$C_1 = O(54\%), C_4 = O$	$C_1 = O(29\%), - C_4 = O$	$C_2 = C_3 (49\%), C_5 = C_6$	$C_2 = C_3 (19\%), - C_5 = C_6$	
	(29%)	(54%)	(18%)	(50%)	
F	1660(25)	1650(726)	1638(66)	1591(201)	
	$C_1 = O(55\%), C_4 = O$	$C_1 = O(27\%), - C_4 = O$	$C_2 = C_3 (39\%), C_5 = C_6$	$C_2 = C_3 (27\%), - C_5 = C_6$	
	(26%)	(56%)	(26%)	(41%)	
G	1663(13)	1655(684)	1638(70)	1590(163)	
	$C_1 = O(54\%), C_4 = O$	$C_1 = O(29\%), - C_4 = O$	$C_2 = C_3 (47\%), C_5 = C_6$	$C_2 = C_3$ (21%), - $C_5 = C_6$	
	(28%)	(55%)	(19%)	(48%)	
Н	1659(4)	1650(740)	1636(67)	1589(199)	
	$C_1 = O(43\%), C_4 = O$	$C_1 = O(39\%), - C_4 = O$	$C_2 = C_3$ (41%), $C_5 = C_6$	$C_2 = C_3 (26\%), - C_5 = C_6$	
	(39%)	(44%)	(25%)	(44%)	
Calc.	1676	1651	1635	1592	
Expt.	1666	1650	1638	1596	
¹³ C labeled at C ₆ position					
A	1678(221)	1650(394)	1635(138)	1592(290)	
	$C_1 = O(81\%)$	$C_4 = O(75\%), -C_2 = C_3$	$C_2 = C_3$ (31%), $C_5 = C_6$	$C_2 = C_3 (25\%), - C_5 = C_6$	
	1	(7%)	(26%)	(39%)	
В	1679(215)	1650(402)	1637(135)	1592(289)	
	$C_1 = O(80\%)$	$C_4 = O(75\%), -C_2 = C_3$	$C_2 = C_3$ (31%), $C_5 = C_6$	$C_2 = C_3 (25\%), - C_5 = C_6$	
	•	(6%)	(26%)	(39%)	
С	1675(315)	1652(274)	1634(211)	1594(284)	
	$C_4 = O(81\%)$	$C_1 = O(70\%), -C_2 = C_3$	$C_2 = C_3$ (26%), $C_5 = C_6$	$C_2 = C_3$ (26%), - $C_5 = C_6$	
		(11%)	$(24\%), C_1 = O(12\%)$	(39%)	
D	1675(316)	1652(255)	1634(237)	1594(284)	
	$C_4 = O(81\%)$	$C_1 = O(67\%), -C_2 = C_3$	$C_2 = C_3$ (26%), $C_5 = C_6$	$C_2 = C_3 (25\%), - C_5 = C_6$	
		(13%)	$(23\%), C_1 = O(15\%)$	(40%)	
Е	1663(9)	1657(681)	1639(88)	1591(158)	
	$C_1 = O(51\%), C_4 = O$	$C_1 = O(32\%), - C_4 = O$	$C_2 = C_3 (47\%), C_5 = C_6$	$C_2 = C_3 (20\%), - C_5 = C_6$	
	(32%)	(49%)	(18%)	(49%)	
F	1660(19)	1651(708)	1638(85)	1591(204)	
	$C_1 = O(51\%), C_4 = O$	$C_1 = O(31\%), - C_4 = O$	$C_2 = C_3 (38\%), C_5 = C_6$	$C_2 = C_3 (28\%), - C_5 = C_6$	
	(30%)	(48%)	(26%)	(39%)	
G	1663(8)	1656(676)	1638(79)	1591(165)	
	$C_1 = O (51\%), C_4 = O$	$C_1 = O(32\%), - C_4 = O$	$C_2 = C_3 (46\%), C_5 = C_6$	$C_2 = C_3 (22\%), - C_5 = C_6$	
	(32%)	(49%)	(20%)	(47%)	
Н	1659(9)	1650(704)	1637(95)	1590(201)	
	$C_1 = O(38\%), C_4 = O$	$C_1 = O(44\%), - C_4 = O$	$C_2 = C_3 (39\%), C_5 = C_6$	$C_2 = C_3 (27\%), - C_5 = C_6$	
	(43%)	(36%)	(24%)	(41%)	

Calc.	1676	1651	1635	1592
Expt.	1666	1650	1638	1596

Note: The calculated frequencies are scaled by 0.973. Experimental values were taken from reference [1]. The minus sign means the out of phase vibration. (Calc. = Calculated, Expt. = Experimental)

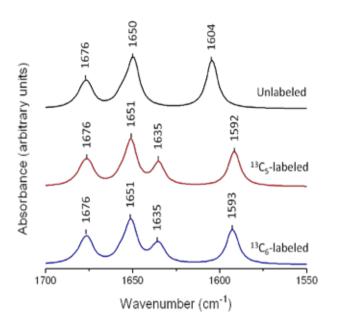


Figure 2. Calculated IR spectra of unlabeled, 13 C₅ and 13 C₆ isotope labeled neutral ubiquinone one in CCl₄. Each spectrum is the weighted average spectra of eight conformers (A, B,C, D, E, F, G and H) using Boltzmann factors 0.2082, 0.2201, 0.2379, 0.1912, 0.0404, 0.0333, 0.0499 and 0.0190 respectively.

Upon site specific asymmetric $^{13}C_5$ or $^{13}C_6$ labeling both C=C symmetric and asymmetric modes downshifted by ~ 18 and 14 cm⁻¹ respectively. The C=C symmetric mode also gained much IR intensity upon asymmetric 13 C labeling at C_5 or C_6 positions and as a result a new band appeared at 1635 cm⁻¹ in the corresponding IR spectrum. However, the C=C asymmetric band loses some intensity. On the other hand, the C=C coupled C=O mode at 1650 cm⁻¹ is up-shifted by ~1 cm⁻¹ and also lost some IR intensity.

6. DISCUSSIONS

Spectral modification of UQ_1 in CCl_4 upon site specific ¹³C labeling can be compared with the experiment carried out by R. Brudler et al. [1] on UQ_{10} in CCl_4 . Their numbering is different from ours in such a way that the atom numbers C_2 and C_3 are same as our atom numbers C_6 and C_5 . They also observed three intense IR bands of unlabeled

neutral UQ₁₀ in the spectral region between 1700 and 1600 cm⁻¹ in CCl₄ at 1666, 1650 and 1611 cm⁻¹ ¹. Upon site specific asymmetric ¹³C labeling at C₂ or C₂ positions (author's naming) they also observed four intense IR bands at 1666, 1650, 1638 and 1996 cm⁻¹. The unchanged values of the frequency and intensity of the highest frequency C=O stretching mode in the experiment carried by R. Brudler et al. is exactly revived in our calculations too. We have also observed decrease in IR intensity of 1650 cm⁻¹ band upon ¹³C labeling at C₅ or C₆ positions (our naming) as the experimenters observed. Our observed result of ~ 12 cm⁻¹ wavenumber downshift of C=C asymmetric band is comparable with the 15 cm⁻¹ downshift of C=C asymmetric band in the experiment of UQ₁₀ in CCl₄.

The origin of new band at 1638 cm⁻¹ upon asymmetric ¹³C labeling at C₂ or C₃ positions (their naming) was not clearly explained by the experimenters. However, they implied that this new band might be 12 cm⁻¹ wavenumber shifted from 1650 cm⁻¹ band. On the basis of the experimental data, they claimed that whatever the origin of the new band the symmetry of IR spectra of ¹³C labeled at C, or C₃ positions of UQ,0 showed that these two positions were chemically equivalent. We have clearly seen that the slight change in the IR intensity and frequency upon asymmetric ¹³C labeling is due to the modification of the scheme of coupling between lower frequency C=O stretching vibration and C=C stretching vibrations. The frequency downshift and intensity change of the asymmetric C=C stretching vibrations are also isotope mass increase and some changes in the coupling scheme between low frequency C=O made and C=C stretching modes of vibrations. Moreover, the asymmetric ¹³C labeling at C₂ or C₃ positions of neutral ubiquinone not only down-shifts the symmetric C=C stretching mode but also increases its IR intensity. The spectral modification observed in the calculations of vibrational frequencies of neutral

unlabeled and asymmetric 13 C labeled ubiquinone molecule may also be useful to explain experimental spectral modifications observed in the ubiquinone spectra in the Q_A binding site of the purple bacteria reaction centers.

7. CONCLUSIONS

Infrared band intensity of very weak symmetric mode of vibration of symmetric molecules can be enhanced by asymmetric isotope labeling of participating atoms. This technique will be useful to explore positions of silent symmetric IR modes of the relevant molecules.

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REFERENCES

- 1. Brudler, R., et al., Asymmetric binding of the 1- and 4-C=O groups of QA in Rhodobacter sphaeroides R26 reaction centres monitored by Fourier transform infra-red spectroscopy using site-specific isotopically labelled ubiquinone-10. The EMBO Journal, 1994. 13(23): p. 5523-5530.
- 2. Foresman, J. and A. Frisch, *Exploring Chemistry with Electronic Structure Methods*. 1996.
- 3. Stuart, B., Infrared Spectroscopy: Fundamentals and Applications. 2004.
- 4. Breton, J., et al., Binding Sites of Quinones in Photosynthetic Bacterial Reaction Centers Investigated by Light-Induced FTIR Difference Spectroscopy: Assignment of the Interactions of Each Carbonyl of QA in Rhodobacter sphaeroides Using Site-Specific 13C-Labeled Ubiquinone. Biochemistry, 1994. 33(48): p. 14378-14386.

- 5. Lamichhane, H., R. Wang, and G. Hastings, Comparison of calculated and experimental FTIR spectra of specifically labeled ubiquinones. Vibrational Spectroscopy, 2011. 55(2): p. 279-286.
- 6. Trumpower, B., Function of Quinones in Energy Conserving Systems. Academic Press, 1982.
- 7. Breton, J., et al., The Unusually Strong Hydrogen Bond between the Carbonyl of QA and His M219 in the Rhodobacter sphaeroides Reaction Center Is Not Essential for Efficient Electron Transfer from QA- to QB†. Biochemistry, 2007. **46**(22): p. 6468-6476.
- 8. Alexov, E.G. and M.R. Gunner, Calculated Protein and Proton Motions Coupled to Electron Transfer: Electron Transfer from QAto QB in Bacterial Photosynthetic Reaction Centers†. Biochemistry, 1999. 38(26): p. 8253-8270.
- 9. Mäntele, W., Reaction-induced infrared difference spectroscopy for the study of protein function and reaction mechanisms. Trends in Biochemical Sciences, 1993. **18**(6): p. 197-202.
- 10. Lamichhane, H.P. and G. Hastings, Calculated vibrational properties of pigments in protein binding sites. Proceedings of the National Academy of Sciences, 2011. **108**(26): p. 10526-10531.
- 11. Brudler, R., et al., FTIR spectroscopy shows weak symmetric hydrogen bonding of the QB carbonyl groups in Rhodobacter sphaeroides R26 reaction centres. FEBS Letters, 1995. 370(1-2): p. 88-92.
- 12. Balakrishnan, G., P. Mohandas, and S. Umpathy, *Ab initio studies on structure and vibrational spectra of ubiquinone and its radical anion.* Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy, 1997. **53**(10): p. 1553-1561.
- 13. Bandaranayake, K.M.P., R. Wang, and G. Hastings, Modification of the Phylloquinone in the A1 Binding Site in Photosystem I Studied Using Time-Resolved FTIR Difference

- *Spectroscopy and Density Functional Theory†*. Biochemistry, 2006. **45**(13): p. 4121-4127.
- 14. Nonella, M., et al., Structures and Vibrational Frequencies of the Quinones in Rb. sphaeroides Derived by a Combined Density Functional/ Molecular Mechanics Approach. The Journal of Physical Chemistry B, 2002. **107**(1): p. 316-322.
- 15. Nonella, M. and C. Brändli, Density Functional Investigation of Methoxy-Substituted p-Benzoquinones: Conformational Analysis and Harmonic Force Fields of 2-Methoxy-and 2,3-Dimethoxy-1,4-benzoquinone. The Journal of Physical Chemistry, 1996. 100(34): p. 14549-14559.
- 16. Boesch, S.E. and R.A. Wheeler, Structures and Properties of Ubiquinone-1 and Its

- Radical Anion from Hybrid Hartree–Fock/ Density Functional Studies1. The Journal of Physical Chemistry A, 1997. **101**(32): p. 5799-5804.
- 17. Frisch, M.J., et al., *Gaussian 03*, *Revision C.02*. Gaussian 03, Revision D.01, 2004.
- 18. Tomasi, J., B. Mennucci, and E. Cancès, *The IEF version of the PCM solvation method:* an overview of a new method addressed to study molecular solutes at the QM ab initio level. Journal of Molecular Structure: THEOCHEM, 1999. **464**(1-3): p. 211-226.
- 19. Martin, J.M.L. and A.C. Van, *GAR2PED*. 1995.
- 20. Patharia, R.K. and P.D. Beale, *Statistical Mechanics*. 1988.