ANTIFUNGAL EFFICACY OF ZANTHOXYLUM OIL AGAINST BIPOLARIS SOROKINIANA (SACC.) SHOEM.

Anshu Manandhar and R.D. Tiwari* *Central Department of Botany Tribhuvan University, Kirtipur, Kathmandu

ABSTRACT

Bipolaris sorokiniana (Sacc.) Shoem. is the most economically important foliar blight pathogen of wheat in all warm zones. Essential oil of *Zanthoxylum armatum* D.C. was used to determine its efficacy against *Bipolaris sorokiniana* by calculating the percentage of mycelial growth inhibition. The oil showed 25, 37.8, 42.8, 75, 84.5 and 100 percent inhibition of mycelial growth at 5.0, 10.0, 15.0, 20.0, 30.0, and 40.0 μ l ml⁻¹ ppm concentrations of essential oil against the test fungus, respectively.

Keywords: Essential oil, fungitoxicity.

Green plants have been found very successful in providing less toxic, more systemic (Fawcett and Spencer 1970), easily biodegradable (Beye 1998), host metabolism stimulatory fungicide (Dixit *et al.* 1983) as well as constituting a renewable source. Certain plants contain products such as alkaloids, tannins, quinines, coumarines, phenolic compounds, in their extracts and exudates, and they are known for their antifungal activities. The presence of antifungal compounds in plant tissues has long been recognized as an important factor for disease resistance (Fawcett and Spencer 1970.)

The average production of wheat in Nepal is very low than the potential yield. This is due to some major wheat diseases. Foliar blight complex (Spot blotch) is one of them. It is commonly known as *Helminthosporium* leaf blight. It is a major disease of wheat and is caused by *H. sativum* P.R. and B (syn. *B. sorokiniana* Sacc) and *Drechslera tritici repentis* Died. The disease is extended from Terai belt to 2600 masl. The incident and severity of blight are comparatively

ECOPRINT VOL 12, 2005

lower in the hills than in the Terai. Considerable loss in wheat grains has been found due to this disease which ranges from 3.1 to 29.0% in a normal year in natural epidemic condition depending upon wheat varities (Mahto 1999).

The main sources of innoculum of *B.* sorokiniana are infected seeds, infected crop residues, volunteer plants, secondary host and free dormant conidia in the soil (Reis 1991). Infected seeds are one of the most important ways for the survival of this pathogen between wheat cropping season. Seed infection by *B. sorokiniana* can adversely affect germination and root system development and can kill seedlings within a few days (Mehta 1993). In Nepal the average seed infection from Terai to hill region varies from 5.0 to 89.1% irrespective of cultivars (Shrestha *et al.* 1998).Recent studies in soil health revealed the presence of *B. sorokiniana* in the soil.

Bipolaris sorokiniana: Mycelium simple, hyaline, conidiophores brown, short, unbranched, aseptate and bear 1-6 conidia. Conidia thickwalled, ellipsoid, dark, olive brown, straight to slightly curved and having 5-9 cells and measures $6-12 \ \mu m \ x \ 12-20 \ \mu m$ (Zillinsky 1983). Colonies on PDA are olivaceous brown to black. In old cultures the number of conidia is so abundant that all cultures turn black and shiny.

Essential oil of Zanthoxylum armatum was obtained from HPPCL (Herb Production and Processing Co. Ltd.) Koteshwar, Kathmandu. Bipolaris sorokiniana was used as test fungus species and its culture was obtained from NARC (National Agriculture Research Council). The fungus was isolated from wheat seeds from Rampur by blotter test method. The pure culture was maintained on Potato Dextrose Agar medium. Seven days old culture of the test fungus was used to prepare inoculum discs (4 mm diameter). A single disc was aseptically placed upside down in the center of each petriplate containing essential oil loaded PDA, so as to establish a direct contact with the medium.

Acetone (80%) was used as the solvent to prepare different concentrations of essential oil (5.0, 10.0, 15.0, 20.0, 30.0 and 40.0 µl ml⁻¹ ppm) for use in experiment (Rao and Srivastava 1994) to assess fungitoxicity following poison food technique (Grover and Moore 1962). A volume of 0.5 ml of each concentration of essential oil was aseptically poured into the petriplate followed by the addition of melted PDA (9.5 ml). The petriplate was kept swirling while adding PDA so as to get a thorough mixing of the contents. Essential oil was, however, replaced by an equal amount of acetone in control set. After the media solidified one inoculum disc of B. sorokiniana mycelium, the test fungus was aseptically placed to each petriplate and incubated at 25+2° C. The average diameter of fungal colonies was measured on 7th day after inoculation and percentage of mycelial growth inhibition was calculated (Rao and Srivastava 1994). In the experiment, three

ECOPRINT VOL 12, 2005

replicates of the petridishes containing PDA were used and the colony size was calculated by taking an average of the three.

Mycelial Growth Inhibition (%) =
$$\frac{gc - gt}{gc} \times 100$$

Where, $g_c = Growth$ of mycelial colony after incubation period in control set subtracting the diameter of inoculum disc, $g_t = Growth$ of mycelial colony after incubation period in treatment set subtracting the diameter of inoculum disc.

Table 1. Assessment of antifungal activity of essential oil of Zanthoxylum armatum against Bipolaris sorokiniana.

SN	Concentration of oil (µl ml ⁻¹)	Mean colony size (mm)	Inhibition of mycelial growth (%)
1	0	64.0 ± 5	-
2	5.0	49.0 ± 4	25.0
3	10.0	41.3 ± 1.53	37.8
4	15.0	38.3 ± 1.53	42.8
5	20.0	19 ± 0.58	75.0
6	30.0	13.3 ± 3.21	84.5
7	40.0	4.0	100

The results showed the mycelial growth inhibition percentage over control. The mycelial growth inhibition percentage was 25.0, 31.26, 36.16, 75.0, 84.5 and 100 percent at essential oil concentrations of 5.0, 10.0, 15.0, 20.0, 30.0 and 40.0 μ l ml⁻¹ ppm, respectively.

During recent years, the essential oils of a number of higher plants have been tested for their *in vitro* antifungal activities, however, the detailed fungitoxic evaluation of such oils in order to prove their usefulness as successful fungitoxicants is limited (Tripathi *et al.* 1983). The toxicity in plants is believed to be due to the presence of certain fungitoxic factors in their tissue (Fawcett and Spencer 1970). The essential oil of *Zathoxylum armatum* possesses such chemical components which on treatment against *Bipolaris sorokiniana* exhibited absolute toxicity by complete inhibition of the growth of the test fungus.

The authors are highly obliged to Dr. N. N. Tiwari, General Manager, HPPCL for providing the essential oil of Zanthoxylum armatum. We would like to acknowledge with deep appreciation to Sarala Sharma, Sarada Joshi and Gyanu Manandhar for their generous gift of the test fungus. The authors are also thankful to Central Department of Botany, Tribhuvan University, Kirtipur for providing the laboratory facilities to carryout this work.

REFERENCES

- Beye, F. 1998. Insecticide from vegetable kingdom. *Plant Res. Devel.* **7:**13-31.
- Bhatta, M.R. 2003. An overview of wheat research and development (2001-2002). In: *Proceedings of Wheat Research Papers*, presented at 25th National Winter Crops Research Workshop Wheat Coordinators Report. *Wheat*. 1:1-18.
- Dixit, S.N., N.N. Tripati and S.C. Tripathi. 1983. Fungitoxicity of some seed extracts. *Nat. Acad. Sci. Letters* 1:287-288.
- Fawcett, C.H. and D.M. Spencer. 1970. Plant chemotherapy with natural products. *Ann. Rev. Phytopath* 8:403-418.
- Grover, R.K. and J.D. Moore. 1962. Toximetric studies of fungicides against brown rot organism. *Sclerotinia fruticsla. Phytopathology* **52:**876-880.
- Mahto, B.N.1999. Loss assessment due to foliar blight of wheat in Nepal. Ann. Agric. Res. 20(2):212-215.

- Mehta, Y.R. 1993. Spot blotch (Bipolaris sorokiniana). In: Seed Borne Diseases and Seed Health Testing of Wheat. (eds.) Mathur, S.B. and B.M. Cunfer. Institute of Seed Pathology for Developing Countries, Copenhagen, Denmark, Pp. 105-112.
- Rao, G.P. and A.K. Srivastava. 1994. Toxicity of essential oils of highter plant against fungal pathogens of sugarcane. *Current Trends in Sugarcane Pathology*. (eds.) Rao, G.P., A.G. Gillasple, P.P. Upadhyaya, A. bergamin, V.P. Agnihotri and C.T. Chen. International Books and Periodicals Supply Service, Pitampura, Delhi, pp. 347-365.
- Reis, E.M.1991. Integrated disease management the changing concepts of controlling head blight and spot blotch. (ed.) D.A. Saunders. *Wheat for the Non-Traditional, Warm Areas*,CIMMYT, Mexico, D.F. pp. 165-177
- Shrestha, K.K., R.D. Timila, B.N. Mahto and H.P.
 Bimb. 1998. Disease incidence and yield loss due to foliar blight of wheat in Nepal. In: *Helminthosporium Blights of Wheat, Spot Blotch and Tan Spot.* (eds.) Duveiller, E.,
 H.J. Dubin, J. Reeves, and A. McNab.
 CIMMYT. Mexico, D.F. pp. 67-72.
- Tripathi, N.N., N.K. Dubey, A. Dikshit, R.D. Tripathi and S.N. Dixit. 1983. Fungitoxic properties of *Alpinia galanga* oil. *Trop. Plant. Sci. Res.* 1(1):49-52.
- Zillinsky, F. 1983. Common Diseases of Small Grain Cereals: A Guide to Identification. CIMMYT. Mexico, D.F. 141 pp.

ECOPRINT VOL 12, 2005